

MEDICAL GRAND ROUNDS

PARKLAND MEMORIAL HOSPITAL

April 15, 1965

HEPATIC ENCEPHALOPATHY

Definition: Clinical syndrome of (1,2,3,4)

1. Abnormal mental state with
 - a. Disturbed consciousness
 - b. Personality changes
 - c. Intellectual deterioration
2. Abnormal neuromuscular state
3. Liver disease
4. Characteristic, but nonspecific, laboratory and EEG findings

Clinical Syndrome

1. Abnormal mental state
 - a. Disturbed consciousness:
 - Somnolence and inversion of sleep rhythm
 - Fixed stare
 - Apathy
 - Slowness of response (especially late in day)
 - Stupor → coma (in children and patients with viral hepatitis may see delirium)
 - b. Personality changes:
 - Euphoria
 - Irritability
 - Loss of concern for family
 - Inappropriate behavior
 - c. Intellectual deterioration:
 - Slight impairment of mental function → gross confusion
 - Inability to carry out simple calculations (subtraction of serial 7's)
 - Constructional apraxia (difficulty in writing and reproducing simple designs with blocks and matches)

2. Abnormal neuromuscular state

Asterixis (See characteristics in detail - Page 2)

Slurred speech

Perseveration

Grasping, incoordination

Ataxia

↑ DTR's early, ↓ or absent DTR's late

Hoffman and/or Babinski (often unilateral and transient)

Ankle clonus

Rigidity (even appearance of decerebration)

Choreoathetosis

Hyperventilation

Coma (at first arousable) → lack of response to any stimuli (loss of corneals)

Convulsions (especially in children and with acute liver disease)

Irreversible spastic paraplegia rarely - 6 cases (5,6).

"Asterixis" - (metabolic flap) (7)

- a. Movements consisting of lateral deviations of fingers and flexion - extension at metacarpo phalangeal and wrist joints. Best seen with arms outstretched, wrist hyperextended and fingers separated. Absent at rest, mitigated by intentional movement and maximal on sustained posture. Usually bilateral, often asynchronous. At intervals of fraction of second to a few seconds.
- b. May be seen on sustained contraction of other muscles (protrusion of tongue, dorsiflexion of foot).
- c. Associated with electromyographic periods of electrical silence. Believed due to inappropriate integration of afferent stimuli normally transmitted by ascending (?extrapyramidal) tracts to brain stem reticular formation (8).
- d. Not specific (9). Seen in uremia, severe pulmonary disease with heart failure, barbiturate intoxication, hypomagnesemia, etc.

3. Liver Disease - Failure of parenchyma and/or of hepatic circulation. "Essential" for development of encephalopathy (10).

Fetor hepaticus - Seen in 46% of one series (11). Sweetish musty smell. Believed due to methyl mercaptan (CH_3SH) or dimethyl sulfide ($\text{CH}_3\text{S:SCH}_3$) - both abnormal derivatives of methionine which accumulates in blood of patients with liver disease (12). Not seen in non-hepatic encephalopathy

4. Laboratory and EEG findings

- a. Laboratory:

Blood: (1) ↑ ammonia concentration, especially arterial. (See Figure 1 - Page 3)

The relation of the arterial ammonia level to the grade of consciousness in hepatic cirrhosis.

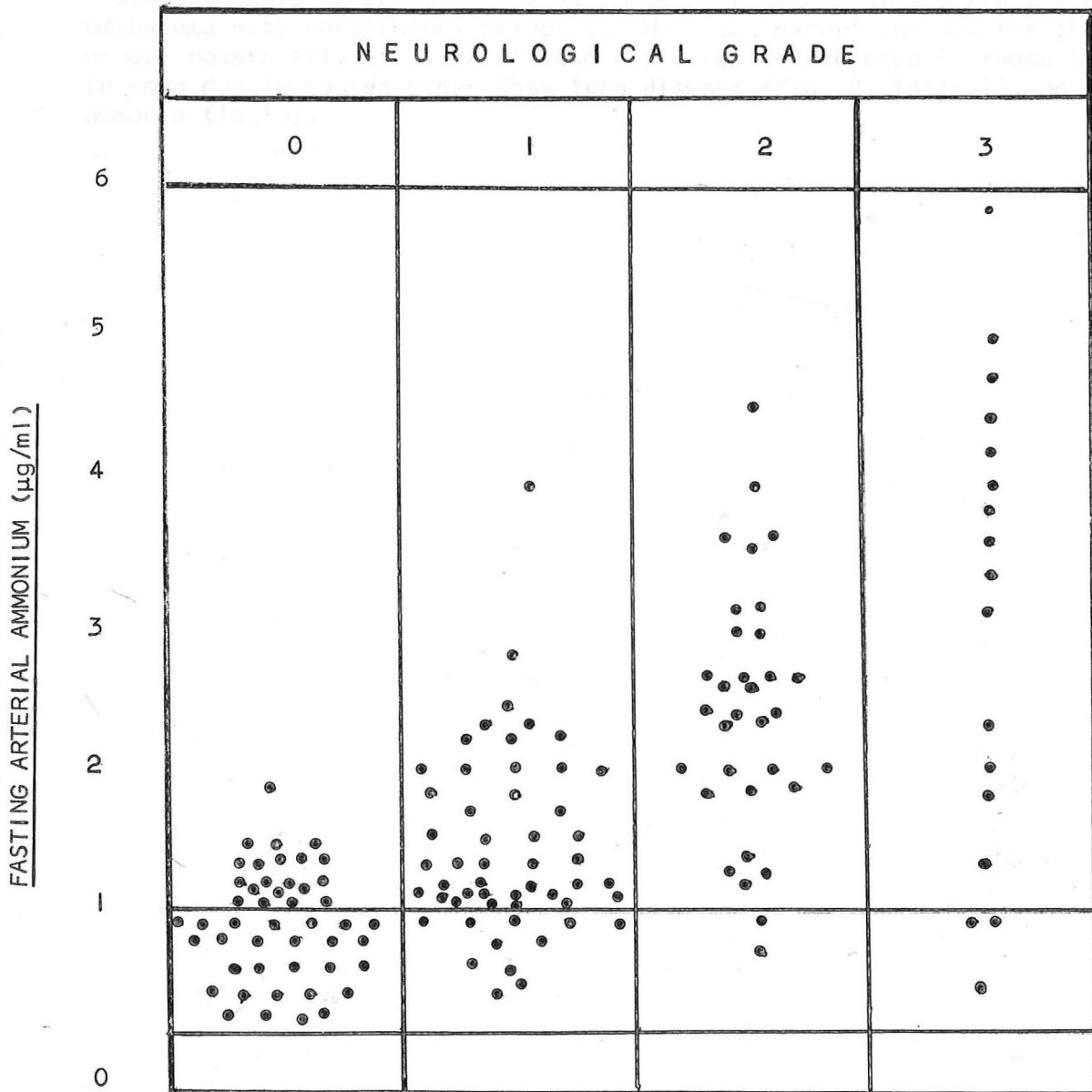
Grade 1: minor disorder of consciousness and the motor system.

Grade 2: gross disorder of consciousness with disorientation in time and space.

Grade 3: coma. Mean normal arterial ammonia level is 0.69 $\mu\text{g/ml}$.

The horizontal lines denote mean $\pm 2 \text{ S.D.}$

Figure 1 (13)



The relation of the arterial ammonium level to the grade of consciousness in hepatic cirrhosis.

Grade 1: minor disorder of consciousness and the motor system.

Grade 2: gross disorder of consciousness with disorientation in time and space.

Grade 3: coma. Mean normal arterial ammonium level is 0.68 µg/ml. The horizontal lines denote mean $\pm$ 2 S.D.

Mean values in patients with impending or fully developed hepatic coma are elevated. However 10 - 25% of arterial samples (Figure 1) and 25 - 50% of venous samples may be in normal range in individual patients and there is often no good correlation in patients with liver disease between degree of cerebral abnormality and ↑ in ammonia (13,14,15).

↑ blood ammonia reported in shock, congestive heart failure and pulmonary emphysema with respiratory acidosis. However, except perhaps for the last group, possibility of liver disease could not be excluded in these instances. In coma due to causes other than lung disease (Fig. 2, Table 1), no ↑ in ammonia (16,15).

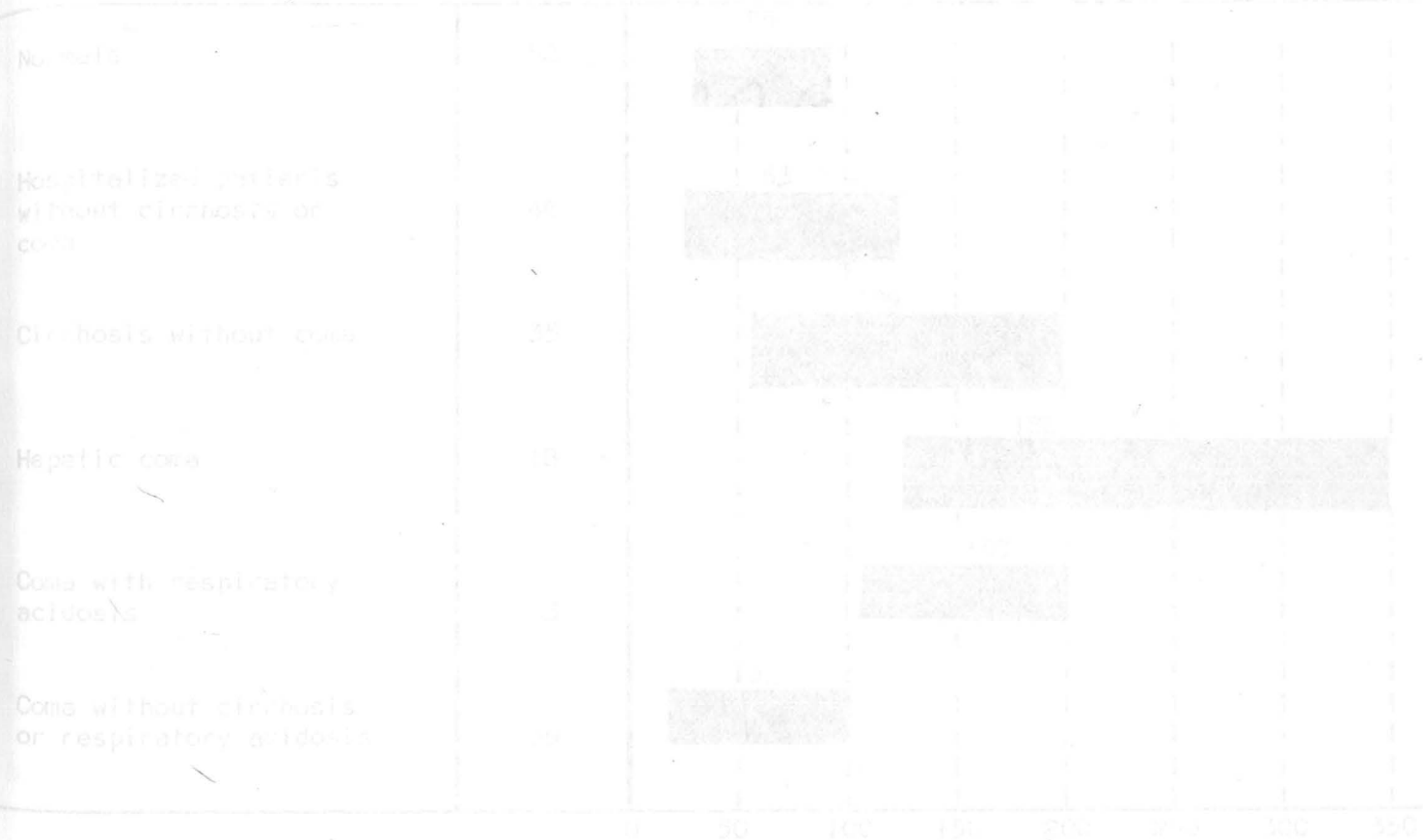
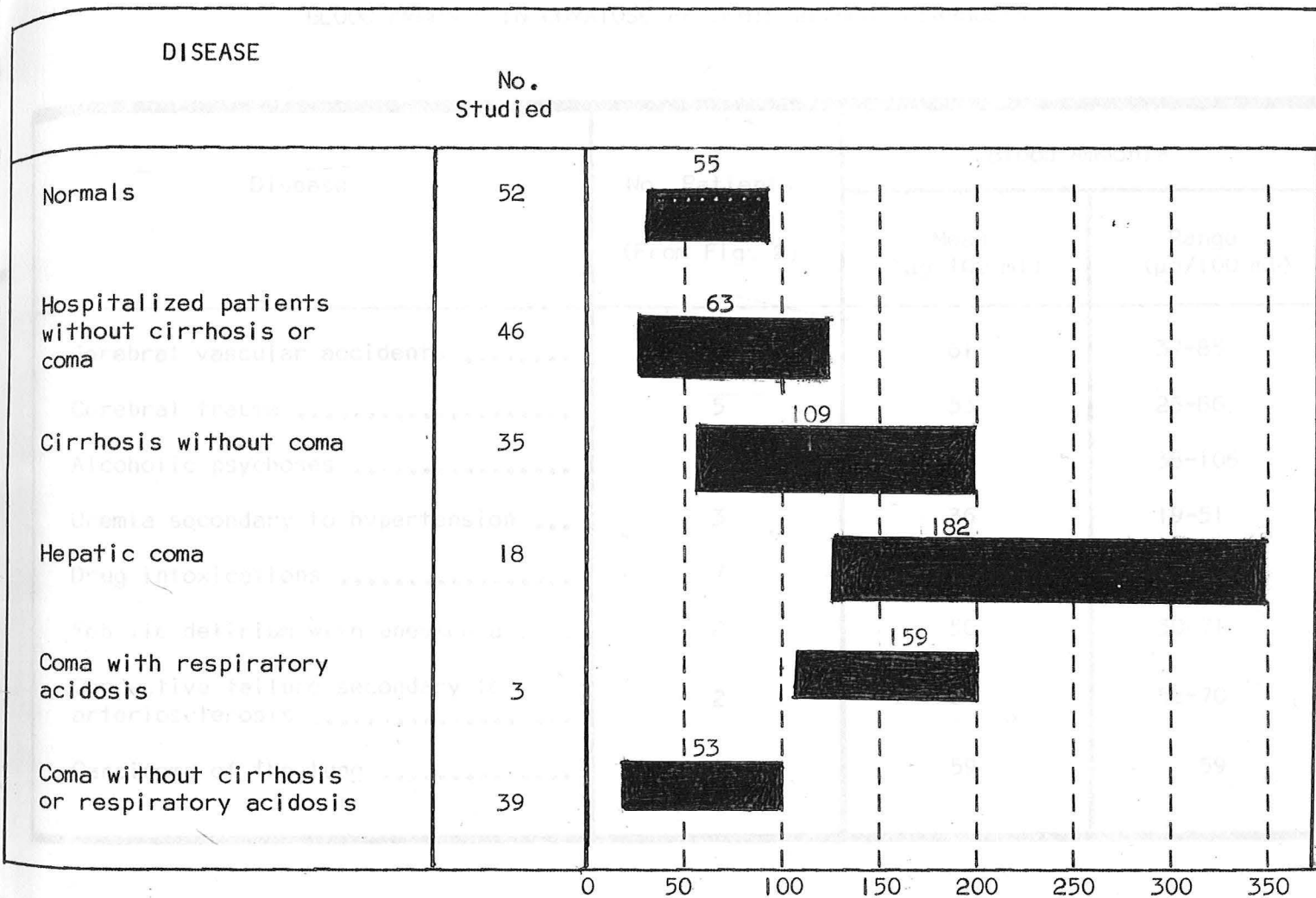


Fig. 2. Range and mean of blood ammonia levels. (O) normal range (mg/dl) (solid values.)

Figure 2 (16)

BLOOD AMMONIA IN $\mu\text{g}/100\text{ ml}$ (venous)



The range and mean of blood ammonia levels. (Figures above bars indicate mean values.)

(2) Amino acids: α -ketoglutarate, γ -amino acids, aspartate, phenylalanine, asparagine, glutamine (17,18). Last correlation with α -ketoglutarate state than ammonia. Probably related to partial impaired metabolism in diseased liver.

CSF:

1. α -ketoglutarate, pyruvate (14).
2. γ -glutamine (Table 11) of diagnostic value (11,12,13).
3. Normal or $\uparrow$ protein - no $\uparrow$ in cells or pressure (14).

Table I

BLOOD AMMONIA IN COMATOSE PATIENTS WITHOUT CIRRHOSIS

Disease	No. Patients Studied (From Fig. 2)	Blood Ammonia	
		Mean ($\mu\text{g}/100\text{ ml}$)	Range ($\mu\text{g}/100\text{ ml}$)
Cerebral vascular accidents	11	61	37-85
Cerebral trauma	5	53	26-86
Alcoholic psychoses	8	64	38-106
Uremia secondary to hypertension ...	3	36	19-51
Drug intoxications	7	57	34-79
Febrile delirium with pneumonia	2	50	30-71
Congestive failure secondary to arteriosclerosis	2	64	52-70
Carcinoma of the lung	1	59	59

Abnormality of most significant in precoma. However, although on average the abnormality correlated with the above, there is no link in both directions.

EFG (e.g., in full coma) is not specific for hepatic encephalopathy. The following are also seen in hypoglycemia, diabetic coma, etc. (27).

Electroencephalogram (EEG) is not specific for hepatic encephalopathy.

- (2) Amino acids: α -ketoglutaric, pyruvic, acetoacetic, methionine, tyrosine, phenylalanine, asparagine, glutamine (17,18). Less correlation with neurologic state than ammonia. Probably related in part to impaired metabolism by diseased liver.

CSF:

1. $\uparrow$ ammonia, α -ketoglutarate, pyruvate (14)
2. $\uparrow$ glutamine (Table II) of diagnostic value (17,18,19,25)
3. Normal or $\uparrow$ protein - no $\uparrow$ in cells or pressure (20)

Table II

GLUTAMINE LEVELS IN CSF (19)

	Hepatic Coma	Cirrhosis	Other Liver Pathology	Other Comatose States	Control Cases
No. of cases	26	17	18	21	98
Range (mg%)	23-96	13-30	6-24	6-19	5-22
Mean $\pm$ S.D. (mg%)	52 $\pm$ 17	21.3 $\pm$ 4.2	15 $\pm$ 5.2	11.4 $\pm$ 3.7	10.3 $\pm$ 3.5

b. EEG:

Paroxysms of bilaterally synchronous, symmetrical, high voltage, slow waves at 1 - 7 cycles/second (normally 8 -13/second). Start anteriorly and progress posteriorly (21).

Abnormality of most significance in precoma. However, although on average EEG abnormality correlates with neurologic disorder, there is overlap (in both directions) in individual cases.

EEG (especially in full coma when it is diffusely slow) is not specific for hepatic encephalopathy. Abnormalities seen in anoxia, uremia, hypoglycemia, diabetic coma, etc., (22), - non-specific.

Electronic frequency analysis detects early cases (23,24), especially after administration of protein or small doses (8 mgm) of morphine (24).

Diagnosis

1. Clinical syndrome (present in full or in part).
2. Exclusion of other causes: hypoglycemia, subdural hematoma, etc.
3. Confirmation by response to therapy is helpful.

Pathogenesis

1. Consciousness (wakefulness, awareness, reactivity): Depends on functional integrity of reticular activating system in brain stem. This polysynaptic network (rete) of neurons receives and correlates afferent stimuli from within and without the body (26,27). Cortex apparently is not essential, but may modify consciousness(28). Rigidity, asterixis and hypernea, often present in hepatic encephalopathy, also are believed to depend on structures located in brain stem (29).

2. CNS pathology in hepatic encephalopathy

- a) Early: None
- b) Few days: ↑ number and size of protoplasmic astrocytes in brain gray matter. Probably specific, but may be seen in hypoxia (30). Occasionally - demyelination of pyramidal tracts in spinal cord (5).

Conclusion: Absent or scant anatomic changes and reversibility of clinical picture suggest a metabolic disorder.

3. CNS physiology in hepatic encephalopathy (31,32,33)

See Tables III and IV on pages 9 and 10.

Table III (32)

Cerebral Metabolism in Hepatic Disease

	Mean CBF (ml blood/100 g brain/min)	Mean CMRO ₂ ml O ₂ /100 gm brain/min	Mean A-V O ₂ (Vol. %)
Normal	54.0	3.3	6.0
Alert	47.1	2.3	5.1
Moderate cerebral dysfunction	41.9	1.7	4.4
Coma	39.6 (↓ 26%)	1.6 (↓ 50%)	4.1

CBF = cerebral blood flow

CMRO₂ = cerebral oxygen consumption

A-V O₂ = arterio-venous oxygen extraction

Table IV (33)Cerebral Metabolism in Hepatic Disease

Degree of Neurologic Impairment	CBF* ml/100 g/min	CMRO ₂ * cc/100 g/mm	A-V O ₂ * Vol. %	Pa CO ₂ mm Hg	Art. pH	Art. O ₂ Sat %	PO ₂ mm Hg
Normal (11) [†]	53	3.4	6.6	38	7.42	95.2	89
Grade (6) 0-1	59.0	3.4	5.9	33	7.48	90.0	57
Grade (9) 2	47.0	2.8	6.1	29	7.51	93.0	64
Grade (6) 3	36	2.0	5.6	30	7.55	89.5	55
Grade (3) 4	22	1.6	7.2	25	7.57	92.0	56

* See footnote Table III
[†] Number of subjects

Summary of findings in Tables III and IV:

1. Progressive decrease in cerebral oxygen consumption ($CMRO_2$) with increasing neurologic impairment.
2. Decrease in cerebral blood flow (CBF), possibly, at least partly, a result of drop in $Pa\ CO_2$ (from hyperventilation).
3. Respiratory alkalosis common (Table IV).
4. In general, $CMRO_2$ more depressed than CBF, and cerebral oxygen extraction ($A-V O_2$) not sufficient to compensate for $\downarrow$ CBF.
5. (?) Decreased cerebral glucose uptake (34) (inconstant).

Comment: Cause or effect?

Decreased $CMRO_2$ occurs in other states of decreased consciousness, as uremic coma, diabetic coma, etc., and is not specific for hepatic encephalopathy (35). However, it does not invariably follow states of somnolence - i.e., normal in sleep and pentathol semi-narcosis (35).

Probability: Hepatic coma is result of $\downarrow$ $CMRO_2$, which may, however, be common denominator of other metabolic disorders.

Conclusion: Impaired cerebral oxidative metabolism, possibly associated with faulty glucose utilization.

Postulated causes of disturbed cerebral metabolism in hepatic encephalopathy:

I Toxin

- a) ammonia
- b) methionine and/or other nitrogenous products
- c) tryptophane metabolites (indoles)

in conjunction with

II Sensitivity of brain in liver disease, possibly as result of depletion of vital substances (? cytidine, ? uridine) (36) synthesized by normal liver.

I_a) Ammonia Toxicity

Evidence in favor:

- 1) $\uparrow$ ammonia levels in blood and CSF of patients with hepatic coma.
- 2) Precipitation of syndrome in susceptible patients and animals by administration of ammonia or substances (blood, proteins) giving rise to it.
- 3) Frequent alleviation of syndrome by therapeutic measures which prevent ammonia influx into brain.

Evidence against:

- 1) Lack of precise correlation in individual patients between degree of neurologic impairment and level of blood or CSF ammonia.
- 2) Much higher levels of ammonia (than in patients with coma) are necessary to produce coma in experimental animals and to demonstrate toxic effect ($\downarrow$ O_2 consumption) of ammonia on brain in vitro (37).

Comment - re: Evidence against

Blood and CSF levels do not necessarily reflect intracerebral concentration of ammonia. Blood ammonia measurement is notoriously difficult and subject to error (45). Blood pH is an important variable - at ↑ pH, NH_3 transfer into brain augmented (2).

Animal and in vitro studies are short-term (minutes-hours) experiments and do not take into account presence of a sensitized brain.

Conclusion:

Arguments against ammonia as a cause of hepatic encephalopathy do not seem convincing.

See Table V on page 13

Relevant Mechanisms

Excretion	Excretion of ammonia as urea, allantoin, and other nitrogenous compounds in urine.	Excretion of ammonia as urea, allantoin, and other nitrogenous compounds in urine.
Excretion	Excretion of ammonia as urea, allantoin, and other nitrogenous compounds in urine.	Excretion of ammonia as urea, allantoin, and other nitrogenous compounds in urine.
Excretion	Excretion of ammonia as urea, allantoin, and other nitrogenous compounds in urine.	Excretion of ammonia as urea, allantoin, and other nitrogenous compounds in urine.
Excretion	Excretion of ammonia as urea, allantoin, and other nitrogenous compounds in urine.	Excretion of ammonia as urea, allantoin, and other nitrogenous compounds in urine.
Excretion	Excretion of ammonia as urea, allantoin, and other nitrogenous compounds in urine.	Excretion of ammonia as urea, allantoin, and other nitrogenous compounds in urine.
Excretion	Excretion of ammonia as urea, allantoin, and other nitrogenous compounds in urine.	Excretion of ammonia as urea, allantoin, and other nitrogenous compounds in urine.
Excretion	Excretion of ammonia as urea, allantoin, and other nitrogenous compounds in urine.	Excretion of ammonia as urea, allantoin, and other nitrogenous compounds in urine.
Excretion	Excretion of ammonia as urea, allantoin, and other nitrogenous compounds in urine.	Excretion of ammonia as urea, allantoin, and other nitrogenous compounds in urine.
Excretion	Excretion of ammonia as urea, allantoin, and other nitrogenous compounds in urine.	Excretion of ammonia as urea, allantoin, and other nitrogenous compounds in urine.
Excretion	Excretion of ammonia as urea, allantoin, and other nitrogenous compounds in urine.	Excretion of ammonia as urea, allantoin, and other nitrogenous compounds in urine.

Conclusion: Ammonia ↑ in hepatic encephalopathy due to its increased production (exogenous type) and/or decreased metabolism (endogenous type). Often "mixed" type seen.

Table V

Ammonia Metabolism in Health and Hepatic Encephalopathy

<u>NORMAL METABOLISM</u>		<u>METABOLISM IN HEPATIC FAILURE</u>
<u>Sources</u>		
<u>Gut:</u>	Effect of bacterial and (?) digestive enzymes on proteins and urea.	↑ NH ₃ from a) intestinal bleeding b) ↑ BUN c) high protein diet
<u>Kidney:</u>	Deamination of glutamine and amino acids with release of NH ₃ into renal veins.	↑ NH ₃ - from diuretics (chlorothiazide, Diamox) which alkalinize urine and may cause hypokalemia
<u>Muscle:</u>	Release on exercise-(?) from adenylic acid.	↑ NH ₃ - released from muscle saturated with ammonia-(?) mechanism.
<u>Removal Mechanisms</u>		
<u>Liver:</u>	a) urea synthesis (Appendix I) b) glutamine synthesis (See Fig. 3)	NH ₃ - removal impaired probably due to decreased urea and glutamine synthesis by diseased liver and ammonia shunted around liver by portosystemic shunts.
<u>Lung:</u> (minor)	Exhalation	May be ↑ with hyperventilation and ↑ blood levels of NH ₃ .
<u>Muscle:</u> (liver)	a) amination of keto acids α - ketoglutaric acid ↓ NH ₃ glutamic acid (ATP) ↓ NH ₃ glutamine b) carbamyl phosphate synthesis (Appendix I)	May be impaired since ketoacids ↑ in blood

Conclusion: Ammonia ↑ in hepatic encephalopathy due to its increased production (exogenous type) and/or decreased metabolism (endogenous type). Often "mixed" type seen.

Importance of pH for ammonia metabolism (38,39,40,41,42,43,44,45)

Ammonia in body fluids exists as NH_4^+ and NH_3 , and proportion of each component is dependent primarily, on pH, as shown below for plasma at 37°C.

$$\text{pH} = \text{pKa} + 10 \log \frac{\text{NH}_3}{\text{NH}_4^+} \quad (\sim 2\%)$$

(7.40) (8.90)

Current methods estimate total ammonia ($\text{NH}_4^+ + \text{NH}_3$) but only NH_3 freely crosses tissue membranes. NH_4^+ diffuses very slowly, probably because of its water solubility and charge.

With $\uparrow$ blood pH, the proportion of NH_3 in blood rises, resulting in a larger quantity of gaseous ammonia which can enter into the brain. The NH_3 distributes itself between the 2 compartments on basis of pH gradient, tending (as a weak base) to move from alkaline to acid side, i.e., from blood to relatively more acid intracerebral milieu ($\text{pH} \sim 7.0$). Any $\uparrow$ in blood pH (without equally altering cerebral pH in some direction) will augment transfer of NH_3 into brain. (This assumes the same pKa for ammonia in both sites).

Thus, metabolic alkalosis will increase plasma NH_3 and will enhance transfer of ammonia into more acid intracellular sites (brain). This explains why alkalosis (viz. diuretic (Diuril) induced) may be detrimental in hepatic encephalopathy.

Evidence of this is abundant from animal experiments where wide pH shifts can be employed (39,40,41,42,43) and less evident (but still convincing) in human studies where lesser pH changes are tolerated (44,45,46).

Respiratory alkalosis in hepatic encephalopathy (33) - (See Table IV)

Etiology unknown.

Theories:

- a) Stimulation of peripheral lung reflexes: - no evidence available
- b) Effect
 - 1) Due to $\uparrow$ ammonia: - conflicting data (47,48)
 - 2) Compensatory for intracerebral accumulation of acid metabolites: - no evidence available.

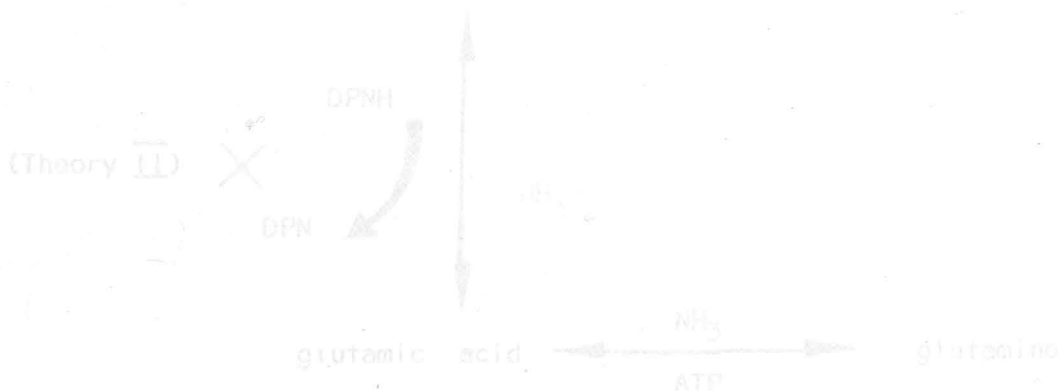


Figure 3

MECHANISM(S) OF AMMONIA "TOXICITY"

PYRUVIC ACID

(CO₂ fix)

(Theory III)

Oxaloacetic acid

Citric acid

KREBS
CYCLE

ATP Formed

(Theory I)

α - Ketoglutarate

DPNH

NH₃

DPN

glutamic acid

NH₃

ATP

glutamine

(Theory II)

X

← NH₃ →

ATP

Theory I Cerebral α -ketoglutarate depletion (Bessman) (49)

Principle: Increased brain ammonia is detoxified to glutamine with resultant depletion of α -ketoglutarate which cannot be replenished from blood since it does not readily cross the "blood-brain barrier". Decrease in α -ketoglutarate slows Krebs cycle and this together with consumption of ATP to form glutamine depletes brain ATP. Such a depletion in strategic sites may interfere with oxidative brain metabolism and/or acetylcholine synthesis (54) which are ATP-dependent.

Evidence for:

- 1) $\downarrow$ α -ketoglutarate (35-50%) and $\uparrow$ glutamine and $\uparrow$ pyruvate in brain of ammonia-intoxicated (short-term) animals (49,50,51).
- 2) Methionine sulfoximine (inhibits synthesis of glutamine) in vivo prevents $\uparrow$ glutamine and pyruvate and $\downarrow$ α -ketoglutarate in ammonia-intoxicated animals. Despite $\uparrow$ brain NH_3 - its toxicity is markedly $\downarrow$ (51).
- 3) Acute NH_3 intoxication does not $\downarrow$ cerebral cortical ATP (52) but does $\downarrow$ (25% $p = < .001$) ATP in medulla and pons (53).

Evidence against:

- 1) Cerebral transaminations can replenish α -ketoglutarate if glutamate adequate and transaminases active. Since glutamine is believed derived from a small metabolically active pool of glutamate (55) and the size of the 2 pools has not been separately assessed, this is undertermined.
- 2) One report (34), α -ketoglutarate released into cerebral venous blood in patients with hepatic coma. However, (?) methodology since pyruvic and lactic acids which accumulate in brain were not released (all these traverse from brain to blood with great difficulty) and glutamine which does also accumulate, but crosses into blood easier, also was not released. In addition, total α -ketoglutarate measurement also may not be helpful, since this keto acid in brain also is functionally compartmented (55).
- 3) CO_2 fixation (Fig. 3) occurs in brain and may replenish Krebs cycle intermediates (α -ketoglutarate). This fixation seems to $\uparrow$ in cerebral cortex under acute ammonia intoxication (55,56).

Pertinent questions re: 3)

- a) Is increase sufficient to provide energy?
- b) Does it take place in reticular formation?
- c) Does it occur in chronic state?

Answers: Unknown

- Conclusion: Theories I and II, perhaps in combination seem most probable.
- 4) It has been calculated that 1% of cerebral ATP is sufficient for acetylcholine synthesis in brain (57), so that part of above hypothesis is unlikely. Interestingly, methionine sulfoximine (which protects versus NH_3 intoxication) prevents an ammonia-induced $\downarrow$ in cerebral acetylcholine (58).

Theory II Decreased supply of DPNH for ATP generation in mitochondria as result of competition for DPNH by α -ketoglutarate $\xrightarrow[\text{NH}_3]{\text{DPNH} \longrightarrow \text{DPN}}$ glutamate reaction.
(Worcel and Ercinska) (61)

Principle: DPNH oxidation necessary for generation of ATP by mitochondrial electron chain.

Evidence for:

- 1) In vitro data shows that NH_3 inhibits O_2 uptake by mitochondria and this is proportional to formation of glutamate from α -ketoglutarate. This cannot be prevented by addition of excess α -ketoglutarate so depletion of this ketoacid is not causal; succinate as substrate (which does not depend on DPNH) is not affected by NH_3 ; addition of DPNH to α -ketoglutarate prevents toxic ammonia effect.*
- 2) Results in vivo with methionine sulfoximine (51) and studies on brain ATP (53) could go along with this theory.

Theory III Impaired oxidative decarboxylation of pyruvic acid. (McKhenn and Tower) (59)

Principle: Pyruvic (and possibly α -ketoglutaric) acid's entry into Krebs cycle via citric acid is impaired - with slowing of cycle.

Evidence for:

- 1) In vitro data shows that mitochondrial O_2 consumption is impaired to a degree as pyruvate utilization was decreased. This was not remedied by adding succinate, DPN. Mechanism of action unknown.

Evidence against:

- 1) Contradicted by another study utilizing similar methods (61).
- 2) Not supported (as sole cause) by in vivo data with methionine sulfoximine.
- 3) Authors neglect own data on α -ketoglutarate that support Theory II.

Theory IV Direct toxic effect of ammonia on neurons. (Weil-Malherb) (60)

Principle: Interference of NH_3 with ionic flux across neuronal membranes.

No data to support this.

In vivo methionine sulfoximine data (51) suggest that intracerebral ammonia increase per se cannot explain the whole effect of NH_3 .

Conclusion: Theories I and II, perhaps in combination seem most tenable.

* Glutamine reverses the effect of ammonia by preventing amination of α -ketoglutarate. The level of TPNH has also been shown to be depressed (70%) on addition of ammonia (104).

I d) Methionine and Other Amine Toxicity

Evidence in favor:

- 1) Methionine, its metabolites and other amines may be elevated in blood, urine, and CSF of patients with hepatic coma (62).
- 2) Administration of methionine may precipitate coma in susceptible patients (63).
- 3) Methyl mercaptan (by-product of methionine) induces unconsciousness (5 mg/per l. inspired air) in rats (64).

Evidence against:

- 1) Methionine I.V. is not toxic in patients in whom coma is precipitated on oral administration (64).
- 2) Rx with oral antibiotics prevents above toxicity.
- 3) Portal vein ammonia ↑ after giving methionine (65).
- 4) Monamine oxidase inhibitors prevent rise in blood ammonia after infusion of ammonium citrate, suggesting that amines (derived from aminated ketoacids) may be source of ammonia (66).

Conclusion: These data suggest that methionine and its products, as well as other amines may precipitate hepatic encephalopathy but a likely mechanism is via release of ammonia. No other specific mechanisms have been postulated.

I c) Tryptophan metabolites (indoles) toxicity (64)

Theory a) Depletion of cerebral serotonin, due to inability of liver to convey tryptophan to 5-OH tryptophan, the precursor of serotonin:

Evidence in favor:

- 1) Serotonin is present in high concentration in brain stem and may be concerned with normal and abnormal brain function (67).
- 2) 5-OH indole acetic acid (5HIAA) (end product of serotonin) may be decreased in urine of patients with severe liver disease (67).
- 3) Administration of 5-OH tryptophan I.V. to patients with hepatic coma, but not other types of coma, improved their EEG (68).

Evidence against:

- 1) Importance of brain serotonin as related to signs of hepatic coma is debatable (67).
- 2) Lowered urinary excretion of 5-HIAA in liver disease not confirmed (69).

Theory b) Tryptophan is converted by gut bacteria to indole and skatole. Small amounts of these enter the circulation and in presence of liver disease and/or collateral circulation may not be conjugated as sulfates and glucuronides for urinary excretion and thus may accumulate in blood as toxins.

Evidence in favor:

- 1) Indoles and skatoles in vitro at high concentrations inhibited respiration of brain (37).

Evidence against:

- 1) Such high concentrations of these substances not found in man (37).

Conclusion: Not enough evidence to assess the role of tryptophan metabolites in human hepatic encephalopathy.

II ↑ Sensitivity of the brain in hepatic disease

Evidence in favor:

- 1) Normal individuals with high blood ammonia levels (maintained by I.V. infusion) were asymptomatic. Note, however, that these are brief infusions (hours) (70). In presence of alkalosis, ammonia in large doses was toxic in "normal" individuals (71).
- 2) Patients with liver disease and especially those with encephalopathy are very sensitive to hypoxia, CO₂, sedatives, etc., - all of which exacerbate neurologic impairment (9).
- 3) Isolated perfused cat brain loses its electrical excitability in absence of liver in perfusion circuit. The liver can be substituted for by inclusion of cytidine and uridine (36).
- 4) Respiration of brain from animals with chronic liver disease is more depressed by toxins (ammonia) in vitro (37).

Conclusion: Evidence strong that liver disease may sensitize brain to various toxins.

Summary on Pathogenesis of Hepatic Encephalopathy

1. Metabolic cerebral disorder with decreased oxygen consumption.
2. Occurs in individuals, with liver disease (parenchymal and/or circulatory hepatic impairment), whose brain is sensitive to a variety of "toxins" which often summate in their deleterious effect.
3. "Toxins" very likely include ammonia and other nitrogenous products. Hypoxia, electrolyte disturbances, alkalosis, etc., - often potentiate this effect.
4. Precise site of action of "toxins" is not known but appears to involve Krebs cycle, probably at level of α -ketoglutarate.

TABLE VII

Cause

Therapy

- I. Standard - of established value.
 1. Removal of precipitating causes and treatment of their effect.
 2. Stop influx of nitrogenous substances into body (stop or decrease ingested protein, antibiotics, etc.).
 3. Maintain caloric, fluid and electrolyte balance.
- II. Semi-Experimental - of possible or probable value.
 1. Arginine
 2. Glutamate
 3. Removal of colon from continuity of intestinal tract.
 4. Steroids
- III. Experimental - unproven value.
 1. Urease immunity
 2. Hemodialysis - peritoneal dialysis - resin dialysis - cross circulation
 3. Cation exchange resins
 4. Protamine

TABLE VI (72) duration with decreased renal function

- I. Removal of precipitating causes and treatment of their effects.
 6. Incidence:

Provoking Causes of Coma in 167 Cirrhotic Patients with Coma (1958-1962)

Cause	%
1. Gastrointestinal hemorrhage (varices 57%)	37
2. Progressive hepatic failure (no "external" precipitating cause)	19
3. Diuretics (paracentesis 10%)	17
4. Unknown	14
5. Infection	10
6. Uremia	2

TABLE VII

<u>Cause</u>	<u>Mechanism(s) of Coma Precipitation</u>
1. Hemorrhage*:	a) ↑ ammonia and other nitrogenous products (100 ml blood = 15-20 gm protein) b) Decreased hepatic and kidney function, the latter resulting in ↑ BUN → NH₃ c) Ammonia in banked transfused blood (73); storage at 4°C - 1 day = 170 µg% 4 days = 330 µg%; 21 days = 900 µg% d) Shock and hypoxia
3. Diuretics:	a) ↓ [K⁺], alkalosis - resulting in ↑ intra-cellular transfer of NH₃, ↑ ammonia released from kidneys (74,75) b) Paracentesis - may lead to oligemia and ↓ renal function with ↑ BUN and sometimes dilutional hyponatremia c) (?) Unknown effects - possibly on up-take of "toxins" by peripheral tissues.
5. Infection (septicemia, peritonitis): Mechanism unknown	
Possible causes:	a) ↑ tissue catabolism with ↑ endogenous nitrogen load b) Dehydration with decreased renal function c) "Toxicity" - hypoxia (76), hyperthermia (77).
6. Uremia:	a) Effect of disease itself on brain b) ↑ BUN → NH₃
7. Sedatives**: Anesthetics:	a) Effect on brain oxygen consumption b) Hypoxia

Excessive protein ingestion is special case of 1.a

* Morphine and other sedatives absolutely contraindicated in hepatic coma.

For Rx of delirium tremens - promazine drugs (used in judicious doses) probably best.

For Rx of convulsions in hepatic coma - barbitol, phenobarbital, which are excreted largely via kidneys. Reduced dosage employed, and patients Rx titrated by clinical response.

For surgery - minimal anesthesia compatible with procedure.

2. Stop influx of nitrogenous products.

a) In coma stop protein ingestion completely.

In precoma - decrease protein ingestion to ~ 40 gm/day.

b) Antibiotics to decrease gut bacterial flora.

Generally used - non-absorbable antibiotics - neomycin, paramomycin.

Initial Rx of coma - 8 gm/day in divided doses. Subsequently, 3-4 g/day.

In precoma 1-2 gms/day may be adequate (minimal dose not well established.)

At 8 gm/day - Stool E. coli essentially disappears in most patients in 2 days; may see ↑ in yeast, diptheroids, strep fecalis (78,79). Feter hepaticus usually disappears in 3 days, paralleling decrease in blood ammonia (78). Clinical condition of patient usually (but not always) improves in 36-72 hours, but may take a week.

In patients with ileus, neomycin may be given as 1% enema in water. Most ammonia is produced in colon (80) and enemas have resulted in ↓ blood ammonia in dogs and man (81). The reliability and consistency of response to this procedure in man are not definitely established.

Relative value of other blood spectrum antibiotics (tetracyclines, chloramphenical) vs. neomycin is not well established clinically. In dogs they have a lesser effect than neomycin on bacterial flora and suppression of ammonia production (80). There is suggestive evidence that they are also less effective in patients (103). Note that intravenous tetracycline (and other antibiotics excreted in bile) may not be adequately excreted into gut in presence of liver disease (Schenker and Combes, unpublished observations).

Caution:

Neomycin is absorbed slightly (1-3%) and may accumulate in blood (ototoxic, nephrotoxic) in patients with renal failure.

Neomycin may lead to malabsorption syndrome - apparently dose-time dependent and probably due to direct effect of drug on gut mucosa (82).

Neomycin may cause staphylococcal enteritis by suppressing endogenous microbial flora, therefore, with diarrhea - stools must be smeared and stained for diagnosis and immediate treatment.

Glutamine

c) Constipation (retention of gastrointestinal blood) must be prevented by judicious use of enemas and laxatives. Excessive (routine) use of former may precipitate dehydration and augment any disturbance of electrolyte status and the latter may be detrimental in patients with gastrointestinal bleeding.

Comment:

Glutamine is eventually deaminated with release of ammonia. Most studies show no consistent value. Only controlled study showed no benefit using single infusion of 25-50 g/2 hour (67).

3. Colonic Exclusion

Principle:

Removal of site of "toxin" formation.

3. Caloric, fluid and electrolyte balance.

On withholding of protein, patient must receive at least 1600 calories as glucose. After 2-4 days of antibiotic therapy, gradual reinstitution of protein (starting with 25 gm/day) has not been detrimental to patients (83). Some wait until evidence of improvement before starting protein. Protein intake is titrated vs. clinical response. Finally, antibiotics are gradually stopped. Best to vary one factor at a time.

Proof of value of above regimen.

- a) No strictly controlled study. It should definitely be considered in the proper patient, who is not a forbidding surgical risk.
- b) Immediate mortality in patients prior to above therapy = ~ 90% (83). Present immediate mortality in comparable patient population = 40-50% (84).

- c) Prognosis best if precipitated by exogenous factor(s) which can be reversed. Worse with "unprovoked" liver failure, high bilirubin, low $[Na]$ (84).

II. Semi-Experimental Therapy

1. Arginine -

Principle:

Serves as substrate for urea cycle (Appendix) therapy with cortisone 500 mg q 8 h for equivalent doses of prednisone.

Comment:

Not established that there is a hepatic deficiency of arginine. Furthermore, arginase decrease in cirrhotic liver has been demonstrated (85) and this may limit usefulness of arginine.

Variable results obtained in practice. Better in patients who may recover spontaneously and worse in those with poorer prognosis. One controlled study (86) failed to show effect on either clinical state or blood ammonia. However, the dose used may have been too low - one infusion of 25 gm/2 hours. Some claim good results with larger doses.

Principle:

Immunize with jackbean urease, induce antibodies to urease. Arginine may lower blood ammonia when given as HCl salt due to $\downarrow$ pH.

2. Glutamate -

Principle:

- a) Combines with $NH_3 \rightarrow$ glutamine by transamination
- b) Yields α - ketoglutarate which takes up NH_3 .

Comment:

Glutamine is eventually deaminated with release of ammonia. Most studies show no consistent value. Only controlled study showed no benefit using single infusion of 25-50 g/2 hour (87).

3. Colonic Exclusion -

Principle:

Removal of site of "toxin" formation.

Comment:

This approach has been used only for patients who:

a) Do not respond to trials of protein restriction plus antibiotics, on assumption that a sufficient number of gut bacteria are resistant to therapy.

Conclusion:

b) Do not follow prescribed therapy and are in danger of becoming mental cripples.

c) Become protein depleted (↑ ascites) on restricted diet

2. Hemodialysis, exchange dialysis (97) -

Principle:

This approach has on occasion given excellent results (88,89,90). It should definitely be considered in the proper patient, who is not a forbidding surgical risk, prior to development of irreversible neurologic abnormalities.

Comment:

4. Steroids -

Principle:

Unknown. In hepatitis may decrease inflammatory response.

Comment:

Most studies (91,84) show no beneficial effect in cirrhosis but an occasional one (92) claims temporary improvement with massive (1000 mg cortisone/day). None are controlled.

3. Cation exchange resins

Principle:

In viral hepatitis, coma outcome almost always fatal (93). Although no strict control studies available, therapy with cortisone 500 mg q 8 h (or equivalent doses of prednisone, etc.) was associated with recovery (93) in some patients (9/23 = 39%). Improving prothrombin time serves as only good guide to prognosis. Two patients had perforations of ulcer and one bled from intestinal tract - out of 23. Therapy with steroids is now recommended for viral hepatitis, pending further data.

Comment:

III. Experimental

1. Urease immunity (94,95,96)

Principle:

Immunize with jackbean urease, induce antibodies to urease which pass into intestinal content and inhibit bacterial intestinal urease with resultant decrease in ammonia production.

4. Protamine -

Principle:

Comment:

Immunization with urease has given ↑ serum antiurease antibody levels (94) and with use of under 8 units of urease no toxicity. Of 8 patients with recurrent encephalopathy - in 6, tolerance for protein increased markedly (100-300%) without increase in blood ammonia levels (95). In animals there was a 60% ↓ in gastrointestinal urease, - in gastrointestinal ammonia and anti-bodies to urease present in feces.

Conclusion regard

However,

1) Study not well controlled.

2) Three patients developed coma after immunization with 10-20 units - in one this may have hastened death.

- 3) Blood ammonia levels were not decreased in any patient.
- 4) Two of eight patients failed to respond despite high serum antiurease antibodies.
- 5) Antibodies may be rapidly digested in gut (96).

Conclusion:

Needed:

- a) Further control studies
- b) Measurement of antiurease in human gut content

2. Hemodialysis, exchange dialysis (97) -

Principle:

Remove circulating ammonia and other toxins via artificial kidney, resin column, etc.

Comment:

Hemodialysis may effectively remove ammonia (20 mg in 4 hours (98) but procedure is tedious, often rendered dangerous by need for heparinization and difficult to apply to large number of patients. Peritoneal dialysis may be of value, especially with concomitant renal failure, but no control studies are available.

Scattered reports available concerning use of resins and one of parabiosis (99). Although some benefit is claimed - insufficient data is available at present.

3. Cation exchange resins (100-101) -

Principle:

Orally administered exchange resins (exchange Na or K for NH_4) bind NH_4 in gut lumen and thus remove it.

Comment:

Excellent results obtained in dogs given blood p o - substantial decrease in blood ammonia (100). In patients with gastrointestinal bleeding, equally good results obtained (101). Use of resin in K cycle may bind ammonia and replete body K which is often low in cirrhotics. Serum electrolytes and ECG monitor necessary. Resin may be given by enema also.

Dose suggested: 20 grams in 30 ml 70% Sorbital q 6 h p o.

4. Protamine -

Principle:

Basic protein consists of 80% arginine - may slowly release arginine into circulation.

Comment:

In experimental animals, protamine lowers blood ammonia (102). Insufficient data for human studies. In large doses drug is an anti-coagulant.

Conclusion regarding therapy:

Standard therapy indicated under I has proven of value. In special instances therapy indicated under II (semi-experimental) may be beneficial. Other procedures are strictly in investigative stage.

Clinical Syndrome

1. *Gabuzda, G.J. Hepatic Coma. Advances in Internal Medicine, Vol. XI, 1962.
2. Davidson, C.S. Hepatic Coma. Disease-a-Month, Jan., 1964.
3. Sherlock, S. Hepatocellular Failure. Diseases of the Liver, 3rd ed., 1963, pp. 84-103.
4. Davidson, E.A., and Summerskill, W.H.J. Psychiatric aspects of liver disease. Postgrad. Med. J. 32:487, 1956.
5. Zieve, L., Mendelson, D.F., and Goepfert, M. Shunt encephalomyelopathy: II. Occurrence of permanent myelopathy. Ann. Int. Med. 53:53, 1960.
6. Scobie, B.A., and Summerskill, W.H.J. Permanent paraplegia with cirrhosis. Arch. Int. Med. 113:805, 1964.
7. *Adams, R.D., and Foley, J.M. Neurological changes in more common types of severe liver disease. Tr. Am. Neurol. Assn. 74:217, 1949.
8. *Leavitt, S., and Tyler, H.R. Studies in asterixis. Arch. Neurol. 10:360, 1964.
9. Conn, H.W. Asterixis in non-hepatic disorders. Am. J. Med. 29:647, 1960.
10. *Warren, K.S., and Rebouças, G. Blood ammonia during bleeding from esophageal varices in patients with hepatosplenic schistosomiasis. New Eng. J. Med. 271:921, 1964.
11. Foulk, W.T., Butt, H.R., Stauffer, M.H., Baggenstoss, A.H., and Gross, J.B. Hepatic coma: A clinical and pathologic study. Gastroenterol. 29:171, 1955.
12. Challenger, F., and Walshe, J.F. Fetor hepaticus. Lancet 1:1239, 1955.
13. *Phear, E.A., Sherlock, S., and Summerskill, W.H.J. Blood ammonium levels in liver disease and "hepatic coma", Lancet 1:836, 1955.
14. Summerskill, W.H.J., Wolfe, S.J., and Davidson, C.S. The metabolism of ammonia and α - ketoacids in liver disease and hepatic coma. J. Clin. Invest. 36:361, 1957.
15. Lieber, C.S., and Lefevre, A. Metabolism ammoniacal et intermediaire au cours du coma hepatic. Acta Clin. Belgica 13:328, 1958.
16. Sullivan, J.F., Linder, H., Holdener, P., and Ortmeyer, L. Blood ammonia in cerebral dysfunction. Am. J. Med. 30:893, 1961.
17. Walshe, J.M. Glutamic acid in hepatic coma. Lancet 1:1238, 1955.
18. Whitehead, T.P., and Whittaker, S.R.F. A method for the determination of glutamine in CSF and the results in hepatic coma. J. Clin. Path. 8:81, 1955.
19. *Gilon, E., Szeinberg, A., Tauman, G., and Bodonyi, E. Glutamine estimation in cerebrospinal fluid in cases of cirrhosis and hepatic coma. J. Lab. Clin. Med. 53:714, 1959.
20. Amatuzio, D.S., Weber, L.J., and Nesbitt, S. Bilirubin and protein in the cerebrospinal fluid of jaundiced patients with severe liver disease with and without hepatic coma. J. Lab. Clin. Med. 41:615, 1953.

21. *Foley, J.M., Watson, C.W., and Adams, R.D. Significance of electroencephalographic changes in hepatic coma. *Trans. Amer. Neurol. Assn.* 75:161, 1950.
22. Parsons-Smith, B.C., et al., Electroencephalograph in liver disease, *Lancet* 2:867, 1957.
23. Laidlaw, J. The application in general medical conditions of a visual method of assessing and representing generalized EEG abnormalities. *J. Neurol. Neurosurg. Psychiat.* 22:69, 1959.
24. Laidlaw, J., Read, A.E., and Sherlock, S. Morphine tolerance in hepatic cirrhosis. *Gastroenterol.* 40:389, 1961.
25. Steigmann, F., Kazemi, F., Dubin, A., and Kissane, J. Cerebrospinal fluid glutamine in the diagnosis of hepatic coma. *Amer. J. Gastroenterol.* 40:378, 1963.
26. Magoun, H.W. The ascending reticular system and wakefulness. Brain Mechanisms and Consciousness, Chas. C. Thomas, Springfield, 1954.
27. French, J.D., and Magoun, H.W. Effects of chronic lesions in central cephalic brain stem of monkeys. *AMA Arch. Neurol. Psych.* 68:591, 1952.
28. Penfield, W. Studies of the cerebral cortex of man. Brain Mechanisms and Consciousness, Chas. C. Thomas, Springfield, 1954.
29. Ganong, W.F. Medical Physiology, Lange Medical Publication, 1963, pp. 126-145.
30. Greenfield, J.G. in Neuropathology, Greenfield, J.G., Blackwood, W., McMenemey, W.H., Meyer, A., and Norman, R.M. Edward Arnold Pub., London, 1958, p.50.
31. Wechsler, R.L., Crum, W., and Roth, J.L.A. The blood flow and oxygen consumption of the human brain in hepatic coma. *Clin. Res. Proc.* 2:74, 1954.
32. Fazekas, J.F., Ticktin, H.E., Ehrmentraut, W.R., and Alman, R.W. Cerebral metabolism in hepatic insufficiency, *Amer. J. Med.* 21:843, 1956.
33. *Posner, J.B., and Plum, F. The toxic effects of carbon dioxide and acetazolamide in hepatic encephalopathy. *J. Clin. Invest.* 39:1246, 1960.
34. Dastur, D.K., Seshadri, R., and Tatageri, V.R. Liver brain relationship in hepatic coma. *Arch. Int. Med.* 112:899, 1963.
35. Kety, S. Blood flow and metabolism of the human brain in health and disease. Neurochemistry, 2nd Ed. Eds. Elliott, K.A.C., Page, I.H., and Quastel, J.H. Chas. C. Thomas, Springfield, 1962.
36. Geiger, A., and Yamasaki, S. Cytidine and uridine requirement of brain. *J. Neurochem.* 1:93, 1956.
37. Walshe, J.M., De Carli, L., and Davidson, C.S. Some factors influencing cerebral oxidation in relation to hepatic coma. *Clin. Sci.* 17:27, 1958.
38. Milne, M.D., Scribner, B.H., and Crawford, M.A. Non-ionic diffusion and the excretion of weak acids and bases. *Am. J. Med.* 24:709, 1958.
39. Warren, K.S. The differential toxicity of ammonium salts. *J. Clin. Invest.* 37:497, 1958.

40. Warren, K.S., and Nathan, D.G. The passage of ammonia across the blood-brain barrier and its relation to blood pH. *J. Clin. Invest.* 37:1724, 1958.
41. *Stabenau, J.R., Warren, K.S., and Rall, S.P. The role of pH gradient in the distribution of ammonia between blood and cerebrospinal fluid, brain and muscle. *J. Clin. Invest.* 38:373, 1959.
42. Warren, K.S., and Schenker, S. Differential effect of fixed acid and carbon dioxide on ammonia toxicity. *Am. J. Physiol.* 203:903, 1962.
43. *Warren, K.S. Ammonia toxicity and pH. *Nature*, 195:47, 1962.
44. Warren, K.S., Iber, F.L., Hille, W., Sherlock, S. Effect of alterations in blood pH on distribution of ammonia from blood to cerebrospinal fluid in patients in hepatic coma. *J. Lab. Clin. Med.* 56:687, 1960.
45. *Moore, E.W., Strohmeyer, G.W., Chalmers, T.C. Distribution of ammonia across the blood. Cerebrospinal fluid barrier in patients with hepatic failure. *Am. J. Med.* 35:350, 1963.
46. Humoller, F.L., Barak, A.J., and Holthaus, J.M. Distribution of ammonia in cells and plasma. *Clin. Chem.* 10:589, 1964.
47. Roberts, K.E., Thompson, F.G. III, Poppell, J.N., and Vanamee, P. Respiratory alkalosis accompanying ammonium toxicity. *J. Appl. Phys.* 9:367, 1956.
48. Wilson, W.P., and Tyor, M.P. Electroencephalographic observations during experimental hyperammonemia. *Neurol.* 8:913, 1958.
49. *Bessman, S.P. Ammoniogenic coma - the biochemistry of an endogenous intoxication. Biochemistry of the Central Nervous System, Pergamon Press, London, 1958, p. 141.
50. Clark, G.M., Eisenman, B. Studies in ammonia metabolism in biochemical changes in brain tissue of dogs during ammonia-induced coma, *New Eng. J. Med.* 259:178, 1958.
51. Warren, K.S., and Weinbach, E.C. Unpublished data.
52. Schenker, S., Mendelson, J.H. Cerebral adenosine triphosphate in rats with ammonia-induced coma. *Am. J. Physiol.* 206:1173, 1964.
53. Schenker, S., and Brophy, E. Unpublished data.
54. Ulshafer, T.R. The measurement of changes in acetylcholine level in rat brain following ammonium ion intoxication and its possible bearing on the problem of hepatic coma. *J. Lab. Clin. Med.* 52:718, 1958.
55. *Berl, S., Takagaki, G., Clarke, A.D., and Waelsch, H. Metabolic compartments in vivo. *J. Biol. Chem.* 237:2562, 1962.
56. Waelsch, H., Berl, S., Rossi, C.A., Clarke, D.D., and Purpura, D.P. Quantitative aspects of CO₂ fraction in mammalian brain in vivo. *J. Neurochem.* 11:717, 1964.
57. McIlwain, H. Biochemistry and the Central Nervous System. J. & A. Churchill, Ltd., London, 1959, pp. 119-211.
58. Braganca, B.M., Faulkerer, P., and Quastel, J.H. Effects of inhibitors of glutamine synthesis on the inhibition of acetylcholine synthesis in brain slices by ammonium ions. *Biochem. and Biophys. Acta* 10:83, 1953.

- 59.* McKhann, G.M., and Tower, D.B. Ammonia toxicity and cerebral oxidative metabolism. *Am. J. Physiol.* 200:420, 1961.
60. Weil-Malherbe, H. The metabolism of ammonia in brain in consciousness and the chemical environment of the brain. 25th Ross Fed. Resch. Conf. 1958, p. 50.
- 61.* Worcel, A., and Erecinska, M. Mechanism of inhibitory action of ammonia on the respiration of rat liver mitochondria. *Biochim. et Biophys. Acta*, 65:27, 1962.
62. Wu, C., Bollman, J.J., and Batt, H.R. Changes in free amino acids in plasma during hepatic coma. *J. Clin. Invest.* 34:845, 1955.
63. Phear, E.A., et al., Methionine toxicity in liver disease and its prevention by chlortetracycline. *Clin. Sci.* 15:93, 1956.
64. Walshe, J.M. Observations on the symptomatology and pathogenesis of hepatic coma. *Quart. J. Med.* 20:421, 1951.
65. Webster, L.T. Jr., and Gabuzda, G.J. Effect on portal blood ammonium concentration of administering methionine to patients with hepatic cirrhosis. *J. Lab. Clin. Med.* 50:426, 1957.
66. Zuidema, G.D., Kirsch, U.M., Cares, H., Kowalchuk, R.S., and Coon, W.W. Effect of monoamine oxidase inhibitor in experimental ammonia intoxication. *Ann. of Surg.* 158:363, 1963.
- 67.* Fazekas, J.P., Taupin, H., and Alman, R.W. Physiologic and pharmacologic basis for chemotherapy of psychiatric states. *Am. J. Med.* 21:825, 1956.
68. Borges, F.J., Merlis, J.K., Bessman, S.P. Serotonin metabolism in liver disease. *J. Clin. Invest.* 38:715, 1959.
69. Donaldson, R.M. Jr., Arabehty, J., and Gray, S.J. In vivo studies of 5-hydroxyindole metabolism in patients with hepatic cirrhosis and in rats. *J. Clin. Invest.* 38:933, 1959.
70. Tyor, M.P., and Wilson, W.P. Peripheral biochemical changes associated with the intravenous administration of ammonium salts in normal subjects. *J. Lab. Clin. Med.* 51:592, 1952.
71. Stauffer, J.C., and Scribner, B.H. Ammonia intoxication during treatment of alkalosis in a patient with normal liver function. *Am. J. Med.* 23:996, 1957.
72. Labby, D.H., Hutchinson, J.H., Stahl, J., Bochel, R., and Imler, M. Mechanism of hepatic coma. Biochemical Clinics - The Liver, No. 3, 1964. R.H. Donnelley Co., New York, N.Y.
73. Spear, P.W., Sass, M., and Cincotti, J.J. Ammonia levels in transfused blood. *J. Lab. Clin. Med.* 48:702, 1956.
74. Casey, T.H., Summerskill, W.H.J., Bickford, R.G., and Rosevear, J.V. Body and serum potassium in liver disease. II. Relationships to arterial ammonia, blood pH and hepatic coma. *Gastroenterol.* 48:208, 1965.
75. Owen, E.E., Flanagan, J.F., and Tyor, M.P. Kidney as a source of blood ammonia: Effect of chlorthiazide. *Proc. Soc. Exp. Biol. Med.* 102:696, 1959.

16. Warren, K.S., and Schenker, S. Hypoxia and ammonia toxicity. *Am. J. Physiol.* 199:1105, 1960.
17. Schenker, S., and Warren, K.S. Effect of temperature on toxicity and metabolism of ammonia in mice. *J. Lab. Clin. Med.* 60:291, 1962.
18. Fisher, C.J., and Faloon, W.W. Blood ammonia levels in hepatic cirrhosis: their control by oral administration of neomycin. *New Eng. J. Med.* 256:1030, 1957.
19. Dawson, A.M., McLaren, G., and Sherlock, S. Neomycin in the treatment of hepatic coma. *Lancet* 2:1263, 1957.
20. Silen, W., Harper, H.A., Mawdsley, D.L., and Weirich, W.L. Effect of antibacterial agents on ammonia production within the intestine. *Proc. Soc. Exp. Biol. & Med.* 88:138, 1955.
21. Najarian, J.S., et al. Control of ammonia production in the colon with neomycin enemas. *AMA Arch. of Surg.* 78:844, 1959.
22. Jacobson, E.D., Prior, J.T., and Faloon, W.W. Malabsorptive syndrome induced by neomycin: Morphologic alterations in the jejunal mucosa. *J. Lab. Clin. Med.* 56:245, 1960.
23. Stormont, J.M., Mackie, J.E., and Davidson, C.S. Observations on antibiotics in the treatment of hepatic coma and on factors contributing to prognosis. *New Eng. J. Med.* 259:1145, 1958.
24. Summerskill, W.H.J., Wolfe, S.J., and Davidson, C.S. The management of hepatic coma in relation to protein withdrawal and certain specific measures. *Am. J. Med.* 33:59, 1957.
25. Ugarte, G., Pino, M.E., and Valenzuela, J. Liver arginase activity in patients with liver cirrhosis and in patients in endogenous hepatic coma. *J. Lab. Clin. Med.* 57:359, 1961.
26. Reynolds, T.B., Redeker, A.G., and Davis, P. A controlled study of the effects of L arginine on hepatic encephalopathy. *Am. J. Med.* 25:359, 1958.
27. Alexander, R.W., Berman, E., and Balfour, D.C. Relationship of glutamic acid and blood ammonia to hepatic coma. *Gastroenterol.* 29:711, 1955.
28. Atkinson, M., and Goligher, J.C. Recurrent hepatic coma treated by colectomy and ileorectal anastomosis. *Lancet* 1:461, 1960.
29. Dienst, S.F. Treatment of progressive ammonia intoxication by exclusion of the colon. *New Eng. J. Med.* 270:555, 1964.
30. Editorial - Coma and the Colon, *Lancet* 1:310, 1963.
31. Sklar, M., and Young, I.I. Failure of ACTH and adrenal corticoids to alter the course of hepatic coma in advanced portal cirrhosis. *Am. J. Med. Sci.* 229:138, 1955.
32. Pessar, T., and Hersing, J.W. Massive doses of cortisone in hepatic coma. *Ann. Int. Med.* 48:1254, 1958.
33. Katz, R., et al. Corticosteroids in the treatment of acute hepatitis in coma. *Gastroenterol.* 42:258, 1962.

94. Visek, W.J., Iwert, M.E., and Burrows, W. Detection of antibody to urease by hemagglutination. *Proc. Soc. Exp. Biol. & Med.* 109:54, 1962.
95. Thomson, A., and Visek, W.J. Some effects of induction of urease immunity in patients with hepatic insufficiency. *Am. J. Med.* 35:804, 1963.
96. Ingelfinger, F., in Yearbook of Medicine, 1964-1965, p. 617-619.
97. Ritchie, H.E., et al. Extra corporeal methods of reducing high blood ammonia levels. *Gut* 3:172, 1962.
98. Sherlock, S. Hepatocellular Failure. Disease of the Liver, 3rd Ed., 1963, pp. 137-138.
99. Nealon, T.F. Jr., and Ching, N.P.H. Ion exchange resins in hepatic coma. Biochemical Clinics - The Liver, Vol. 3, R.H. Donnelley and Co., New York, N.Y., 1964.
100. Zuidema, G.D., Fletcher, M.M., and Burton, W.D. Orally administered cation exchange resin-sorbital regimen for treatment of ammonia intoxication in dogs. *Proc. Soc. Exp. Biol. & Med.* 109:732, 1962.
101. Zuidema, G.D., et al. Oral cation exchange resins for treatment of ammonia intoxication. *S.G. and O.* 119:790, 1964.
102. Brown, H., and Brown, G. Lowering blood ammonia with protamine. *Arch. Surg.* 85:771, 1962.
103. Summerskill, W.H.J. Hepatic coma in liver failure and gastrointestinal hemorrhage treated with neomycin. *Brit. Med. J.* 11:1322, 1958.
104. Erecinska, M., and Worcel, A. Reversal of the inhibitory action of ammonia on the respiration of rat-liver mitochondria. *Biochim. and Biophys. Acta.* 71:305, 1963.

Urea Cycle Enzymes

- (1) carbamyl phosphate synthetase
- (2) ornithine transcarbamylase
- (3) arginine synthetase (condensing and cleavage)
- (4) arginase

11/17/58
 3 134-0020-12

UREA SYNTHESIS

Case #1. The patient, a 3-year-old Negro female, was first seen January 15, 1958, at Children's Medical Center in Dallas. The chief complaint was pain in the right hip for 6 weeks and pain in the left knee for 2 weeks with progressive weakness and lassitude. Physical examination showed decreased rotary motion of both hips. X-ray films of hips, knees, shoulders, hands, and chest taken during this period showed no calcification.

Roentgenograms taken in May 1958 showed extensive calcification of the soft tissue around the knees. By September this had increased in the legs and was also present in the hands and arms. A biopsy of the skin and muscle of the left lower leg showed predominantly acanthosis in the subcutaneous tissue. There was lobulated deposition of calcium varying from fine granular to grossly visible deposits with osteoclasts surrounding the calcium deposits. The muscle showed focal areas of nonspecific degeneration but no inflammatory reaction.

Administration of acetate was begun in November, 1959. The range of motion performed before and after therapy was unchanged. Post-therapy roentgenograms showed no loss of tissue calcium. Subjective improvement was noted, and patient appeared to be more active after therapy. Repeat roentgenograms in May 1960, showed a slight increase in the calcium deposits in the hips.

In November, 1960, she had an episode of acute pain and inability to move the left leg, with extreme tenderness just above the popliteal space on the left. Flexion deformities of the knees and recurrent episodes of pain in the area of calcific deposits above the popliteal space bilaterally led to operation on March 13, 1961, hamstring release and removal of extensive calcific deposits in the muscles and tendons.

Urea Cycle Enzymes

- (1) carbamyl phosphate synthetase
- (2) ornithine transcarbamylase
- (3) arginine synthetase (condensing and cleavage)
- (4) arginase

Case #2. The patient, a 14-year-old white female, developed low-grade generalized weakness at age 8. She was seen at Texas Scottish Rite Hospital March 12, 1958, and found to have "weakness in the arms and thighs." A skin and muscle biopsy from the left lower leg showed extensive areas of calcification in fibrofatty tissues and heavy inflammatory-cell infiltration around these areas. No changes were noted in the muscle tissue. Radiographic examination revealed calcification of the entire right anterior tibial muscle and tendon, and diffuse calcification in the subcutaneous tissue of the left lower leg and about the hip muscles.

