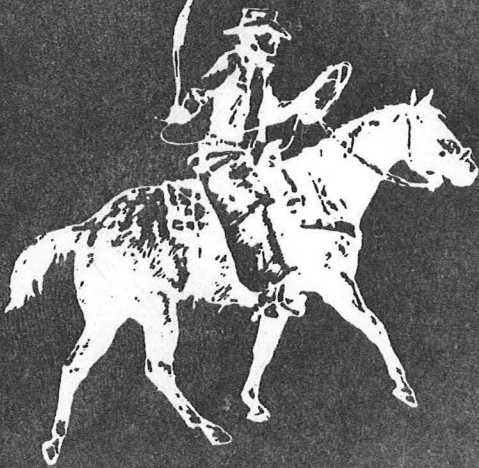


SOMATOSTATINOMA
SYNDROME



INTERNAL MEDICINE GRAND ROUNDS - PARKLAND MEMORIAL HOSPITAL
APRIL 26, 1979 GUENTER J. KREJS, M.D.

PROLOGUE

THE FOUR MODES OF
FUNCTION SUGGESTED
FOR SOMATOSTATIN:

1. NEUROHORMONE REGULATING
PITUITARY ENDOCRINE SECRETION
2. NEUROTRANSMITTER IN CENTRAL
AND PERIPHERAL NERVOUS SYST.
3. PARAHORMONE ACTING LOCALLY
IN THE GI-TRACT AND PANC. ISLETS
4. CONVENTIONAL HORMONE
REACHING TARGET ORGAN VIA
BLOOD CIRCULATION

APRIL 26 , 1979

CASE REPORT

E.L. is a 52 year old Canadian railroad inspector, who in 1966 and 1967 experienced episodes of melena which were attributed to peptic ulcer disease, although no ulcer was found by x-ray and endoscopic examination. In 1970 he had hematemesis and melena requiring blood transfusions, and again no ulcer was found. In 1973 he was troubled by postprandial eructations and abdominal pain, as well as periumbilical cramping after heavy meals. In 1974 a large single gallstone was shown on oral cholecystogram and, on barium meal examination, a filling defect was detected in the second portion of the duodenum (Fig 1).



Figure 1: Duodenal C-loop with filling defect (3 x 4cm) in the second portion (film dated January 24, 1974)

At endoscopy this filling defect was described as broad-based sessile polyp located just proximal to a normal appearing papilla of Vater. Subsequently in 1974, a cholecystectomy (large single gallstone), truncal vagotomy and pyloroplasty were performed, but no ulcer was found. During the pyloroplasty, a firm lobulated tumor mass, measuring about 3 x 4 cm, was palpated digitally in the area of the ampulla.

It was not fixed and appeared to be in the wall of the medial portion of the second part of the duodenum. It was not possible to biopsy it or see it through the pylorotomy; the pyloroplasty was therefore completed and the duodenum and head of the pancreas mobilized by Kocher maneuver and the tumor inspected through a duodenotomy in the second part of the duodenum. No mucosal lesion was noted. A Vim-Silverman needle biopsy was performed through the duodenal wall, but the biopsy did not survive processing. A subsequent barium meal confirmed the filling defect on the concavity of the duodenal loop above the level of the ampulla. This was unchanged on another barium meal in 1976.

The patient was relatively well until 1977, when he returned complaining of a dry mouth and belching. Another barium meal study showed no change of the duodenal filling defect. Late in 1977, he developed weight loss (84 to 75 kg, height 178 cm) associated with occasional vomiting two hours after meals. He also complained of abdominal discomfort relieved by eructation and loose stools. One of three stools was positive for occult blood. Sudan stain was positive for fat globules on a stool smear. Blood sugar was 246 mg/dl (3 hours after a meal), and a subsequent glucose tolerance test showed a fasting blood sugar of 180 and one and two hour values of 324 and 303 mg/dl, respectively, with 4% glucosuria. Except for an alkaline phosphatase at the upper limit of normal, blood chemistry tests were normal.

The patient's weight fell further to 73 kg. Repeat endoscopy showed duodenal mucosa stretched over what appeared as an extrinsic lesion, distorting the medial wall above the ampulla. Biopsies at that time revealed normal duodenal mucosa and cytology brushings were negative for tumor cells. The patient was begun on 14 units of Lente insulin per day and a 2,000 calorie diabetic diet, but he continued to note fatigue, together with periodic vomiting and diarrhea, consisting of 3-4 loose, foul smelling stools daily. Endoscopic retrograde cholangiopancreatography was performed and showed a normal pancreatic duct. Biopsies of the tumor at ERCP showed tissue consistent with an endocrine lesion such as gastrinoma or a carcinoid. The duodenal aspirate following secretin injection (1 clinical unit/kg) revealed a normal volume (4.1 ml/kg/80 min, normal >2) and bicarbonate output (0.27 meq/kg/80 min, normal >0.2). Serum gastrin was 99 pg/ml (normal <200), and 5-HIAA was not elevated in a 24-hour urine collection. A 72-hour stool collection yielded 20 g of fat per day on a 100 g fat diet. Pancreatic enzyme replacement was started, and stool frequency declined to one per day, but weight loss continued to 68 kg.

The clinical picture of combined exocrine and endocrine pancreatic insufficiency (steatorrhea and diabetes mellitus), and cholelithiasis in the presence of a duodenal tumor of endocrine type, was felt to be suggestive of a somatostatinoma. This was confirmed by somatostatin radioimmunoassay, which showed a plasma level in excess of 2,000 pg/ml (fasting level 88 ± 8 pg/ml in 10 normal controls) (see below). Subsequently the patient was admitted to the General Clinical Research Center of The University of Texas Health Science Center at Dallas for further workup (August 1978).

Physical exam was essentially negative on admission. The patient's weight had dropped to 64 kg. Abnormal laboratory results included hemoglobin 12.9 g/dl, hematocrit 36%, total bilirubin 1.3 mg/dl, and alkaline phosphatase 136 U per liter (30-115).

Endoscopy revealed a patent pyloric channel (pyloroplasty) and a 3 x 4 cm intramural mass located on the medial side of the second portion of the duodenum. The overlying mucosa was of normal appearance, and multiple biopsies showed normal duodenal mucosa. Vigorous duodenal motility was also noted during endoscopy.

Ultrasonography revealed a 4-5 cm mass in the region of the head of the pancreas. Liver spleen scan showed multiple filling defects in the liver consistent with metastases. Angiography revealed a hypervascular mass in the region of the head of the pancreas (2 x 3 cm) and multiple areas of hypervascular stain in the late arterial phase of hepatic artery injections, compatible with liver metastases. At peritoneoscopy, multiple white implants with hyperemic margins slightly raised above the liver surface were seen. Light microscopy of biopsies taken from the hepatic implants at peritoneoscopy showed tumor formed by small masses and cords of cells separated by a fibrovascular stroma. The tumor cells were cuboidal in shape, though some assumed columnar appearance. The tumor nuclei appeared to occupy more than half of the cell volume. No mitotic figures were identified. Secretory granules were seen in the tumor cells on electronmicroscopy performed at Parkland Hospital.

After completion of most of the special studies that I will subsequently discuss the patient was referred to the National Institutes of Health in Bethesda, Maryland (September 1978) for treatment. After several weeks of intravenous hyperalimentation and further investigation the patient was taken to surgery. A preoperative repeat angiogram suggested a rapid increase in size of the liver metastases and this was also reflected by the plasma somatostatin level which had doubled, (from around 10,000 to 20,000 pg/ml) during this period. A 5 x 5 cm tumor in the head of the pancreas was resected by a Whipple's procedure (follow up see page 22).

SOMATOSTATIN

Somatostatin is a cyclic polypeptide consisting of 14 amino acids (tetradecapeptide, molecular weight 1818).

The original observation that growth hormone secretion is under dual control of both stimulating and inhibiting factors was made in this institution by Krulich and associates in 1968 (7-9). At that time it had been proven that the hypothalamus controls and regulates the secretion of two pituitary hormones namely thyrotropin (TSH) and the gonadotropin, luteinizing hormone (LH). A hypothalamic releasing factor for growth hormone (somatotropin) was postulated on the basis of a series of experimental and clinical observations. The nature of this postulated hypothalamic releasing factor for growth hormone, however, had remained elusive. Searching to demonstrate the presence of this still hypothetical somatotropin releasing factor Krulich et al (7-9) incubated anterior pituitaries in vitro with crude sheep hypothalamic extract and measured growth hormone release into the medium. When crude extract was fractionated on Sephadex G-25, a consistent picture was obtained with a zone containing GH-releasing activity and a zone that produced a strong inhibition in GH release.

In 1972, Guillemin* and coworkers (10,11) succeeded to isolate the active principle using hypothalami from 500,000 sheep. The hypothalamic tetradecapeptide was called somatostatin in the belief that it was a specific hypothalamic factor which modulates the release of growth hormone (somatotropin) analogous to the factor which inhibits release of prolactin. The purity and yield of their isolation procedure permitted Guillemin and his colleagues to determine the amino acid sequence of somatostatin (12). This breakthrough then led to the production of a biologically active synthetic replica which in turn allowed the development of a radioimmunoassay (13,14) and a host of pharmacologic and physiologic studies.

Routine protein synthesis techniques are used to produce somatostatin which is now available to a great number of investigators. It is of interest that another method

* Nobel Prize 1977

for the synthesis of somatostatin has been brought about by the advances in molecular genetics. Synthesis of somatostatin can be performed by *E. coli* bacteria whose genetic code was changed (15).

Besides the hypothalamus, somatostatin has been found in numerous other areas of the central nervous system as well as in peripheral autonomic ganglia and plexus. The numerous examples of neurones containing somatostatin thus satisfy one of the criteria for a neurotransmitter or neuromodulator (6).

During such localization studies of somatostatin in central nervous tissue Dubois and others (17,18) used liver, spleen, pancreas and intestine as presumably negative control tissues. However, somatostatin was either found in high concentration in tissue extracts or identified by immunocytochemical studies in the digestive tract, particularly in the pancreas.

Figure 2 shows the somatostatin content of the digestive tract.

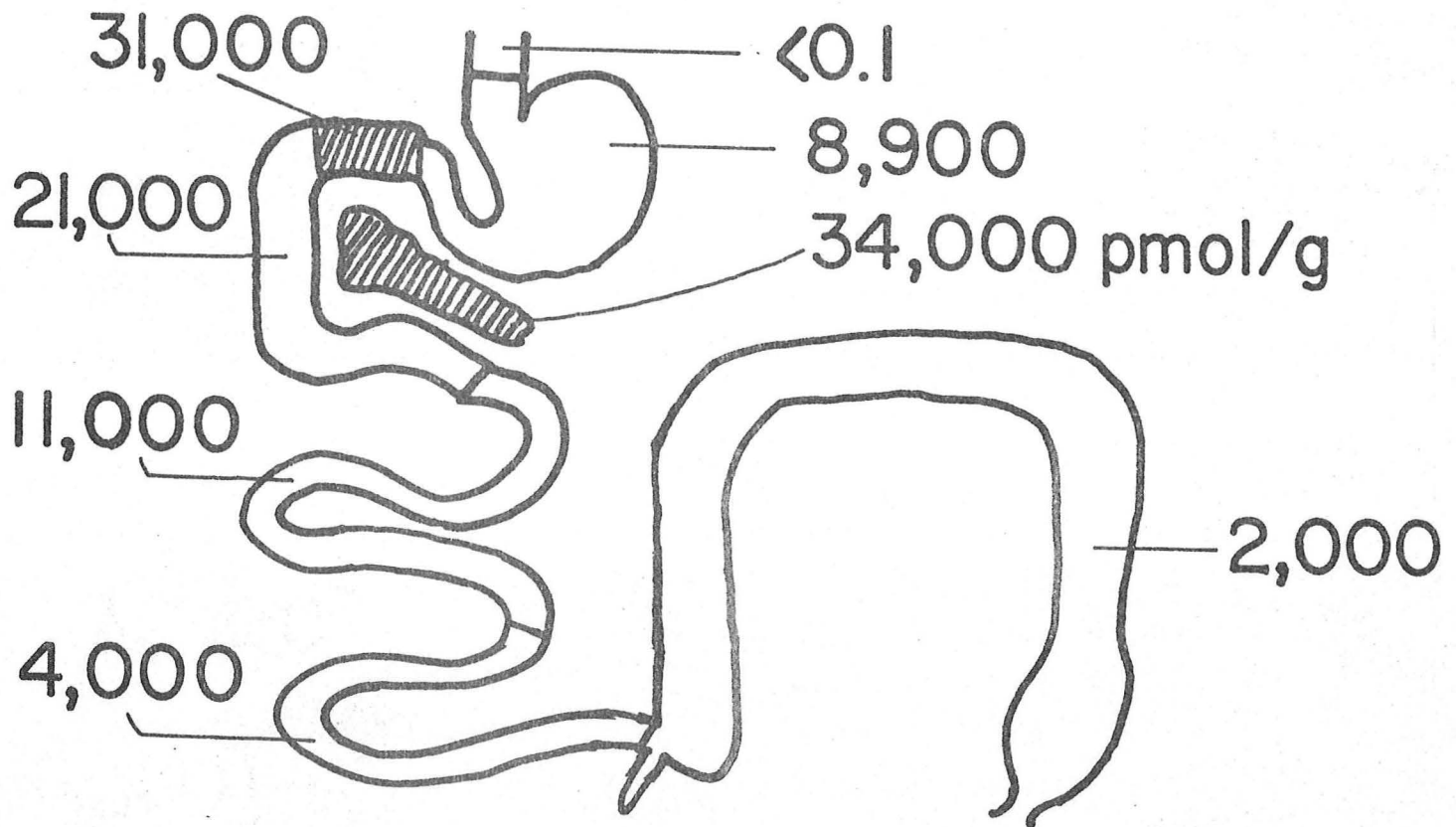


Figure 2: Somatostatin content (pmol/g tissue) of the digestive tract (modified from Bloom, S.R., 1978, *GASTROINTESTINAL HORMONES*)

In the pancreas somatostatin was localized to the D cells (17-23). In the normal islets of Langerhans of mammals, B cells are mainly located in the center of the islet, and a so-called "mantle" of A and D cells is seen on the islet periphery (23-25). This type of islet of Langerhans is observed in the body and tail of the pancreas. In the head and uncinata process, the islets also contain many pancreatic polypeptide-producing cells.

Ultrastructurally the D cells have been characterized by their large (200-400 nm) weakly osmiophilic secretion granules. The core of the secretion granule exhibits a fine homogeneous granulation and its membrane is closely applied.

PROOF OF DIAGNOSIS IN E.L.

The diagnosis of somatostatinoma in patient E.L. was proven by 1) somatostatin radioimmunoassay, and 2) immunocytochemical studies in tumor tissue obtained at peritoneoscopy (liver metastases) and surgery (primary tumor).

1. Radioimmunoassay (1,14)

The patient's fasting plasma level of SLI ranged from 9,000 to 13,000 and averaged $9,800 \pm 600$ pg/ml (mean \pm SEM)(July, August 1978).

For comparison the fasting SLI plasma level of healthy subjects (n=10) was found to be 88 ± 8 pg/ml (mean \pm SEM) and after a 1,000 calorie hamburger meal, SLI plasma levels rise to 135 ± 26 pg/ml (P. Raskin, unpublished observation).

When we infused somatostatin in (probably pharmacological) doses ($8 \mu\text{g}/\text{kg}/\text{h}$) in healthy subjects (26) plasma SLI levels rose to $2,062 \pm 192$ pg/ml (n=20), which is about one-fifth of the patient's fasting levels.

Thus, SLI in the patient was over 100 times the normal average. Extreme and constant hormonal excess is consistent with a malignant functioning islet cell tumor with metastatic spread.

To characterize plasma SLI, gel filtration chromatography of plasma samples was carried out on Biogel P-10 and Sephadex G-200 columns.

The SLI of canine plasma is undistinguishable from synthetic somatostatin, but it circulates bound to high molecular weight components ($150,000$) that are saturable by endogenous somatostatin (27). Although SLI in human plasma has been less thoroughly characterized, a typical elution profile on Biogel P-10 columns (pH 7.4) of plasma from a normal subject (Figure 3, upper panel) indicates that bound SLI also represents the predominant form in human plasma. If free somatostatin is present, it is below the detection limit of the radioimmunoassay (3 pg/tube). When the plasma samples were chromatographed under a dissociating condition, e.g. in 2 M acetic acid, however, over 90% of the total SLI was eluted from the column in the same position as synthetic somatostatin, thus confirming the earlier report by Kronheim et al (28). The lower panel on Figure 3 shows the elution profile of a basal plasma sample from the patient. In addition to a peak of somatostatin at the void volume of the column, multiple peaks of SLI were observed in the molecular weight regions 10,000-15,000, 2,000-3,500 and in the molecular weight region of free somatostatin. Chromatography of a plasma sample on Sephadex G-200 indicated that the amount of SLI eluted in the $>150,000$ molecular weight zone was not greater than in plasma of normal subjects.

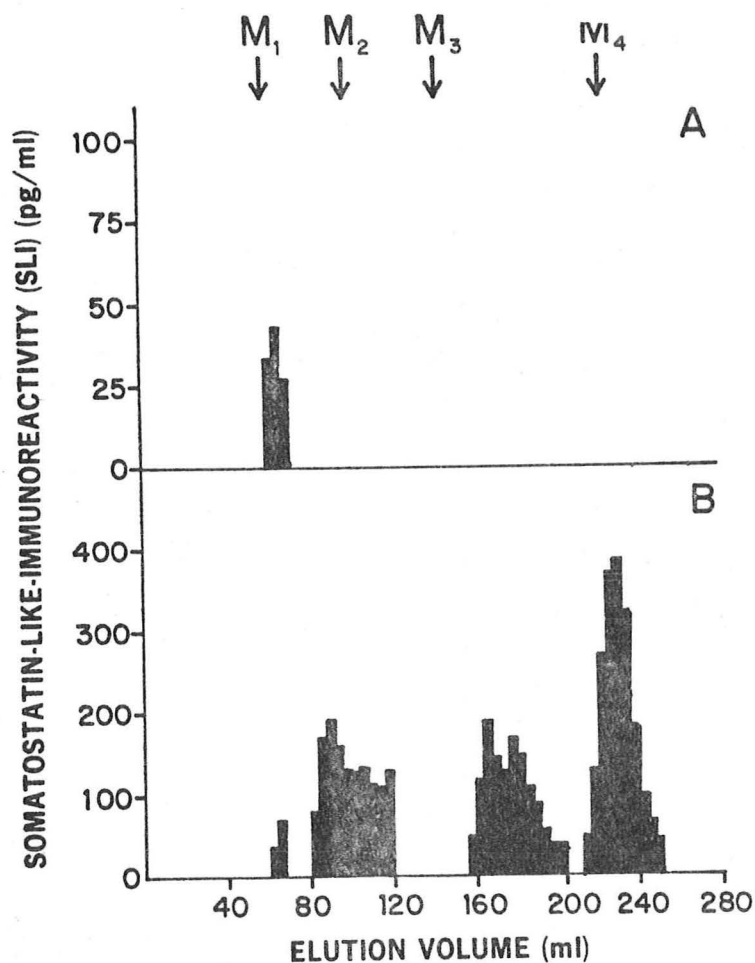


Figure 3: Elution profiles of SLI from the plasma of a healthy control subject (A), 4 ml plasma, 100 pg/ml SLI, and the patient with somatostatinoma (B), 3.0 ml plasma, >2,000 pg/ml SLI, on Biogel P-10 column at pH 7.4. The column was calibrated with markers of known molecular weight: M₁ (blue dextran; void volume), M₂ (cytochrome-c), M₃ (glucagon), and M₄ (somatostatin).

Thus, the SLI in our patient's plasma was distributed in four main peaks on gel filtration chromatography: these were a 1600-dalton peak, corresponding to synthetic somatostatin, a 3,000-dalton and 12,000-dalton peak corresponding to compounds that may represent precursors of somatostatin and correspond to peaks present in extracts of normal somatostatin containing tissues (29-42) but not previously found in plasma, and fourthly, a >150,000-dalton peak, corresponding to the fraction in which all measurable SLI appears when plasma of normal subjects is chromatographed by these techniques. This component is believed to represent somatostatin bound to large molecular weight plasma components (27). The latter was not increased above normal in this somatostatinoma patient.

Extracts of the primary tumor contained $\approx 5 \mu\text{g}/\text{mg}$ net tissue weight. The somatostatin content of normal pancreas has been reported to be 253 ± 43 (range 31-640)

pg/mg tissue (43). SLI was distributed into the same 1,600, 3,000 and 12,000 molecular weight fractions present in the plasma of this patient and in extracts of normal dog pancreas (41). Multiple molecular forms of SLI were also reported in extracts of the tumor of the patient described by Larsson et al (2). Analogous to insulinomas, glucagonomas and other polypeptide releasing pancreatic tumors (44-46), the recognition of such large molecular weight forms of SLI may aid in the diagnosis of pancreatic endocrine neoplasms.

2. Morphology

Liver metastasis: On electron microscopy, the cytoplasm of the tumor cells, obtained by biopsy of a liver metastasis at peritoneoscopy, was filled with small electron dense secretory granules of various shapes. Immunofluorescent staining of the biopsy material with several different antisera (see Table 2) revealed a positive reaction only with antisomatostatin antiserum. All control tests were negative, confirming the specificity of the immunofluorescent reaction.

Primary pancreatic tumor:

Light microscopy. The primary pancreatic tumor was composed of solid nests and cords of cells forming anastomosed ribbons or follicular structures. The latter were surrounded by cuboidal or columnar cells and filled with PAS positive material. Broad and thin bands of connective tissue penetrated the tumor tissue. No amyloid was detected.

Immunofluorescence. The results of immunofluorescence study are summarized in Table 2. Virtually all tumor cells displayed a positive reaction with the antisomatostatin antiserum. The immunofluorescence appeared consistently brighter at the periphery of the blocks of tumor tissue, whereas the central part displayed a weaker staining (Fig 4a,b). Exposure to antigen-absorbed antiserum completely abolished the immunoreaction (Fig 4). Calcitonin positive cells showing bright to weak fluorescence were also found, being much more numerous in one of the two blocks examined. Control tests showed a total inhibition of the specific antisomatostatin and anticalcitonin staining.

Electron microscopy. At the ultrastructural level the tumor was characterized by two populations of well granulated cells (Fig 5). The granules of one type were large and contained material that varied from dense and homogeneous to pale and flocculent. The other type of tumor cells showed polymorphic dense granules of small size, like the tumor cells of the liver metastasis. Junctional complexes were usually seen joining adjacent cells of the follicle-like structures. The apical parts of these cells were characterized by a well developed cell web and numerous microvilli.

Immunocytochemistry. With the antisomatostatin antiserum, using the two steps method, gold particles indicating antigen-antibody reaction sites, were consistently found on the secretory granules of all cells, irrespective of the granule size, shape and electron density (Fig 5). By contrast, with the anticalcitonin antiserum, gold particles were localized only within the population of small granule cells. Each antiserum staining was abolished by the homologous but not by the heterologous antigen.

Somatostatin and calcitonin immunoreactive material thus coexisted in the secretory granules of a minor population of tumor cells. However, since not all granules were labeled and consecutive sections were not examined, it cannot be decided whether somatostatin and calcitonin immunoreactivity are stored in the same secretory granules.

TABLE 2. Immunofluorescence Microscopy of Tumor Tissue

ANTISERUM	DILUTION	PRIMARY TUMOR	METASTASIS
Anti-somatostatin ^a	1:200	+	+
Anti-calcitonin ^b	1:300	+	-
Anti-insulin ^c	1:200	-	-
Anti-glucagon ^d	1:100	-	-
Anti-pancreatic polypeptide ^e	1:100	-	-
Anti-glicentin ^f	1:50	-	-
Anti-secretin ^g	1:10	-	-
Anti-gastrin ^h	1:50	-	-
Anti-ACTH ⁱ	1:10	-	-
Anti-TRH ^j	1:50	-	-

^aprovided by Dr. S. Ito, Niigata, Japan

^bprovided by Dr. J. McIntyre, London, England and directed against residues 16-19, 29-32 of calcitonin M

^cprovided by Dr. P.H.Wright, Indianapolis, Indiana, USA

^dprovided by Dr. R. Donald, Christchurch, New Zealand

^eprovided by Dr. R. Chance, Indianapolis, Indiana, USA

^fprovided by Dr. A.J. Moody, Bagsvaerd, Denmark

^gprovided by Dr. N. Yanaihara, Shizuoka-Shi, Japan

^hprovided by Dr. W. Gepts, Brussels, Belgium

ⁱprovided by Dr. J.A. Edwardson, London, England and directed against residues 17-39 of human ACTH

^jprovided by Dr. J.P. Leppäluoto, Oulu, Finland

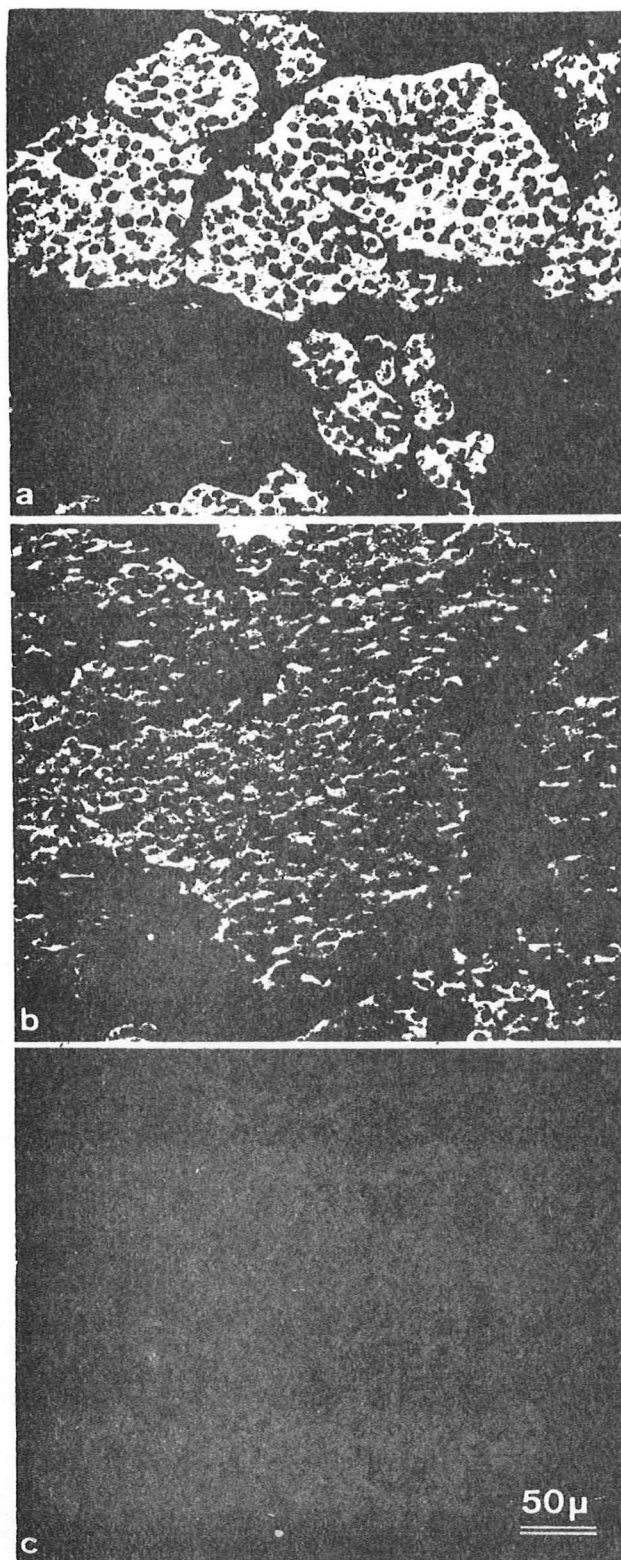


Figure 4: Paraffin sections of the primary pancreatic tumor stained with the indirect immunofluorescence method with anti-somatostatin serum. a. Clusters of brightly positive cells surrounded by abundant connective tissue. b. Masses of tumor cells showing a weak immunofluorescent reaction. c. Somatostatin immunostaining appears completely abolished on the section consecutive to b. which has been stained with an anti-somatostatin serum absorbed with synthetic somatostatin (X 200).

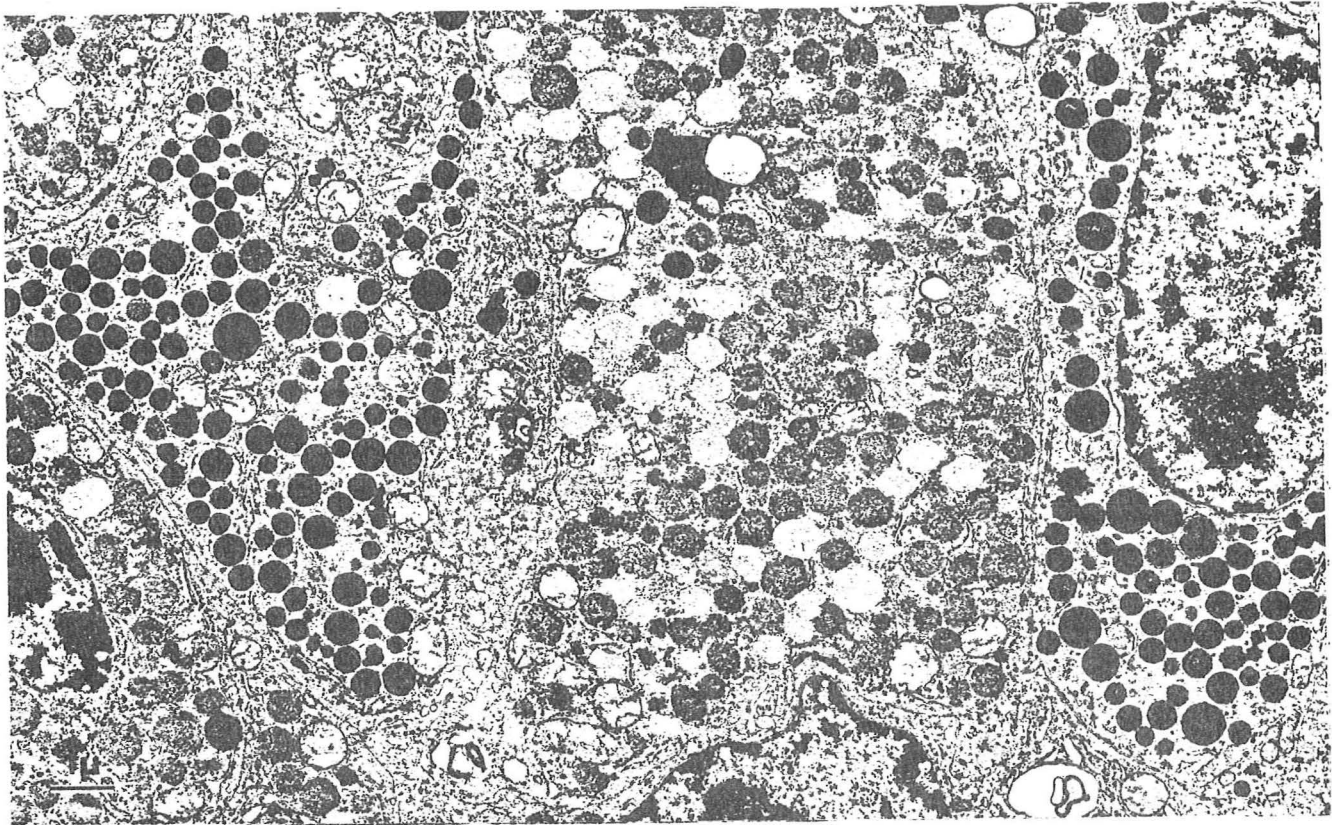


Figure 5: Low power electron micrograph showing several tumor cells. Note the large size and the variable degree of electron density of the secretory granules (X 8,000).

PLASMA CALCITONIN IN PATIENT E.L.

As may be expected from the morphological studies, plasma calcitonin was elevated in the patient's fasting plasma: 4,650 pg/ml (normal <120). High calcitonin levels were also observed in the patient reported by Galmiche et al (6) and we have recently seen a patient with a VIP-producing malignant islet cell tumor who also had a high plasma calcitonin concentration (47). Elevated serum calcitonin may be a marker of a variety of endocrine tumors.

ACTIONS OF SOMATOSTATIN

A. ENDOCRINE FUNCTIONS

1. Hypothalamic and Pituitary Function

In pharmacological doses somatostatin is a powerful inhibitor of human growth hormone secretion, suppressing the response to insulin-induced hypoglycemia (48),

L-Dopa (49), arginine infusion (50), exercise (51) and sleep (52).

In our patient, both insulin-induced hypoglycemia (blood sugar 30 mg/dl) and arginine infusion failed to stimulate growth hormone secretion with the plasma concentration remaining below 0.8 ng/ml.

Although somatostatin is known to suppress TRH-stimulated TSH secretion in euthyroid subjects (48,50,53) we were unable to demonstrate suppression of the TSH response to TRH in our patient. The reason for this is not clear but it is possible that the suppression of TSH secretion by somatostatin in acute experiments is not seen in the presence of persistently elevated levels of somatostatin.

Plasma levels of prolactin, FSH, LH and cortisol were normal in our patient which is in agreement with reports that somatostatin has no effect on these hormones (48, 49,53).

2. Pancreatic Islet Cell Function

One of the three cardinal symptoms in our patient was diabetes mellitus.

Somatostatin lowers basal and stimulated serum concentrations of insulin (54), glucagon (54) and pancreatic polypeptide (55). The combined suppression of insulin and glucagon release should give rise to a syndrome of mild diabetes with impaired glucose tolerance, but, because of concomitant suppression of glucagon, neither severe endogenous hyperglycemia nor hyperketonemia are to be expected (56).

In our patient islet cell function was investigated by oral glucose tolerance test, arginine infusion and the response to a mixed meal (1,000 calorie hamburger meal). Basal levels of glucagon (IRG) and C-peptide* in all three tests were normal.

During the glucose tolerance test (Figure 6) there was no suppression of IRG and the rise in C-peptide appeared to be normal. Basal SLI levels rose fourfold by 120 minutes after glucose ingestion.

During the arginine infusion test IRG rose more than fourfold and there was a 50 mg/dl rise in blood glucose (Fig 7). C-peptide levels also rose: however, the normal C-peptide response to arginine infusion has not been reported. SLI levels rose more than fourfold in response to arginine infusion.

The excessive rise in glucagon levels in response to arginine stimulation simulated the A-cell response of ordinary diabetes. Thus, despite the enormously increased levels of somatostatin, the A-cells were resistant to it.

After the mixed meal (Fig 8) there was a normal rise in IRG and although somewhat delayed, C-peptide rose in a normal fashion. SLI showed a dramatic (eightfold) increase in response to the meal.

The C-peptide response in these tests suggests that B-cell function was only moderately impaired. In fact, while in our General Clinical Research Center the patient's diabetes could be satisfactorily controlled with carbohydrate restriction and without insulin injection.

*Direct measurement of immunoreactive insulin was impossible in this patient due to insulin antibodies presumably resulting from insulin therapy. For the same reason it was impossible to perform the pancreatic polypeptide radioimmunoassay.

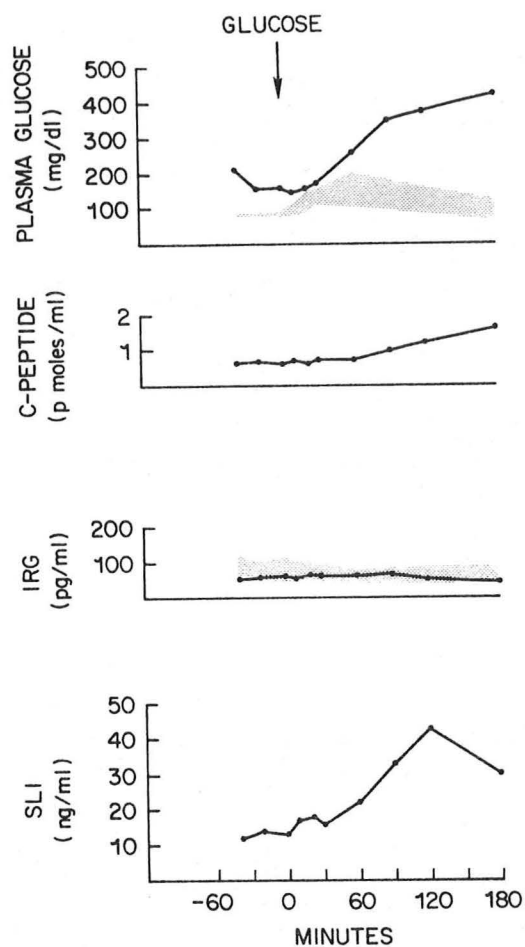


Figure 6: Plasma glucose, C-peptide, immunoreactive glucagon (IRG) and somatostatin-like immunoreactivity (SLI) response to an oral glucose load in the patient with somatostatinoma. The shaded areas represent the response of nine normal subjects (mean \pm SD) to the same glucose load (57).

It is also noteworthy that a needle biopsy of the patient's quadriceps muscle revealed thickening of the capillary basement membrane ($2219 \pm 349 \text{ \AA}$) (diabetic range above 1600 \AA) (59). This finding also makes the patient's diabetes indistinguishable from the common forms of diabetes mellitus.

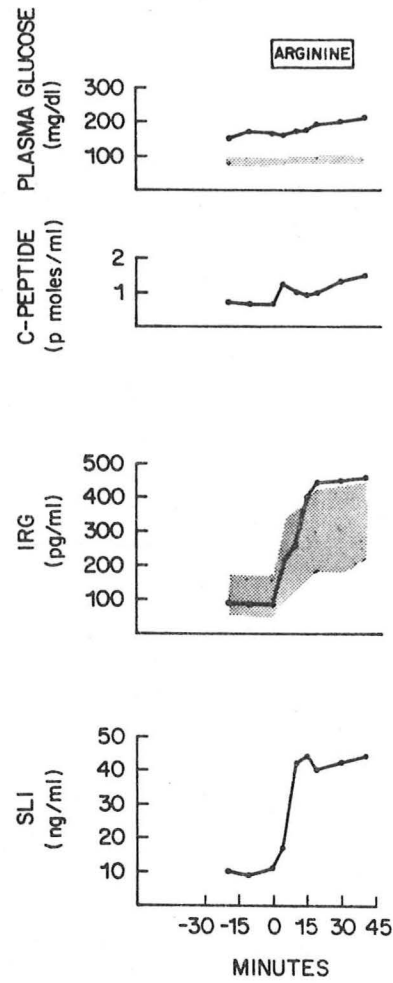


Figure 7: Plasma glucose, C-peptide, IRG and SLI response to intravenous arginine in the patient with somatostatinoma. The shaded areas represent the response of 28 normal subjects (mean \pm SD) to a similar intravenous infusion of arginine (58).

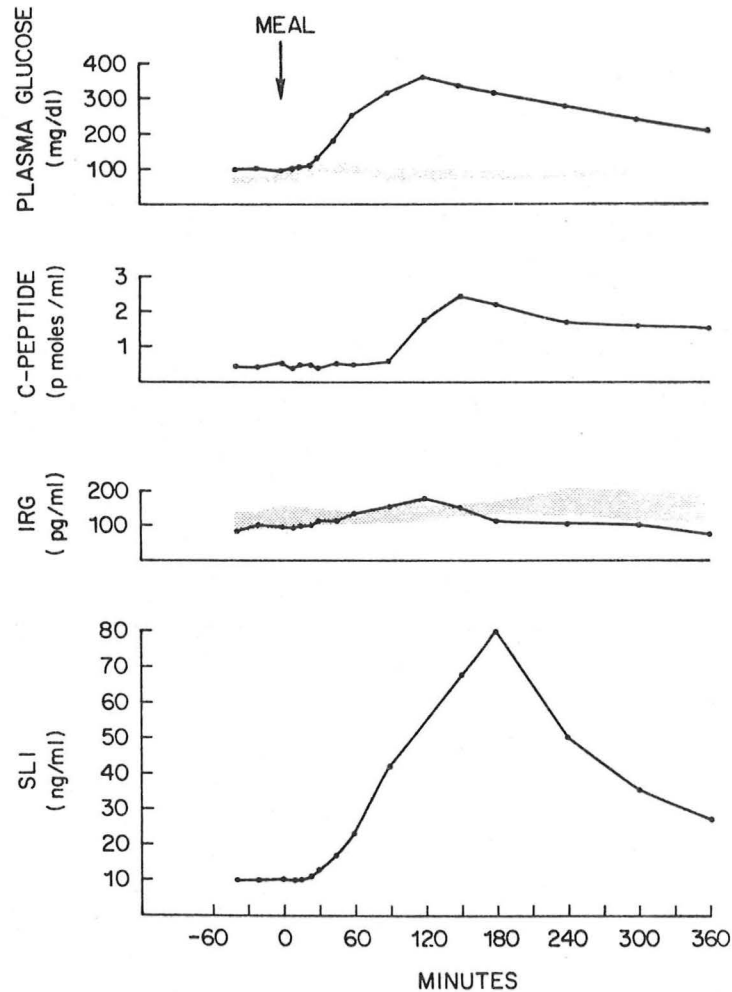


Figure 8: Plasma glucose, C-peptide, IRG and SLI response to a mixed meal in the somatostatinoma patient. The shaded areas represent the response of ten normal subjects (mean \pm SD) to the same meal.

3. Gastrointestinal Hormones

Somatostatin suppresses the release of gastrin (60), CCK (61), secretin (62), motilin (63), GIP (64) and GLI (65). This may lead to a variety of gastrointestinal manifestations, including hypochlorhydria, indigestion, postprandial fullness, eructations, vomiting, abdominal pain, maldigestion, steatorrhea, diarrhea, and susceptibility to cholelithiasis secondary to reduced postprandial gallbladder contraction.

Table 3 shows gastrin and GIP responses to the mixed meal. Gastrin levels were normal. Gastrin rose to 210 pg/ml at a time when plasma somatostatin increased several fold (Fig 8). GIP levels were suppressed and did not rise in response to the meal in patient E.L.

TABLE 3. Postprandial serum concentration of gastrin and GIP. Blood samples were drawn during the mixed meal study depicted in Figure 8. GIP radioimmunoassay was performed by Drs. A. Shulkes and J. Walsh, Los Angeles, California (normal fasting level: <60 pmoles/l).

	Basal	<u>Time (min) after meal</u>						
		10	20	45	90	120	240	360
GASTRIN pg/ml	79	113	147	189	210	180	151	138
GIP fmoles/l	<10	<10	<10	<10	<10	<10	<10	<10

B. GASTROINTESTINAL FUNCTION

1. Gastric Acid Secretion

Somatostatin has been shown to be a potent inhibitor of gastric acid secretion (60,66).

Gastric acid analysis in patient E.L. revealed a BAO of 0.04 meq/h and a MAO (following pentagastrin) of 1.92 meq/h. Despite the patient's truncal vagotomy and pyloroplasty these numbers appear low and may be considered hypochlorhydric (maximal acid output following truncal vagotomy and pyloroplasty has been reported to average 6.7 ± 4.3 (range 2.6 to 16.0 meq/h) (67).

The patient's relative hypochlorhydria may be explained by a direct inhibitory influence of somatostatin on the parietal cells. However, with hypochlorhydria and a history of vagotomy one might expect very high postprandial gastrin levels and thus, this patient's normal gastrin values (Table 3) may indeed reflect an inhibitory effect of somatostatin on gastrin release.

Suction biopsy from the body of the stomach showed normal mucosal architecture with parietal cells appearing normal in number and morphology. There were no signs of superficial or atrophic gastritis. Histological examination of the resected antrum (Whipple's procedure) revealed no abnormalities.

2. Gastric Emptying

Several observations suggest that somatostatin may play a role in controlling motor activity of the digestive tract. Inhibition of gastric emptying has been described by several investigators (63,68,70).

In patient E.L. gastric emptying was studied by scintigraphy using a ^{99m}Tc -labeled meal (71). Gamma counts from the stomach were recorded with a computer via the gamma camera every five minutes for one hour. In the patient 100% of the activity was still in the stomach at 60 minutes. In normal controls $52 \pm 2\%$ (mean \pm SEM) of the

radioactivity is found to remain in the stomach at 60 minutes following the meal. The vagotomy and pyloroplasty in the patient would even be expected to enhance gastric emptying (72). Thus, delayed gastric emptying in this patient is consistent with the known action of high circulating levels of somatostatin.

3. Small Bowel Motility

The effect of somatostatin on small bowel motility is very complex and has not been investigated to a great extent. Somatostatin may inhibit motility by suppressing the release of such gastrointestinal hormones as motilin, gastrin, and cholecystokinin that are involved in the control of intestinal motor activity.

For the purpose of this Grand Rounds I would like to pick two motility parameters to discuss. One is intestinal transit time and the other is the interdigestive myoelectric complex.

When we studied normal controls by small bowel perfusion during somatostatin administration (26) we found that somatostatin almost doubled transit time in the test segment (Fig 9). Intestinal transit time was not studied in patient E.L.

Prolongation of transit time by somatostatin may be of benefit in diarrheal states that are at least in part caused by an intestinal motor disorder. One possible application is the malignant carcinoid syndrome and we have, in fact, markedly reduced daily stool weight in such a patient during somatostatin infusion (74).

The second motility parameter is the interdigestive migrating complex (75). The cyclic recurrence of the phenomenon and the migration of the activity front down the intestine is thought to clean up the remnants of the preceding meal and function as the "housekeeper" of the small intestine (76).

This migrating myoelectric complex is abolished by feeding or infusion of gastrin, cholecystokinin and insulin.

An interesting observation was recently made in dogs where somatostatin infusion was shown to double the frequency of the interdigestive myoelectric complex. Also, after feeding, somatostatin converted the fed-type pattern to the fasting-type pattern with the onset of migrating complexes (77).

Small bowel motility was evaluated in patient E.L. by using a four-lumen polyvinyl tube with side openings 6 cm apart in the proximal small bowel. The tubes were constantly perfused with water at a slow rate and connected to external Statham transducers and a multichannel recorder.

In patient E.L. six migrating myoelectric complexes were observed in the fasting state at a mean interval of 38 min (range 33 to 48), complexes lasted for 274 sec (range 220 to 284) and propagation velocity was 2.1 cm/min (range 1.2 to 3.2). There was myoelectrical quiescence between the migrating myoelectrical complexes. However, the lack of a crescendo activity before and a decrescendo activity following the complexes may have been due to the patient's vagotomy. In 11 normal subjects migrating myoelectric complexes were observed at intervals of 120.3 ± 14.0 min, the duration of complexes was 420 ± 36 sec and the propagation velocity was 3.9 ± 0.5 cm/min. The patient did not tolerate eating with the tube in place and the motility pattern of the fed state could not be studied.

The results of this patient's motility study are thus consistent with experimental results observed in the canine intestine (77).

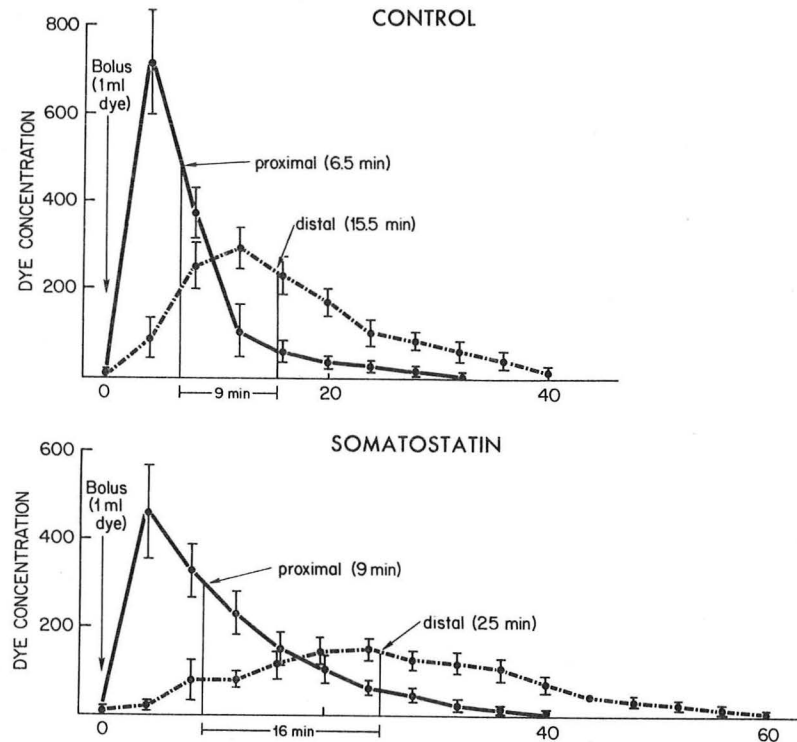


Figure 9: Dye dilution curves for estimating mean transit time in a 30 cm jejunal test segment. Ordinate gives normalized dye concentration (^3H -mannitol, ^{14}C urea and BSP were used). Abscissa gives time in minutes. Dye concentration was measured every 4 minutes at the proximal and distal sampling site. Mean transit time is estimated between the vertical lines that separate half the area under each curve (73). Somatostatin prolonged mean transit time from 9 to 17 minutes. Atropine (bottom) prolonged transit time from 9 to 19 minutes.

It is possible that this motility disturbance played a role in the patient's abdominal discomfort and dyspeptic symptoms.

4. Small Intestinal Absorption

Somatostatin reduces blood levels of sugars, Ca isotope and triglycerides after oral test meals (78-81). This suggests but does not prove reduction of absorption. One physiological role proposed for somatostatin by Dr. Unger and his associates (82) is the regulation of nutrient influx from the gastrointestinal tract into the circulation in coordination with the insulin-mediated regulation of nutrient disposal to the tissues. Thus the islets of Langerhans with its A, B and D cells would have control over all routes of nutrient flux: influx from storage sites (glucagon), disposal to tissues (insulin) and nutrient uptake by the intestine (somatostatin).

To test whether somatostatin directly affects small bowel absorption we have used a steady-state perfusion technique in the jejunum of healthy volunteers. The test solutions contained various nutrients and perfusion was performed during either saline (control) or somatostatin infusion (26).

Figure 10 shows the effect of somatostatin on glucose absorption from the human jejunum. Glucose absorption is decreased by somatostatin with a reduction of V_{max} by about 30 percent.

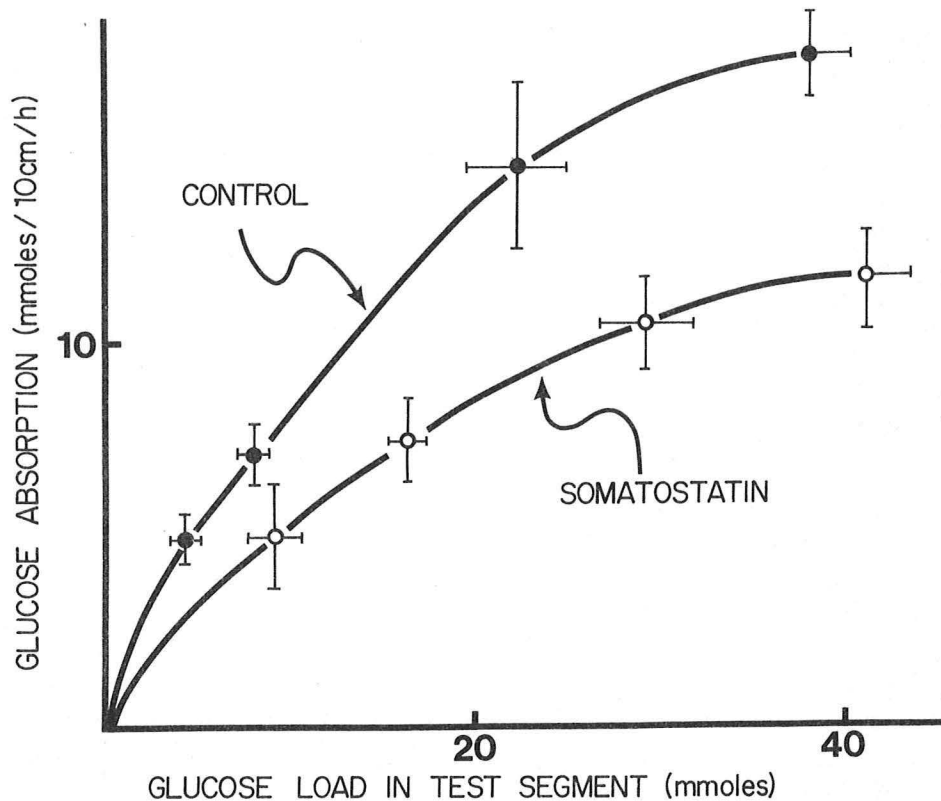


Figure 10: Kinetic curves for glucose absorption from 10 cm test segments of human jejunum. Each point represents mean \pm SEM of 5 observations. Lineweaver-Burk analysis showed reduction of the V_{max} for glucose absorption (17.9 and 11.4 mmoles/10 cm/h for control and somatostatin periods, respectively) while the k_m values were similar (8.2 and 9.3 mmoles, respectively).

Table 4 compares jejunal absorption of sugars, amino acids and oleic acid in healthy subjects and the patient. The values observed in the patient are similar to the absorption rates seen in healthy subjects when somatostatin is given by continuous intravenous infusion (8 μ g/kg/h).

TABLE 4. SMALL BOWEL PERFUSION STUDY
NET ABSORPTION FROM TEST SEGMENT
(PERCENT OF TOTAL AMOUNT OF SOLUTE ENTERING TEST SEGMENT)

ABSORBED SOLUTE	<u>HEALTHY SUBJECTS</u>		<u>PATIENT</u>
	CONTROL	SOMATOSTATIN INFUSION	"NO INFUSION"
GLUCOSE	65	36	49
FRUCTOSE	36	24	29
XYLOSE	18	16	14
GLYCINE	35	21	20
LYSINE	22	18	14
OLEIC ACID	57	47	49

In our patient pancreatic exocrine function was assessed by a secretin test, a Lundh meal and stool fat analysis.

In response to exogenous secretin the patient's duodenal aspirate was normal with respect to volume (3.8 ml/kg/80 min, normal >2.0) but bicarbonate concentration and output were borderline low (conc: 60 meq/liter, normal >70; output: 0.2 meq/kg/h, normal >0.2).

In response to the Lundh meal, however, there was only a minimal rise in amylase and lipase concentration suggesting either failure to release the hormones in the duodenal mucosa that induce pancreatic secretion or a direct inhibitory effect of somatostatin on the pancreas (increased postprandial SLI levels: see Fig 8). This abnormal test result explains the patient's third cardinal symptom, namely steatorrhea. That the steatorrhea was of pancreatic etiology is further supported by the decrease of fecal fat on pancreatic enzyme replacement (Table 5).

TABLE 5. FECAL FAT ON 100 g FAT DIET IN PATIENT E.L.

	Stool Weight (g/24h)	Stool Fat (g/24h)
OFF MEDICATION	520	31
PANCREATIC ENZYME REPLACEMENT THERAPY	357	4

Table 6 summarizes the comparison between the known pharmacological actions of somatostatin and the findings in the patient with malignant somatostatinoma syndrome.

TABLE 6.

<u>PHARMACOLOGICAL ACTIONS OF SOMATOSTATIN</u>		<u>FINDINGS IN PATIENT E.L.</u>
<u>CARDINAL SYMPTOMS</u>	<u>REDUCTION OR INHIBITION OF</u>	
	Gallbladder motility (CCK-PZ release)	Gallstones
	Insulin release	Mild diabetes
	Pancreatic bicarbonate and enzyme secretion	Steatorrhea
	<u>OTHER ENDOCRINE FUNCTIONS</u>	
	GH-release	Absent growth hormone response to arginine and insulin
	TSH release	Normal TSH response to TRH
	Glucagon	Normal glucagon response to arginine
	Gastrin	Normal gastrin (relative hypo-gastrinemia?)
	GIP	No postprandial rise of GIP
	Renin activity	Not studied
	Erythropoietin	Not studied (normocytic, normochromic anemia)
	<u>OTHER GASTROINTESTINAL FUNCTIONS</u>	
	HCl and pepsin secretion	"Relative hypochlorhydria" (vagotomy)
	Gastric emptying	Delayed despite vagotomy and pyloroplasty
	Small bowel absorption of sugars, amino acids and fatty acids	Reduced in comparison to normal controls
	Intestinal motility	Abnormal
	Salivary secretion	Not studied (dry mouth)

The clinical features observed in the six patients with somatostatinoma syndrome that have thus far been reported are listed in Table 7.

TABLE 7. Review of clinical manifestations in 6 patients with somatostatinoma. (+) present, (-) absent, NS: not studied, NR: not reported, W: Whipple's procedure, CH: Chemotherapy with streptozotocin, EL: Exploratory laparotomy

Author	Patient's Age and Sex	Patient's Home Country	Cholelithiasis	Diabetes Mellitus	Steatorrhea	Diarrhea	Hypochlorhydria	Anemia	Weight loss	Liver Metastases	Localisation of Primary Pancreatic Tumor	Additional Features	Treatment	Outcome
Larson (2)	55 f	Sweden	+	+	+	+	+	NR	+	+	Head	none	W	Died postop.
Ganda (3,84)	46 f	USA	+	+	NS	-	-	+	+	-	Head	none	W	Asymptomatic 24 months later
Kovacs (4)	54 m	Canada	-	+	NS	-	NS	NR	+	+	Tail	ACTH production	EL	Died postop.
De Nutte (5)	NR	Belgium	+	+	+	-	NS	NR	+	+	NR	none	CH	Not known
Galmiche (6)	70 f	France	+	+	+	+	+	NR	+	+	NR	Calcitonin production	W	Died postop.
Present Case (1)	52 m	Canada	+	+	+	+	+	+	+	+	Head	Calcitonin production	W,CH	Clinical and biochemical improvement

FOLLOWUP OBSERVATIONS IN E.L.

Following intravenous hyperalimentation the patient's plasma somatostatin level had risen from about 10,000 to 20,000 pg/ml at the time of surgery. This was accompanied by

a marked increase in the size of the liver metastases as followed by angiography.

After resection of the primary tumor plasma SLI dropped to 17,000 pg/ml and decreased further after subsequent chemotherapy (Table 8).

An interesting observation was made when the patient's plasma after tumor resection was again subjected to gel filtration chromatography. While pure somatostatin was the major fraction of total plasma SLI before surgery (1600 dalton), postoperatively the precursor form (3,000 dalton) is predominating. This suggests that the liver metastases mainly release the high molecular precursor form of somatostatin while the primary pancreatic tumor released the pure decapeptide.

Complications during his workup and treatment included a postoperative wound infection which resolved, an a-v fistula in a liver metastasis and a hepatic artery aneurysm, both of which were successfully occluded by balloon and embolization.

TABLE 8. COURSE OF PLASMA SLI (PG/ML) IN PATIENT E.L.

Aug./Sept. 1978	Diagnostic workup	9,000 - 13,000
October 1978	Preoperatively, (after six weeks of intravenous hyperalimentation)	20,000 - 25,000
November 1978	Postoperatively	17,000
December 1978	Chemotherapy	14,000
January 1979	(streptozotocin by	11,000
February 1979	hepatic artery infusion)	7,000

The patient is clinically improving, has gained 4 kg since surgery, and his insulin requirements which had risen to 40 U Lente per day preoperatively are now down to 7 U in April, 1979. Followup angiography showed 30% shrinkage of the liver metastases in April, 1979.

EARLY DIAGNOSIS

The early diagnosis of the somatostatinoma syndrome is clearly difficult. A benign somatostatinoma, such as that reported by Ganda et al. (3), in which plasma SLI was not measured, might well produce only intermittent hypersomatostatinemia. In this event a provocative test with a substance such as tolbutamide, a powerful stimulus of pancreatic somatostatin release (85), may be required to expose the abnormality, as was first employed by Nutte et al. (5). The early features are nonspecific and common, a

triad of dyspepsia, mild diabetes and cholelithiasis. Both diabetes, which was present in all cases, are common in the older age group in which all of the somatostatinomas have thus far been found.

Steatorrhea in the absence of pancreatitis may be a clue to the diagnosis, but the only patient without evidence of metastases did not have steatorrhea (3).

Perhaps the early diagnosis of benign somatostatinoma may require localization of the primary tumor. In the present case, the tumor was present at least 4 years prior to the definite diagnosis. During that period, symptoms were limited to postprandial eructation and periumbilical cramping after heavy meals and were attributed to the patient's gallstones. Late in 1977, however, the relatively abrupt onset of severe weight loss, steatorrhea, diabetes and the finding of a serum alkaline phosphatase at the upper limit of normal may herald the development of hepatic metastases and of more severe hypersomatostatinemia. When a presumably endocrine pancreatic tumor distorts the duodenum (Fig 1), its removal is urgently indicated and studies of survival in patients with the Zollinger-Ellison syndrome indicate that this location is associated with the most favorable prognosis. With the availability of ultrasonography and arteriography, pancreatic tumors are more easily detected. Since somatostatinoma may not be rare, one wonders if blood somatostatin should be measured in the workup of a diabetic patient with dyspepsia, or gallstones.

A plasma SLI radioimmunoassay is available at several centers. A survey of patients with such diseases as gallstones, diabetes mellitus, and exocrine pancreatic insufficiency may reveal if such a screening is clinically relevant. No disease other than the somatostatinoma syndrome is known to cause severe hypersomatostatinemia, although juvenile-type diabetes may be associated with modest elevations (86,87). If basal levels of SLI are normal in a patient with a pancreatic tumor, a provocation test with tolbutamide or another stimulus may be of value.

SUMMARY

Knowledge of the pharmacological actions of somatostatin allowed recognition of the cause of this patient's syndrome and availability of a radioimmunoassay confirmed the diagnosis of somatostatinoma. Cardinal symptoms were cholelithiasis, diabetes and steatorrhea associated with a pancreatic tumor. Further pathophysiological studies showed - with a few exceptions - that the patient's endocrine and gastrointestinal function was consistent with the effects of somatostatin excess predicted from experimental observations.

Six patients with somatostatinoma - including the present case - have been reported. The true incidence of the disease, however, is not known at this time, since general awareness of this syndrome among physicians has probably not been reached.

Future experience will have to show whether a provocation test with tolbutamide or screening of plasma somatostatin levels by radioimmunoassay may be beneficial in the early recognition of this disease.

APPENDIXPotential Clinical Use of Somatostatin

The original publication by Guillemin and co-workers about the discovery of somatostatin ends with the following statement: "Should SRIF be active in humans, its possible clinical significance, particularly in the treatment of acromegaly and the management of juvenile diabetes, has not escaped our attention"(10).

Limitations for clinical use are caused by two characteristics of somatostatin: its short-lived action and its multiple effects. Attempts are being made to produce somatostatin analogues that are long acting and only enhance certain actions of the peptide (88).

1. Diseases for which clinical studies or single case reports have suggested a potential benefit:

Acromegaly (48,89, 90)

Diabetes mellitus (91-93)

Insulinoma, glucagonoma, VIPoma, gastrinoma: preoperative or palliative use suggested (88,94)

Malignant carcinoid syndrome (74,95)

Upper gastrointestinal bleeding due to peptic ulcer disease or esophageal varices (96-98)

2. No benefit:

Extrapyramidal disorders and EEG abnormalities (99)

Experimental pancreatitis in rats (100)

REFERENCES

1. Krejs, G.J., Orci, L., Conlon, J.M., Ravazzola, M., Davis, G.R., Raskin, P., Collins, S.M., McCarthy, D.M., Baetens, D., Rubenstein, A. and Unger, R.H.: Somatostatinoma Syndrome - Biochemical, Morphological and Clinical Features. (Submitted for publication)
2. Larsson, L.I., Holst, J.J., Kuhl, C., Lundquist, G., Hirsch, M.A., Ingemansson, S., Lindkaer Jensen, S., and Rehfeld, J.F.: Pancreatic somatostatinoma: Clinical features and physiological implications. LANCET 26:666-668, 1977
3. Ganda, O.P., Weir, G.C., Soeldner, J.S., Legg, M.A., Chick, W.L., Patel, Y.C., Ebeid, A.M., Gabbay, K.H. and Reichlin, S.: "Somatostatinoma": A somatostatin-containing tumor of the endocrine pancreas. N ENGL J MED 296:964-967, 1977
4. Kovacs, K., Horvath, E., Ezrin, C., Sepp, H. and Elkan, I.: Immunoreactive somatostatin in pancreatic islet-cell carcinoma accompanied by ectopic A.C.T.H. syndrome. LANCET i:1365-1366, 1977
5. De Nutte, N., Somers, G., Gepts, W., Jacobs, M. and Pipeleers, D.: Pancreatic hormone release in tumour-associated hypersomatostatinemia. DIABETOLOGIA 15:227, 1978
6. Galmiche, J.P., Colin, R., DuBois, P.M., Chayvialle, J.A., Descos, F., Paulin, C. and Geffroy, Y.: Calcitonin secretion by a pancreatic somatostatinoma. NEW ENGL J MED 299:1252, 1978
7. Krulich, L., Dhariwal, A.P.S. and McCann, S.M.: Stimulatory and inhibitory effects of purified hypothalamic extracts on growth hormone release from rat pituitary in vitro. ENDOCRINOLOGY 83:783-790, 1968
8. Krulich, L. and McCann, S.M.: Effect of GH-releasing factor and GH-inhibiting factor on the release and concentration of GH in pituitaries incubated in vitro. ENDOCRINOLOGY 85:319-324, 1969
9. Krulich, L., Illner, P., Fawcett, C.P., Quijada, M., and McCann, S.M.: Dual hypothalamic regulation of growth hormone secretion. EXCERPTA MEDICA INTERNATIONAL CONGRESS SERIES 244:306-316, 1972
10. Brazeau, P., Vale, W., Burgus, R., Ling, N., Butcher, M., Rivier, J. and Guillemin, R.: Hypothalamic polypeptide that inhibits the secretion of immunoreactive pituitary growth hormone. SCIENCE 179:77-79, 1973
11. Burgus, R., Brazeau, P. and Vale, W.: Isolation and determination of the primary structure of somatostatin (a somatotropin release inhibiting factor) of ovine hypothalamic origin. DHEW Publ. 74-612 (NIH), 1973, p. 144-158
12. Rivier, J.E.F.: Somatostatin. Total solid phase synthesis. J AMER CHEM SOC 96:2986-2992, 1974
13. Arimura, A., Sato, H., Coy, D. and Schally, A.V.: Radioimmunoassay for GH-release inhibiting hormone. PROC SOC EXPER BIOL MED 148:784-789, 1975
14. Harris, V., Conlon, J.M., Srikant, C.B., McCorkle, K., Schusdziarra, V., Ipp, E. and Unger, R.H.: Measurements of somatostatin-like immunoreactivity in plasma. CLIN CHIM ACTA 87:275-283, 1978

15. Itakura, K., Hirose, T., Crea, R., Riggs, A.D., Heyneker, H.L., Bolivar, F. and Boyer, H.W.: Expression in *Escherichia coli* of a chemically synthesized gene for the hormone somatostatin. *SCIENCE* 198:1056-1063, 1977
16. Barker, J.L.: Peptides: roles in neuronal excitability. *PHYSIOL REV* 56:435-452, 1976
17. Dubois, M.: Immunoreactive somatostatin is present in discrete cells of the endocrine pancreas. *PROC NATL ACAD SCI USA* 72:1340-1343, 1975
18. Luft, R., Efendic, S., Hokfelt, T., Johansson, O. and Arimura, A.: Immunohistochemical evidence for the localization of somatostatin-like immunoreactivity in a cell population of the pancreatic islets. *MED BIOL* 52:428-430, 1974
19. Hokfelt, T., Efendic, S., Hellerstrom, C., Johansson, O., Luft, R. and Arimura, A.: Cellular localization of somatostatin in endocrine-like cells and neurons of the rat with special reference to the A₁-cells of the pancreatic islets and to the hypothalamus. *ACTA ENDOCRINOL* 80:(Suppl. 200), 1-40, 1975
20. Orci, L., Baetens, D., Dubois, M.P. and Rufener, C.: Evidence for the D-cell of pancreas secreting somatostatin. *HORM METAB RES* 7:400-402, 1975
21. Pelletier, G., Leclerc, R., Arimura, A. and Schally, A.V.: Immunohistochemical localization of somatostatin in the rat pancreas. *J HISTOCHEM CYTOCHEM* 23:699-701, 1975
22. Polak, J.M., Grimelius, L., Pearse, A.G.E., Bloom, S.R. and Arimura, A.: Growth-hormone release-inhibitory hormone in gastrointestinal and pancreatic D-cells. *LANCET* ii:1220-1222, 1975
23. Rufener, C., Dubois, M., Malaisse-Lagae, F. and Orci, L.: Immuno-fluorescent reactivity to anti-somatostatin in the gastro-intestinal mucosa of the dog. *DIABETOLOGIA* 11:321-324, 1975
24. Forssman, W.G., Helmstaedter, V., Metz, J., Muhlmann, G. and Feuerle, G.E.: Immunohistochemistry and ultrastructure of somatostatin cells with special reference to the gastro entero pancreatic system. *METABOLISM* 27:1179-1191, 1978
25. Orci, L. and Unger, R.: Hypothesis: functional subdivisions of islets of Langerhans and possible role of D cells. *LANCET* 2:1243-1244, 1975
26. Krejs, G.J., Raskin, P. and Fordtran, J.S.: Inhibition of jejunal sugar and amino acid absorption in man: a nonspecific effect of somatostatin. *CLIN RES* 26:420A, 1978
27. Conlon, J.M., Srikant, C.B., Ipp, E., Schusdziarra, V., Vale, W. and Unger, R.H.: Properties of endogenous somatostatin-like immunoreactivity and synthetic somatostatin in dog plasma. *J CLIN INVEST* 62:1187-1193, 1978
28. Krohnheim, S., Berelowitz, M. and Pimstone, B.L.: The characterization of somatostatin-like immunoreactivity in human serum. *DIABETES* 27:523-529, 1978
29. Arimura, A., Sato, H., Dupont, A., Nishi, N. and Schally, A.V.: Somatostatin: abundance of immunoreactive hormone in rat stomach and pancreas. *SCIENCE* 189:1007-1009, 1975

30. Schally, A.V., Dupont, A., Arimura, A., Redding, T.W. and Linthicum, G.L.: GH-release inhibiting hormone (GH-RIH): the existence of three forms of GH-RIH. *FED PROC* 34:584, 1975
31. Schally, A.V., Dupont, A., Arimura, A., Redding, T.W., Nishi, N., Linthicum, G. and Schlesinger, D.: Isolation and structure of somatostatin from porcine hypothalamic. *BIOCHEMISTRY* 15:509-514, 1976
32. Noe, B.D., Weir, G.C. and Bauer, G.E.: Biosynthesis of somatostatin in pancreatic islets. *METABOLISM* 27 (Suppl 1): 1201-1205, 1978
33. Vale, W., Ling, N., Rivier, J., Villarreal, J., Rivier, C., Douglas, C. and Brown, M.: Somatostatin, anatomic and phylogenetic distribution of somatostatin. *METABOLISM* 25 (Suppl 1): 1491-1494, 1976
34. Pradayrol, J., Chayvialle, J. and Mutt, V.: Pig duodenal somatostatin: extraction and purification. *METABOLISM* 27 (Suppl 1): 1197-1200, 1978
35. McIntosh, C., Arnold, R., Bothe, E., Becker, H., Kobberling, J. and Creutzfeldt, W.: Gastrointestinal somatostatin: extraction and radioimmunoassay in different species. *GUT* 19:655-663, 1978
36. Dupont, A., Alvarado-Urvina, G.: Conversion of big pancreatic somatostatin without peptide bond cleavage into somatostatin tetradecapeptide. *LIFE SCI* 19: 1431-1433, 1976
37. Chayvialle, J.A.P., Descos, F., Bernard, C., Martin, A., Barbe, C. and Paetensky, C.: Somatostatin in mucosa of stomach and duodenum in gastroduodenal disease. *GASTROENTEROLOGY* 75:13-19, 1978
38. Patel, Y.C. and Reichlin, S.: Somatostatin in hypothalamus, extrahypothalamic brain and peripheral tissues of the rat. *ENDOCRINOLOGY* 102:523-530, 1978
39. Millar, J.: Somatostatin immunoreactive peptides of higher molecular weight in ovine hypothalamic extracts. *J ENDOCRINOL* 77:429-430, 1978
40. Spiess, J. and Vale, W.: Evidence for larger forms of somatostatin in pigeon pancreas and rat brain. *METABOLISM* 27 (Suppl 1): 1175, 1978
41. Conlon, J.M., Zyznar, E. and Unger, R.H.: Multiple forms of somatostatin-like immunoreactivity in canine pancreas. *FEBS LETT* 94:327-330
42. Noe, B.D., Fletcher, D.J., Bauer, G.E., Weir, G.C. and Patel, Y.: Somatostatin biosynthesis occurs in pancreatic islets. *ENDOCRINOLOGY* 102:1675-1685, 1978
43. McIntosh, C., Arnold, R., Bothe, E., Kobberling, J. and Creutzfeldt, W.: Gastrointestinal somatostatin in man. *METABOLISM* 27 (Suppl 1): 1317-1320, 1978
44. Weir, G.C., Horton, E.S., Aoki, T.T., Slovik, D., Jaspar, J. and Rubenstein, A.H.: Secretion by glucagonomas of a possible glucagon precursor. *J CLIN INVEST* 59: 325-330, 1977
45. Bloom, S.R. and Polak, J.M.: The glucagonoma syndrome. Gastrointestinal Hormones and Pathology of the Digestive System. Edited by M. Grossman, V. Speranza, N. Basso, E. Lezoche, New York and London, Plenum Press 1978, pp. 183-194

46. Bloom, S.R., Bryant, M.G. and Adrian, T.E.: Multiple hormone forms in PPomas, VIPomas and glucagonomas. SCAND J GASTROENT 13 (Suppl 49): 24, 1978
47. Krejs, G.J. and Fordtran, J.S.: Diagnostic and pathophysiologic studies in patients with chronic secretory diarrhea. In: Secretory Diarrhea. Edited by M. Field, S. Schultz and J.S. Fordtran. In print.
48. Hall, R., Schally, A.V., Evered, D., Kastin, A.J., Mortimer, C.H., Turnbridge, W.M.G., Besser, G.M., Coy, D.H., Goldie, D.J., McNeilly, A.S., Phenekos, C. and Weightman, D.: Actions of growth hormone release inhibiting hormone in healthy men and acromegaly. LANCET ii:581-584, 1973
49. Siler, T.M., Vandenberg, G., Yen, S.S.C., Brazeau, P., Vale, W. and Guillemin, R.: Inhibition of growth hormone release in humans by somatostatin. J CLIN ENDOCRINOL METAB 37:632-634, 1973
50. Siler, T., Yen, S., Vale, W. and Guillemin, R.: Inhibition by somatostatin of the release of TSH induced in man by thyrotropin-releasing factor. J CLIN ENDOCRINOL METAB 38:742-754, 1974
51. Prange, H.A., Orskov, H., Seyer-Hansen, K. and Lundback, K.: Source actions of growth hormone release inhibiting factor. BR MED J 3:523-524, 1973
52. Parker, D.C., Rossman, L.G., Siler, T.M., Rivier, J., Yen, S.S.C. and Guillemin, R.: Inhibition of sleep related peak in physiologic human growth hormone release by somatostatin. J CLIN ENDOCRINOL METAB 38:496-499, 1974
53. Carr, D., Gomez-Pan, A., Weightman, D., Roy, V.C.M., Hall, R., Besser, G.M., Thorner, M.O., McNeilly, A.S., Schally, A.V., Kastin, A.J. and Coy, D.H.: Growth hormone-release inhibiting hormone. Actions on thyrotropin and prolactin secretion after thyrotropin releasing hormone. BR MED J 3:67-69, 1975
54. Gerich, J.E., Lovinger, R. and Grodsky, G.M.: Inhibition by somatostatin of glucagon and insulin release from the perfused rat pancreas in response to arginine, isoproterenol, and theophylline: evidence for a preferential effect on glucagon secretion. ENDOCRINOLOGY 96:749-754, 1975
55. Marco, J., Hedo, J.A. and Villaneuva, M.L.: Inhibitory effect of somatostatin on human pancreatic polypeptide secretion. LIFE SCI 21:789-792, 1977
56. Unger, R.H.: Somatostatinoma. N ENGL J MED 296:998-1000, 1977
57. Aydin, I., Raskin, P. and Unger, R.H.: The effect of short term intravenous insulin administration on the glucagon response to a carbohydrate meal in adult onset and juvenile type diabetes. DIABETOLOGIA 13:629-637, 1977
58. Unger, R.H., Aquilar-Parada, E., Muller, W.A. and Eisentraut, A.M.: Studies of pancreatic alpha-cell function in normal and diabetic subjects. J CLIN INVEST 49:837-848, 1970
59. Siperstein, M.D., Raskin, P. and Burns, H.: Electron microscopic quantification of diabetic microangiopathy. DIABETES 22:514-527, 1973
60. Vatn, M.H., Schrupf, E., Hanssen, K.F. and Myren, J.: The effect of somatostatin on pentagastrin-stimulated gastric secretion and on plasma gastrin in man. SCAND J GASTROENT 12:833-839, 1977

61. Schlegel, W., Harvey, R.F., Raptis, S., Oliver, J.M. and Pfeiffer, E.F.: Inhibition of cholecystokinin-pancreozymin release by somatostatin. *LANCET* ii:166-168, 1977
62. Hanssen, L.E., Hanssen, K.F. and Myren, J.: Inhibition of secretin release and pancreatic bicarbonate secretion by somatostatin infusion in man. *SCAND J GASTROENT* 12:391-394, 1977
63. Bloom, S.R., Ralphs, D.N., Besser, G.M., Hall, R., Coy, D.H., Kastin, A.J. and Schally, A.V.: Effect of somatostatin on motilin levels and gastric emptying. *GUT* 16:834, 1975
64. Peterson, R.A., Dryburgh, J.R. and Brown, J.C.: The effect of somatostatin on release and insulinotropic action of gastric inhibitory polypeptide. *CAN J PHYSIOL PHARMACOL* 53:1200-1205, 1975
65. Sakurai, H., Dobbs, R.E. and Unger, R.H.: The effect of somatostatin on the response of GLI to the intraduodenal administration of glucose, protein and fat. *DIA-BETOLOGIA* 11:427-430, 1975
66. Konturek, S.J., Swierczek, J., Kwiecien, N., Mikos, E., Oleksy, J. and Wierzbicki, Z.: Effect of somatostatin on meal-induced gastric secretion in duodenal ulcer patients. *AM J DIG DIS* 22:981-983, 1977
67. Bell, P.R.F.: The long term effect of vagotomy on the maximal acid response to histamine in man. *GASTROENTEROLOGY* 46:387-391, 1964
68. Stadaas, J.O., Schrupf, E. and Hanssen, K.F.: Somatostatin inhibits gastric motility in response to distention. *SCAND J GASTROENT* 13:145-148, 1978
69. Boden, G., Jacoby, H.I. and Staus, A.: Somatostatin interacts with basal and carbachol stimulated antral and duodenal motility. *GASTROENTEROLOGY* 70:961, 1976
70. Johansson, C., Efendic, S., Wisen, O., Uvnäs-Wallensten, K. and Luft, R.: Effect of short-time somatostatin infusion on the gastric and intestinal propulsion in humans. *SCAND J GASTROENT* 13:481-483, 1978
71. McClelland, R.N. and Horton, J.W.: Relief of acute, persistent postvagotomy atony by metoclopramide. *ANN SURG* 188:439-447, 1978
72. Colmer, M.R., Owen, G.M. and Shields, R.: Pattern of gastric emptying after vagotomy and pyloroplasty. *BRIT MED J* 2:448-450, 1973
73. Dillard, R.L., Eastman, H. and Fordtran, J.S.: Volume-flow relationship during the transport of fluid through the human small intestine. *GASTROENTEROLOGY* 49: 58-66, 1965
74. Davis, G.R., Camp, R.C., Raskin, P. and Krejs, G.J.: Effect of somatostatin infusion on jejunal water and electrolyte transport in a patient with secretory diarrhea due to malignant carcinoid syndrome. (Submitted for publication).
75. Szurszewski, J.H.: A migrating electric complex of the canine small intestine. *AM J PHYSIOL* 217:1757-1763, 1969
76. Code, C.F. and Schlegel, J.F.: The gastrointestinal interdigestive housekeeper: motor correlates of the interdigestive myoelectric complex of the dog. In: *Proceedings of the Fourth International Symposium on Gastrointestinal Motility*. E.E. Daniel, J.A.L. Gilbert, B. Schofield, T.K. Schnitka and G. Scott, editors. Vandouwer Mitchell Press Ltd., Banff, Canada. 631-634

77. Thor, P., Krol, R., Konturek, S.V., Coy, D.H. and Schally, A.V.: Effect of somatostatin on myoelectrical activity of small bowel. *AM J PHYSIOL ENDOCRINOL METAB GASTROINTEST PHYSIOL* 4:E249-E254, 1978
78. Wahren, J. and Felig, P.: Influence of somatostatin on carbohydrate disposal and absorption in diabetes mellitus. *LANCET* ii:1213-1216, 1976
79. Pointner, V.H., Hengl, G., Bayer, P.M. and Flegel, U.: Hemmung des postprandialen Triglyzeridanstiegs im Serum durch Somatostatin beim Menschen. *WIEN KLIN WOCHENSCHR* 89:224-227, 1977
80. Evensen, D., Hanssen, K.F. and Berstad, A.: The effect on intestinal calcium absorption of somatostatin in man. *SCAND J GASTROENT* 13:449-451, 1978
81. Pott, G., Wagner, H., Zierden, E., Hilke, K.H., Jansen, H., Hengst, K. and Gerlach, V.: Influence of somatostatin on carbohydrate absorption in human small intestine. *KLIN WOCHENSCHR* 57:131-133, 1979
82. Schusdziarra, V., Ipp, E., Harris, V., Dobbs, R.E., Raskin, P., Orci, L. and Unger, R.H.: Studies of the physiology and pathophysiology of the pancreatic D-cell. *METABOLISM* 27 (Suppl 1): 1227-1232, 1978
83. Creutzfeldt, W., Lankisch, P.G. and Folsch, U.R.: Inhibition by somatostatin of pancreatic juice and enzyme secretion and gallbladder contraction in man induced by secretin and cholecystokinin-pancreozymin administration. *DTSCH MED WSCHR* 100:1135-1138, 1975
84. Ganda, O.P. and Soeldner, J.S.: "Somatostatinoma": Follow-up studies. *N ENGL J MED* 297:1352-1353, 1977
85. IPP, E., Dobbs, R.E., Arimura, A., Vale, W., Harris, V. and Unger, R.H.: Release of immunoreactive somatostatin from the pancreas in response to glucose, amino acids, pancreozymin-cholecystokinin, and tolbutamide. *J CLIN INVEST* 60:760-765, 1977
86. Schusdziarra, V., Dobbs, R., Raskin, P. and Unger, R.H.: Increased plasma immunoreactive somatostatin (IRS) levels in alloxan diabetic dogs and juvenile diabetic humans. *DIABETOLOGIA* 13:430, 1977
87. Hirsch, H.J. and Gabbay, K.H.: Radioimmunoassay of somatostatin-like immunoreactivity (SLI) in human plasma. *DIABETES* 27 (Suppl 2): 441, 1978
88. Lundbaek, K.: Somatostatin: clinical importance and outlook. *METABOLISM* 27 (Suppl 1): 1463-1469, 1978
89. Mortimer, C.H., Carr, D., Lind, T., Bloom, S.R., Mallinson, C.N., Schally, A.V., Tunbridge, W.M.G., Yeomans, L, Coy, D.H., Kastin, A., Besser, G.M. and Hall, R.: Effects of growth-hormone release-inhibiting hormone on circulating glucagon, insulin, and growth hormone in normal, diabetic, acromegalic, and hypopituitary patients. *LANCET* i:697-701, 1974
90. Yen, S., Siler, T. and DeVane, G.: Effect of somatostatin in patients with acromegaly. *N ENGL J MED* 290:935-938, 1974
91. Gerich, J.E.: Somatostatin. Its possible role in carbohydrate homeostasis and the treatment of diabetes mellitus. *ARCH INTERN MED* 137:659-666, 1977

92. Raskin, P. and Unger, R.H.: Hyperglucagonemia and its suppression. Importance in the metabolic control of diabetes. *N ENGL J MED* 299:433-436, 1978
93. Raskin, P.: The role of somatostatin in managing diabetes. *DRUG THERAPY* 8: 81-90, 1978
94. Guilleman, R. and Gerich, J.: Somatostatin: physiological and clinical significance. *ANN REV MED* 27:379-388, 1976
95. Dharmathaphorn, K., Sherwin, R.S., Binder, H.J. and Dobbin, J.W.: Somatostatin (SRIF) inhibits intestinal fluid secretion. *CLIN RES* 26:496A, 1978
96. Kayasseh, L., Gyr, K., Stalder, G.A. and Allgoewer, M.: Somatostatin in acute gastroduodenal hemorrhage. *LANCET* ii:833-834, 1978
97. Raptis, S. and Rosenthal, J.: Somatostatin: potential diagnostic and therapeutic value. *ACTA HEPATO GASTROENTEROL* 24:61-63, 1977
98. Tyden, G., Samnegard, H., Thulin, L., Friman, L. and Efendic, S.: Treatment of bleeding esophageal varices with somatostatin. *N ENGL J MED* 299:1466-1467, 1978
99. Dupont, E., Prange Hanse, A., Juul-Jensen, P., Lundbaek, K., Magnussen, I. and De Fine Olivarius, B.: Somatostatin in the treatment of patients with extrapyramidal disorders and patients with EEG abnormalities. *ACTA NEUROL SCANDINAV* 57:488-493, 1978
100. Lankisch, P.G., Koop, H., Winckler, K., Folsch, U.R. and Creutzfeldt, W.: Somatostatin therapy of acute experimental pancreatitis. *GUT* 18:713-716, 1977