

ANCA-Associated Vasculitis

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Introduction

The systemic vasculitides are a group of rare and life-threatening diseases characterized by inflammation of blood vessel walls and adjacent tissues.¹⁻⁴ The end result of this inflammatory response is necrosis, stenosis, and thrombosis of vascular channels leading to ischemia, infarction or hemorrhage in affected organs. These disorders should be considered in any patient with multisystem organ dysfunction or persistent constitutional complaints. Some of the primary vasculitides are associated with the presence of anti-neutrophil cytoplasmic antibodies (ANCA).⁵⁻¹⁰ These disorders include Wegener's granulomatosis (WG), microscopic polyangiitis (MPA), and the Churg-Strauss syndrome (CSS).¹¹⁻¹⁸ They occur at a rate of approximately 1-3 per 100,000, with CSS being the least common.¹⁹⁻²⁵ While each of these conditions has been associated with characteristic "patterns" of organ involvement, and particular laboratory and histopathologic findings, there is actually considerable overlap between them.^{6, 18, 26-29} In some cases, it is impossible to tell them apart. They are all idiopathic, pauci-immune forms of predominantly small vessel vasculitis (mainly affecting arterioles, capillaries and venules) that have the potential to target any organ in the body.^{3, 30, 31} They are best considered "variations on a theme" of a single disease entity, ANCA-associated systemic vasculitis (AASV).^{32, 33}

Clinical Features

Wegener's Granulomatosis

Patients with AASV often present with constitutional symptoms including fever, weight loss, malaise, fatigue, arthralgias and myalgias.³⁴⁻³⁶ WG additionally features involvement of the upper and lower respiratory tract, as well as the kidneys.^{14, 34, 37, 38} Sinonasal disease presenting as chronic nasal congestion, recurrent sinus infections or epistaxis is common at presentation, and ultimately develops in 90%.^{34, 37, 39, 40} Examination may reveal nasal crusting, ulceration, septal perforation and, occasionally, saddle nose deformity. Other important head and neck manifestations include serous otitis, sensorineural hearing loss and ocular disease.^{39, 41-44} Ocular abnormalities have been identified in 15-50% of patients during the course of the disease.^{34, 37, 44-47} Findings range from proptosis caused by retroorbital inflammatory pseudotumors, to scleritis, uveitis, optic neuritis or occlusion of the central retinal vessels. Oral ulcers and gingivitis may also be seen.

Pulmonary disease is identified in 65-85% of patients with WG.^{34, 37, 48} Nodules, masses or alveolar infiltrates, often cavitating, are the most common radiographic abnormalities (figure 1).^{34, 37, 40, 49, 50} Computed tomography is more sensitive than plain chest radiography for detecting abnormalities in these patients.^{49, 51, 52} Cough, pleurisy or hemoptysis may be seen, but such symptoms are absent in one third.³⁴ Pleural effusions occur in 20%.^{40, 49-51, 53}

The most immediately life-threatening complication of AASV is diffuse alveolar hemorrhage (DAH).⁵⁴⁻⁵⁷ This occurs in 5-10% of patients with WG and carries a 25% mortality risk.^{34, 58-60} Pulmonary hemorrhage typically produces dyspnea, hemoptysis and bilateral airspace disease (ground glass opacities or frank consolidation).^{54, 56, 61, 62} The diagnosis is confirmed by bronchoalveolar lavage demonstrating increasingly bloody return upon instillation of serial saline aliquots.^{63, 64}

Granulomatous inflammation and scarring may lead to subglottic or endobronchial stenosis.⁶⁵⁻⁶⁷ Subglottic stenosis occurs in 15% of WG patients and may produce dyspnea, hoarseness, or no symptoms at all.^{68, 69} Management usually involves mechanical dilatation with corticosteroid injection, or sometimes tracheostomy.⁶⁹

Cutaneous involvement is seen in up to half of patients during the course of their disease.^{34, 37, 41, 70, 71} Palpable purpura, associated with leukocytoclastic vasculitis, is the most

common lesion.⁷² Digital ischemia, skin nodules, pyoderma gangrenosum-like lesions and even livido reticularis may also be seen in patients with AASV.⁷¹⁻⁷³

Colonic or small bowel ulceration may occur due to mesenteric vasculitis, and lead to abdominal pain, diarrhea, gastrointestinal bleeding or perforation.⁷⁴⁻⁸⁰ Clinically significant gastrointestinal involvement is seen in fewer than 15% of WG patients, although more have evidence of disease at autopsy.

Neurologic complications develop in 20-50% of patients with WG, usually later in the disease course.^{34, 37, 40, 81-84} Peripheral neuropathies, particularly mononeuritis multiplex, and cranial neuropathies are most common, but stroke, seizure, cerebritis and meningitis can be seen.

Renal involvement in WG is usually asymptomatic until severe impairment in glomerular filtration occurs.^{34, 37, 85} Proteinuria, hematuria and red blood cell casts characteristic of glomerulonephritis (GN) are seen on urinalysis, along with elevations in blood urea nitrogen and serum creatinine. Although it ultimately develops in between 50 and 90% of WG patients, only about 20% have glomerulonephritis at the time of presentation.^{33, 34, 37, 48} Consequently, only one patient in five will have the classic triad of WG when they first come to medical attention. This, and a lack of physician familiarity with other systemic disease manifestations, may be an important factor contributing to delays in diagnosis and treatment initiation.

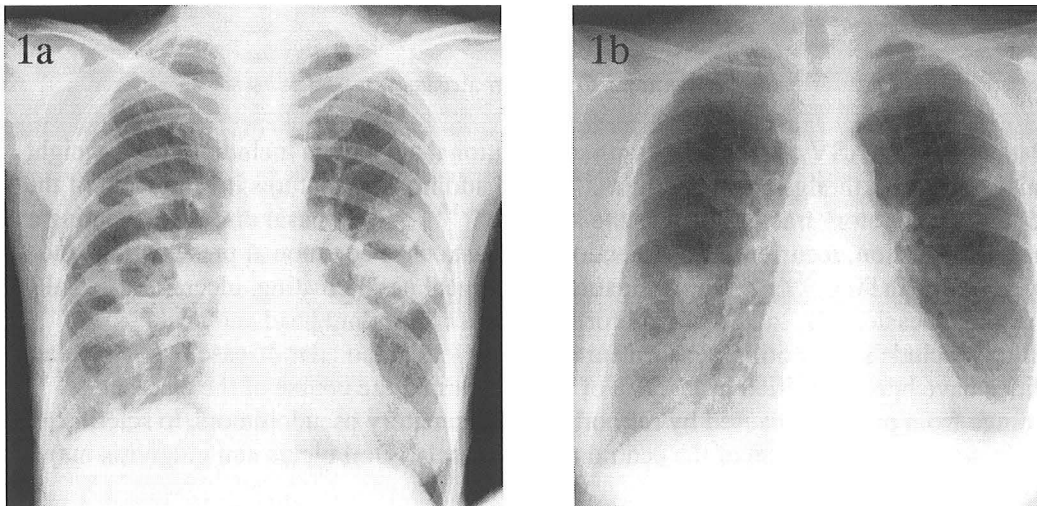


Figure 1. Typical plain film patterns in two subjects with Wegener's granulomatosis. Figure 1a demonstrates multiple, bilateral cavitating masses in a patient with chronic sinusitis, cough and hemoptysis. Figure 1b shows bilateral nodular opacities and a right lower lobe alveolar infiltrate. This patient had a history of epistaxis and dry cough, and presented with acute renal failure.

Microscopic Polyangiitis

MPA is one of the most common small vessel vasculitides.^{13, 86} All of the clinical complications of vasculitis noted above can also occur in microscopic polyangiitis, with the exception of cavitating lung opacities and the granulomatous upper airway disease.³⁰ Relative to WG, renal involvement is reported even more frequently (80-100%), and is more often evident at initial presentation.^{3, 13, 87-89} Microscopic hematuria and/or proteinuria are almost always present, even in the few patients with a normal serum creatinine level.^{3, 87} Diffuse alveolar hemorrhage is usually an ANCA-associated condition, and MPA is a more frequent cause than WG.^{88, 90-92} Between 10% and 40% of MPA patients develop this complication (figure 2).^{13, 87, 93, 94} Constitutional complaints are more frequent compared to WG, and may be present for several

months before a diagnosis is secured.^{13, 35, 87, 95} About one half of MPA patients have skin disease, typically palpable purpura.^{13, 87, 95} Ocular pathology, sinusitis, epistaxis and oral ulcers occur, but less frequently than in WG.^{11, 13, 92} The destructive upper airway disease characteristic of WG does not occur, however. Abdominal pain and gastrointestinal bleeding are twice as common in MPA as in WG.^{87, 96} The reported frequency of central and peripheral nervous system disease in MPA is quite variable.^{13, 87, 95-97}

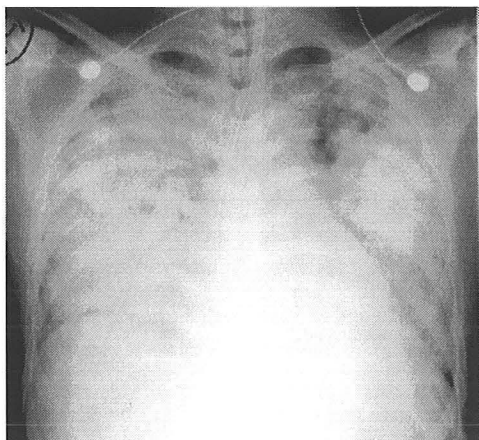


Figure 2. Microscopic polyangiitis. Diffuse alveolar hemorrhage presenting as hemoptysis and acute respiratory failure in a 24 year old man with necrotizing, crescentic glomerulonephritis. The patient was ANCA positive with a perinuclear pattern. Anti-MPO antibodies were also present. The plain film reveals extensive, bilateral airspace disease that cleared completely within two weeks following pulse corticosteroid treatment and plasma exchange.

Churg Strauss Syndrome

Churg Strauss syndrome is diagnosed when asthma and prominent eosinophilia are seen along with a necrotizing vasculitis.^{16, 38, 98-100} Asthma, which is uncommon in other vasculitic syndromes, is almost universally present here.^{98, 101-103} The organs most commonly involved in CSS include the lungs, skin, nervous system, heart, GI tract and kidneys (table 1).^{99, 100, 104} Chest films often reveal patchy alveolar infiltrates at the time of presentation, and computed tomography can detect additional more subtle findings such as ground glass opacities.^{101, 105-109} As in chronic eosinophilic pneumonia, symmetrical peripheral alveolar consolidation resembling the “photographic negative of pulmonary edema” may be seen (figure 3a). Nodular opacities may also be identified, but unlike WG, they rarely cavitate. Diffuse alveolar hemorrhage occurs in CSS, but much less commonly than in MPA or WG.⁹⁹ Thirty percent have pleural involvement.¹¹⁰ Enlargement of the cardiac silhouette is not uncommon (figure 3b).

Table 1. Approximate frequency of various organ involvement in patients with ANCA-associated small vessel vasculitis.

| | WG | MPA | CSS |
|-------------------------|-----|-----|-----|
| Eye/ENT | 90% | 30% | 65% |
| Pulmonary | 75% | 30% | 60% |
| Renal | 80% | 95% | 35% |
| Cutaneous | 50% | 50% | 65% |
| Gastrointestinal | 15% | 30% | 50% |
| Nervous system | 35% | 35% | 70% |
| Cardiac | <5% | <5% | 40% |
| Musculoskeletal | 35% | 65% | 50% |
| Constitutional symptoms | 40% | 70% | 70% |

In many individuals, chronic rhinosinusitis and nasal polyposis are present for years prior to the onset of asthma.^{89, 98} Patients often transition from this prodromal phase to one with simple

tissue eosinophilia that may involve any organ.¹¹¹ Asthma becomes increasingly difficult to control.¹⁶ Marked elevations in serum IgE are common.^{98, 103} Finally, the vasculitic phase appears, usually coincident with the development of constitutional symptoms.¹¹² Paradoxically, the asthmatic component may enjoy significant remission during this phase of the disease, only to return after disease remission is achieved.^{89, 99}

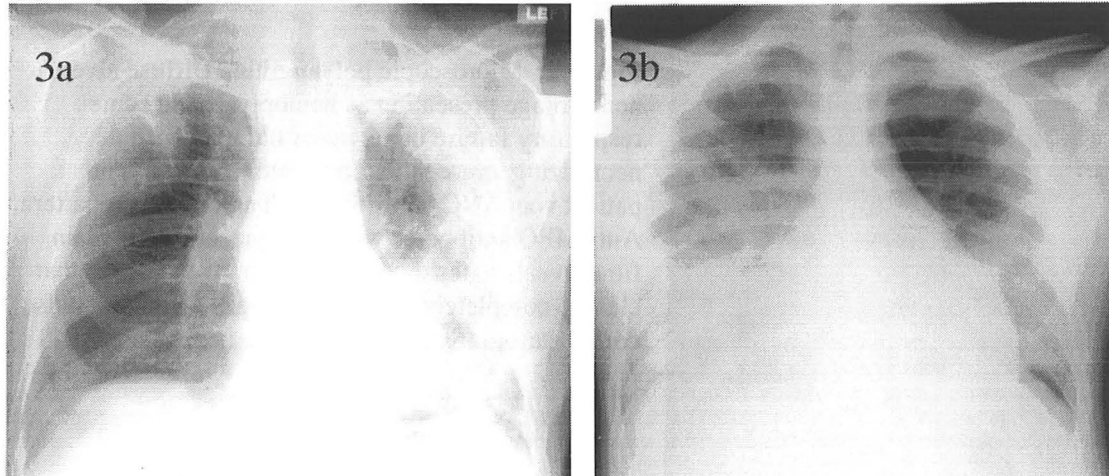


Figure 3. Plain chest radiographs in two patients with Churg Strauss syndrome. 3a) peripheral, axillary predominant consolidation, a finding classic for chronic eosinophilic pneumonia, that is sometimes also seen in CSS. 3b) chest film of a patient with a two year history of asthma and long-standing allergic rhinitis with nasal polyposis. He presented with dyspnea, orthopnea, abdominal pain and marked eosinophilia. He had a pericardial effusion, an ejection fraction of 35%, mononeuritis multiplex, and bilateral, basilar predominant airspace disease with eosinophilic pneumonia (but no vasculitis) by surgical lung biopsy.

Palpable purpura or subcutaneous nodules are seen in two thirds.^{73, 98, 113, 114} Gastrointestinal involvement ranging from an eosinophilic gastroenteritis to mesenteric ischemia, and producing abdominal pain, bleeding, ulceration or perforation, is seen in about 50% of patients at some point in the disease course.^{98, 101, 115-119} Peripheral neuropathies, usually mononeuritis multiplex, develop in 65-75%.^{89, 98, 120, 121} CSS is the form of vasculitis most strongly associated with neurologic impairment. Central nervous system involvement leading to psychiatric disturbances, hemorrhage, infarction or seizures has also been reported in up to 25%.^{82, 98, 120, 122} Cerebral bleeding was responsible for 16% of deaths in the series by Lanham and colleagues.⁹⁸ Involvement of the heart is also common in CSS, and is the leading cause of disease-related death.^{98, 102, 123, 124} It may manifest as congestive heart failure from eosinophilic myocarditis, pericardial effusion or myocardial infarction due to coronary vasculitis.^{98, 125-127} Although renal disease is less frequent, less severe and easier to treat in CSS relative to the other AASVs, it is hardly inconsequential.^{98, 102, 128} Renal abnormalities including proteinuria, hematuria, or renal insufficiency were identified in 84% of 19 patients by Clutterbuck and colleagues.¹²⁹ Four of these patients had a serum creatinine level above 6 mg/dl, and three had nephrotic syndrome. In Lanham's series, renal failure was responsible for 18% of CSS deaths.⁹⁸

Anti-Neutrophil Cytoplasmic Antibodies (ANCA)

ANCA are autoantibodies that react with a variety of proteins within the cytoplasmic granules of neutrophils and monocytes.¹³⁰ While strongly associated with small vessel vasculitis,

ANCA may be seen in a number of other conditions that may appear on the differential diagnosis of AASV.¹³¹ These include collagen vascular disease, certain infections, inflammatory bowel disease, some fibrotic lung conditions and lymphoproliferative disorders.^{5, 132-136} A consensus statement released in 1999 recommended ANCA testing for patients with glomerulonephritis, alveolar hemorrhage, cutaneous vasculitis, chronic sinusitis or otitis, retro-orbital mass, unexplained peripheral neuropathy, or subglottic stenosis.¹³² A positive ANCA with one or more of these clinical features indicates a greater than 90% probability of a pauciimmune small vessel vasculitis.¹³⁷

ANCA are detected by indirect immunofluorescence (IIF) staining of alcohol fixed neutrophils exposed to patient sera. Two major patterns may be identified: perinuclear (p-ANCA) and cytoplasmic (c-ANCA). IIF alone lacks specificity, so ELISA assays are performed on positive sera to confirm the antigen involved.¹³⁸ In AASV, that antigen is almost always proteinase 3 (PR3) or myeloperoxidase (MPO). c-ANCA staining is usually associated with anti-PR3 antibodies and are present in 65-90% of Wegener's granulomatosis patients.¹³⁹⁻¹⁴² p-ANCA are classically associated with anti-MPO antibodies and MPA or CSS.^{3, 4, 10, 17, 99, 143} Approximately 70% of CSS patients and 80% of MPA patients have positive ANCA assays.^{115, 144} Occasionally, patients with MPO antibodies show a cytoplasmic IIF pattern, and patients with PR3 antibodies demonstrate a perinuclear pattern.^{145, 146} One large study was done to assess ANCA sensitivity and specificity in WG and MPA.¹⁴⁴ The sensitivity of the various assays in WG patients was: c-ANCA 64%, p-ANCA 21%, anti-PR3 66%, and anti-MPO 24%. In MPA, the sensitivities were: p-ANCA 58%, c-ANCA 23%, anti-MPO 58%, and anti-PR3 26%. The specificity of these assays for idiopathic small vessel vasculitis, in general, were: c-ANCA 95%, p-ANCA 81%, anti-PR3 87%, and anti-MPO 91%. The diagnostic specificity of the c-ANCA/anti-PR3 and p-ANCA/anti-MPO combinations was 99%.¹⁴⁴

ANCA titers are often high at presentation and during disease exacerbations, and low or absent when the vasculitis is quiescent.^{141, 142, 147} Antibody levels typically decline with treatment, and an increase in ANCA titer often precedes clinical relapse.^{8, 132, 148-150} In a meta-analysis by Tervaert and colleagues, however, clinical relapse occurred in only 94 out of 197 patients with rising ANCA titers; and 157 disease flares were preceded by a rising ANCA titer in only 81 instances.¹⁵¹ This suggests about a 50% sensitivity and specificity in the relationship between changing antibody levels and disease activity. A rising c-ANCA titer accompanied by a rising anti-PR3 titer may indicate a higher probability of disease flare in WG, but the two do not necessarily occur simultaneously.⁸⁸ Stegeman and co-workers also found that anti-PR3 and anti-MPO levels were useful for predicting relapse in AASV.¹⁵² Patients with MPO-ANCA more frequently have persistent antibody elevations despite clinical remission.^{88, 150, 153} It does appear, however, that those with persistently high titers are more prone to disease relapse.¹⁵⁴

Some clinical differences have been noted between patients with PR3-ANCA and those with MPO-ANCA.¹⁵⁵ Patients with anti-PR3 antibodies have more eye, ear and upper respiratory tract involvement, granuloma formation, and a higher relapse rate.^{9, 156} Patients with PR3-ANCA also tend to have more active renal lesions and experience more rapid declines in renal function compared to MPO-ANCA patients, but the long-term renal outcome is essentially the same.^{9, 157}

Histopathology and the Need for Biopsy in AASV

Whether the presence of ANCA in certain clinical settings can replace biopsy as a diagnostic tool is debatable. Some suggest that the presence of MPO- or PR3-ANCA with glomerulonephritis abrogates the need for biopsy confirmation.¹⁵⁸ Others believe that the treatments employed in these disorders are too toxic to be used without first obtaining tissue, and that other useful prognostic information can be gleaned at least from examination of renal tissue.¹⁷ When certain characteristic patterns of organ involvement are combined with the

presence of anti-MPO or PR3 antibodies, the diagnosis of AASV is virtually assured. A more specific diagnosis may require examination of biopsy material, but that information may have little impact on the ultimate approach to treatment.

Many times, even examination of biopsy material only reveals “supportive” evidence that still must be interpreted in the light of other clinical and serologic parameters. For example, the histopathologic hallmarks of WG are vasculitis (involving the small and medium sized arteries), granulomatous inflammation and tissue necrosis (fig 4a). Renal biopsies in WG patients, however, typically reveal only a segmental, necrotizing and crescentic glomerulonephritis with no direct evidence of vasculitis or granulomas (figure 4b).⁵⁵ Immunohistochemistry reveals few, if any, immune deposits (pauci-immune GN), but this does not distinguish WG from MPA or CSS. Renal biopsies are primarily useful for excluding immune complex-mediated GN, anti-basement membrane antibody disease and other causes of GN, and for projecting long-term renal outcome (based on the percentage of glomerular crescents). Upper airway biopsies are easy to perform, but reveal the “diagnostic” combination of granulomas and vasculitis in only one fifth of patients.¹⁵⁹ Surgical lung biopsies have the best overall diagnostic yield, but are harder to obtain.^{55, 160} In patients presenting with diffuse alveolar hemorrhage, pulmonary capillaritis may be seen without any specific findings (i.e., granulomas) to suggest WG.^{161, 162} Capillaritis is characterized by neutrophilic infiltration of alveolar septae with fibrinoid necrosis of alveolar walls leading to the extravasation of erythrocytes into alveolar spaces (figure 4c).^{57, 58, 163, 164} Some of the neutrophils are apoptotic with pyknotic nuclei, and nuclear dust is often seen.^{57, 58} This lesion is the pulmonary equivalent of the leukocytoclastic vasculitis in the skin that produces palpable purpura, and the glomerular capillaritis that leads to necrotizing, crescentic GN.^{30, 162, 165} It may be seen in any of the ANCA-associated vasculitides, and in a variety of other autoimmune conditions as well.^{58, 166-168}

MPA is not associated with granulomatous inflammation, but the histopathology is otherwise indistinguishable from WG.^{11, 30, 87} MPA mainly involves small vessels, but can also involve medium sized arteries. Capillaritis and rapidly progressive necrotizing, crescentic GN are common (figure 4b and 4c).^{87, 160, 169} In some cases, a more slowly progressive glomerulosclerosis may be seen.^{88, 157, 170, 171} Skin, sural nerve and kidney biopsies are the most commonly employed biopsy sites because they are accessible.¹¹

In CSS, the classic histopathologic findings include a necrotizing vasculitis involving small and medium sized vessels, granulomatous inflammation, and eosinophilic infiltration of blood vessel walls and tissues (figure 4d).^{160, 172, 173} It is not essential, however, to see all of these features to make the diagnosis. The original patients described by Churg and Strauss had severe, systemic vasculitis.¹⁷² More subtle presentations are now prevalent due to the common use of exogenous corticosteroids in problematic asthmatics.¹¹¹ In a retrospective study of 23 CSS patients, Reid and colleagues found that only four patients demonstrated all of the classic histologic findings of allergic angiitis and granulomatosis.¹⁷⁴ Eosinophilic infiltration of any organ should raise suspicion for CSS even in the absence of other findings.¹¹¹ Renal disease can be due to a pauci-immune GN or interstitial nephritis.^{99, 128, 129, 175} In the appropriate clinical setting, a positive ANCA with MPO (or occasionally PR3) specificity is good evidence for CSS even when vasculitis is not identified on biopsy.^{15, 111}

Diffuse alveolar hemorrhage occurs rarely in CSS, but subclinical alveolar bleeding may be very common in all of the ANCA-associated vasculitides.^{55, 99, 176-179} Hemosiderin-laden macrophages (indicating prior leakage of red blood cells into the alveolar spaces) were identified in bronchoalveolar lavage fluid from 53% of WG and CSS patients in one study, compared to only a small percentage of patients with collagen vascular disease.¹⁷⁹ At autopsy, capillaritis has been reported in 40% of MPA patients and 31% of WG patients.^{55, 177}

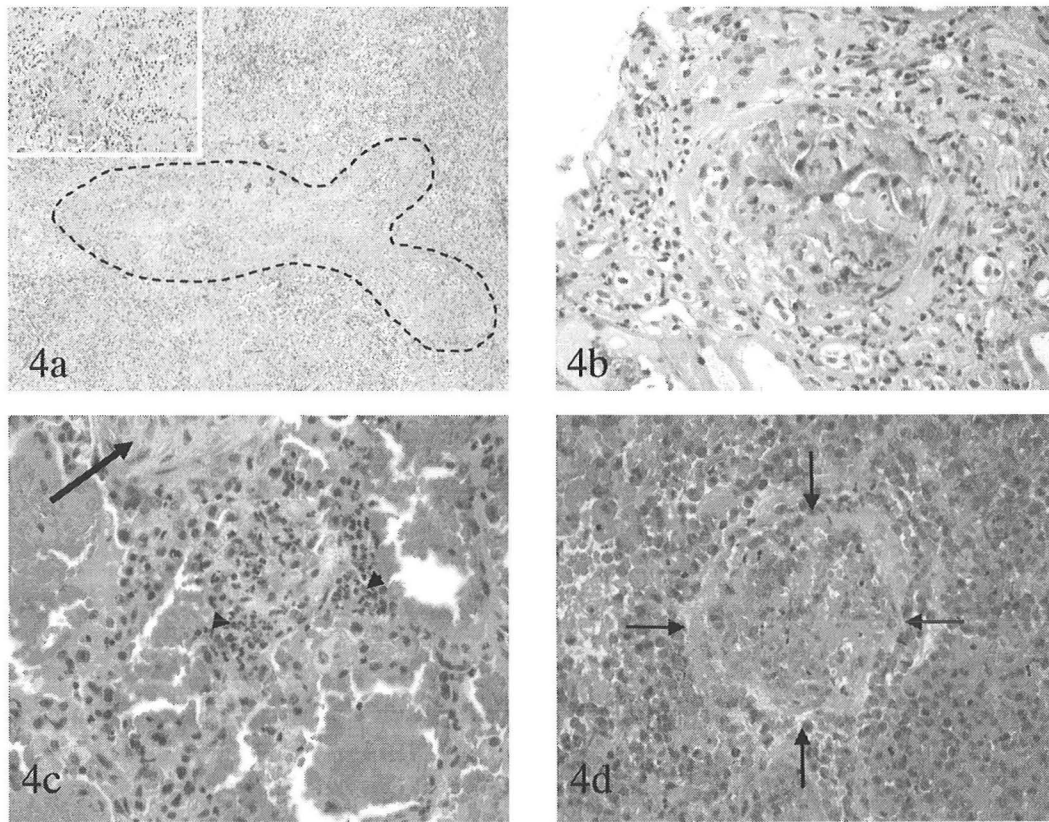


Figure 4. Histopathology of ANCA-associated vasculitis. 4a) necrotizing granulomatous vasculitis in a lung biopsy from a patient with WG. The dotted line outlines the vessel. There is infiltration of the vessel by a poorly formed granuloma (inset). Necrosis is visible in the upper right field. 4b) necrotizing and crescentic glomerulonephritis. 4c) lung biopsy from a patient with microscopic polyangiitis and diffuse alveolar hemorrhage. The specimen shows capillaritis with collections of neutrophils and nuclear debris within the alveolar septae (arrowheads). The adjacent airspaces are filled with blood. A focus of organizing pneumonia is also visible (arrow). 4d) Necrotic small vessel (arrows) in a patient with CSS. The vessel is infiltrated by numerous eosinophils. The adjacent alveolar spaces are filled with blood and eosinophils.

Pathogenesis of AASV

Anti-neutrophil cytoplasmic antibodies participate directly in the pathogenesis of small vessel vasculitis, which is illustrated in figure 5.^{162, 180-183} In order to be fully activated by ANCA, the various effector cells in AASV (neutrophils, monocytes, lymphocytes, eosinophils and endothelial cells) must first be primed by proinflammatory cytokines. Studies have shown that circulating IL-6, IL-8 and TNF- α levels are increased in patients with active disease, but not during remission.^{130, 184, 185} TNF- α and IL-1 levels are high at sites of active vascular inflammation as well.¹⁸⁶ It has been suggested that infection may play a role in the development of vasculitis simply by increasing TNF- α levels and facilitating leukocyte priming.^{7, 180} ANCA, themselves, have also been shown to stimulate the production of proinflammatory cytokines like IL-1, IL-8, monocyte chemoattractant protein-1 (MCP-1) and leukotrienes.¹⁸⁷⁻¹⁹¹ ANCA-binding activates cytokine-primed neutrophils causing a respiratory burst and the release of lysosomal enzymes.³⁵ Neutrophils must first adhere to endothelium, however, before ANCA can fully

activate them.¹⁹² ANCA facilitate this process as well by upregulating adhesion molecule expression ($\beta 2$ integrins, ICAM-1, VCAM-1).^{7, 193-197}

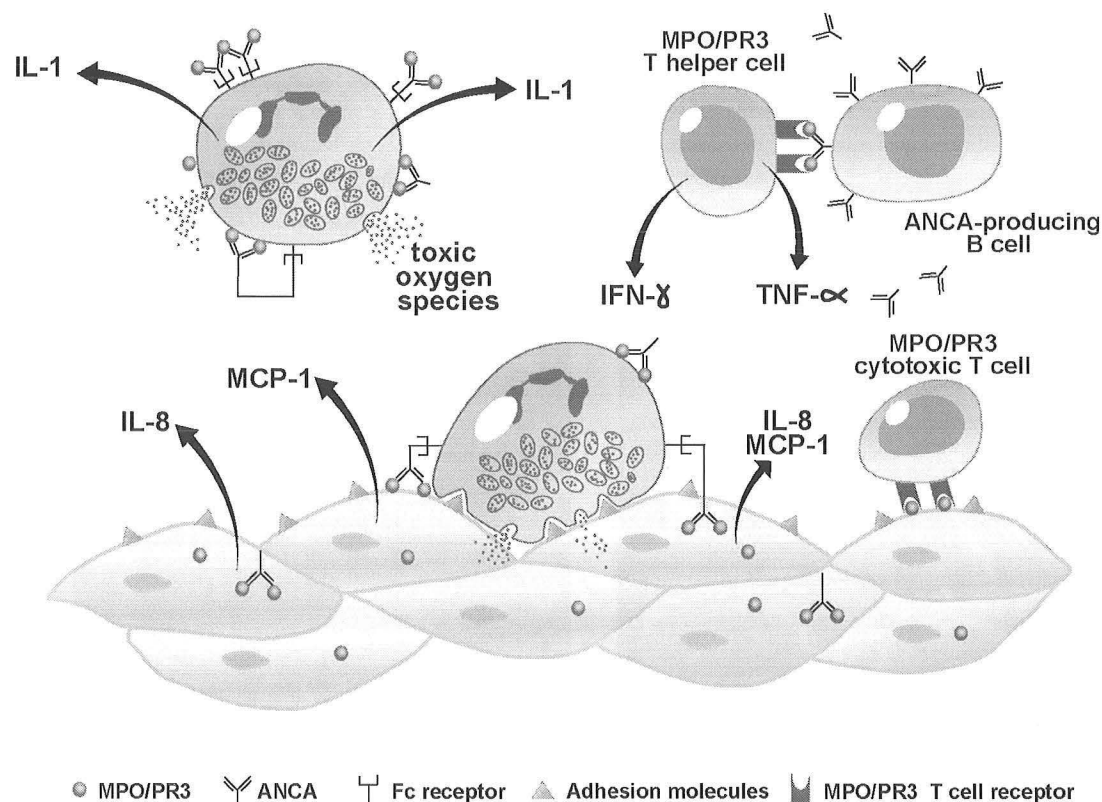


Figure 5. Schematic of major cellular and humoral events contributing to the pathogenesis of AASV. ANCA bind to, and activate, neutrophils and endothelial cells, augmenting the respiratory burst in PMNs, and leading to the elaboration of pro-inflammatory cytokines like IL-1, IL-8 and MCP-1. PR3 binding to endothelium has also been shown, *in vitro*, to elicit IL-8 and MCP-1 production, while also increasing endothelial apoptosis. ANCA promote antibody-dependent cellular cytotoxicity in the presence of neutrophils, contributing to endothelial injury. ANCA-specific T helper cells assist with B cell production of autoantibodies, and release additional cytokines that promote neutrophil and monocyte activation. Cytotoxic T cells with specificity for ANCA target antigens also contribute to vascular injury.

After neutrophils degranulate, cytoplasmic antigens including MPO and PR3 can bind to the neutrophil membrane. When ANCA interact with these proteins, the degree of cellular activation, adhesion molecule expression and oxidant liberation increase.^{196, 198-200} In the presence of cytokines, endothelial cells may also express PR3 on their cell surface.²⁰¹ Free cationic PR3 and MPO can also bind directly to the anionic endothelium after release by degranulation. ANCA binding then promotes antibody-dependent cellular cytotoxicity in the presence of neutrophils.²⁰²⁻²⁰⁷

ANCA also trigger signal transduction pathways within primed endothelial cells that lead to impairment of endothelial barrier function.²⁰⁸ IL-8 inhibits the transendothelial migration of neutrophils, which may result in additional bystander injury to the vascular surface.^{162, 209} Injured and activated endothelial cells release even more proinflammatory substances that attract T cells and monocytes, leading to further damage and, in some cases, granuloma formation.¹⁶²

Several lines of evidence in animal models have confirmed the pathogenicity of ANCA. When MPO antibodies are induced in mice, the animals develop necrotizing glomerulonephritis.^{210, 211} Even in knockout mice that have no functioning lymphocytes, the infusion of anti-MPO antibodies produces a pauci-immune necrotizing, crescentic GN.^{180, 212} When rats are immunized with human MPO, they develop cross-reactive MPO antibodies leading to pulmonary hemorrhage and necrotizing, crescentic GN.²¹³

Apoptosis of neutrophils is associated with translocation of the normally sheltered PR3 and MPO antigens to the cell surface, but the ability of ANCA to induce neutrophil activation of these cells is actually decreased.^{214, 215} Still, apoptosis may play a key role in the pathogenesis of AASV. Primed neutrophils exposed to ANCA undergo dysregulated apoptosis.¹⁶² These cells fail to express an important consumption signal for scavenging phagocytes. Not only do more cells undergo programmed cell death, but they are cleared less efficiently as evidenced by the frequent appearance of cells with pyknotic nuclei and nuclear dust in biopsy specimens of capillaritis or leukocytoclastic vasculitis.⁵⁸ Their persistence in tissues may lead to continued inflammation or the actual induction of autoimmunity.^{162, 215-220} In humans, dendritic cells, but not macrophages, have been shown to efficiently present antigen derived from apoptotic cells, stimulating a cytotoxic T cell response.²²¹ This may even be true for self-antigens. The clonal expansion of self-reactive T helper cells may facilitate the production of ANCA by B cells in this setting. In fact, when Brown Norway rats are injected with syngeneic apoptotic neutrophils (but not fresh neutrophils), they develop ANCA.²²² Interestingly, once ANCA are present, they further amplify neutrophil apoptosis, possibly creating a positive feedback loop and contributing to the development of AASV.^{215, 223}

Under normal circumstances, there are few ANCA target molecules present on the cell surface of neutrophils or other cells. It appears, however, that most of the biological effects of ANCA occur via Fc receptor (FcR) dependent mechanisms.^{191, 224-227} Certain FcR alleles, when engaged by ANCA, trigger a more intense oxidative burst,²²⁵ and certain alleles have been associated with a greater risk of developing WG, and an increased rate of disease relapse.²²⁸

The role of anti-endothelial cell antibodies (AECA) in small vessel vasculitis is less clear. These antibodies have been reported to occur frequently in this setting by some authors, and uncommonly by others.²²⁹⁻²³¹ Under some circumstances, AECA may exert a direct cytotoxic effect on endothelial cells.^{229, 232} AECA have also been shown to increase adhesion molecule expression and proinflammatory cytokine production by endothelial cells.²³³ In CSS, AECA are found in patients with active and inactive disease.²³⁴ They could simply represent an epiphenomenon related to endothelial cell damage.

The neutrophilic proteases targeted by ANCA are not merely degradative enzymes. They participate in complex biological functions including intracellular signaling.²³⁵ PR3, for example, is linked to downregulation of myeloid differentiation. This process of feedback control may be defective in patients with ANCA, and contribute to excessive inflammation.²³⁶ PR3, itself, has been shown to augment IL-8 and MCP-1 production by endothelial cells *in vitro*.²³⁷⁻²³⁹ PR3 can also induce apoptosis in endothelial cells, leading to further vascular damage.^{235, 240, 241} It is also found in normal lung, and is upregulated in alveolar tissue of patients with WG.²⁴² Within an individual, PR3 may be present on the cell membrane of some neutrophils, but absent on others. The size of the membrane PR3+ subset can range from 0-100% of the cells, but is constant in each person.²⁴³ Patients with a high membrane PR3+ phenotype may be more likely to develop AASV.²⁴³⁻²⁴⁵ Certain PR3 (or MPO) polymorphisms may also result in enhanced immunogenicity contributing to more autoantibody production.²³⁶

PR3 activity in normally limited by serum anti-proteases like α 1-antitrypsin.²⁴⁶ For MPO, normal scavenging appears to involve ceruloplasmin.²⁴⁷ Interestingly, allelic alterations associated with α 1-antitrypsin deficiency have been linked to the development of AASV.^{246, 248-254} Patients with the Z phenotype have more widespread organ involvement and higher mortality.^{249,}

^{254, 255} It also appears that ANCA, themselves, may block the normal inhibition of PR3 by α 1-antitrypsin, allowing PR3 to incite vascular injury.^{256, 257}

T lymphocytes are certainly important participants in the granulomatous response of WG and CSS, and in the glomerular lesions of rapidly progressive GN.^{162, 258-260} There is evidence of excessive Th1 lymphocyte activity in WG. T cells from these patients elaborate increased amounts of TNF- α and IFN- γ .^{261, 262} Serum markers of T cell activation have been shown to correlate with disease activity in WG.^{234, 258, 263, 264} Autoreactive T cell populations specific for PR3 and MPO have been identified in patients with AASV.²⁶⁵⁻²⁶⁸ These cells may assist B lymphocytes with the production of ANCA, or participate in direct cell-mediated cytotoxicity.²⁶⁹ The adoptive transfer of anti-MPO specific lymphocytes in MPO-knockout mice immunized with MPO has been shown to cause glomerulonephritis and systemic necrotizing vasculitis (including pulmonary capillaritis).²¹² While T cell responses (exogenous and autoimmune) are normally closely linked to HLA molecule expression, no consistent HLA associations have yet been identified in AASV.²⁶⁹

The major effector cells in CSS are eosinophils, lymphocytes and endothelial cells.^{234, 270} Derangements in CD95 mediated apoptosis of eosinophils and lymphocytes, and oligoclonal T cell expansion has been identified in CSS.²⁷¹ Eosinophil products like eosinophilic cationic protein and major basic protein are important contributors to tissue damage in CSS. These proteins are cytotoxic, can activate complement and coagulation cascades, trigger the release of proinflammatory cytokines and upregulate adhesion molecule expression.¹⁵

There is increasing evidence that dysfunctional co-stimulatory pathways may also play a role in the pathogenesis of vasculitis.²⁷² Cytotoxic T lymphocyte associated antigen-4 (CTLA-4) is expressed on activated T cells and downregulates inflammation after binding to the antigen presenting cell.²⁷²⁻²⁷⁴ Animals that are deficient in CTLA-4 develop fatal lymphoproliferation and vasculitis.^{275, 276} Certain CTLA-4 polymorphisms have been associated with WG, and could contribute to the development of vasculitis by allowing for uncontrolled T cell activation by antigen.²⁷⁷

Cell adhesion molecules play an essential role in the interaction between circulating leukocytes and vascular endothelium. They permit attachment of white blood cells, provide co-stimulatory activation signals, and direct the migration of inflammatory cells into tissues.^{197, 278} Adhesion molecule expression is clearly elevated in patients with vasculitis.²⁷⁸⁻²⁸³ Polymorphisms in CD18, a leukocyte adhesion molecule, have also been associated with MPO-ANCA positive vasculitis.^{284, 285}

Treatment of AASV

There is no doubt that combination therapy with cytotoxic agents and corticosteroids markedly reduces mortality and helps reclaim organ function in patients with AASV.²⁸⁶ Unfortunately, it does so at a considerable physiologic price. Patients survive longer, but suffer from high rates of disease and treatment-related morbidity.²⁸⁶ Once almost uniformly fatal diseases, the ANCA-associated vasculitides have been transformed into chronic, often relapsing disorders that may require several years of therapy.²⁸⁷⁻²⁸⁹ The focus of initial therapy, of course, is the rapid induction of disease remission. The intensity of treatment, however, should be tailored to the severity of the presentation. Once the disease is controlled, a less toxic maintenance regimen can be introduced.^{33, 94, 288} Unfortunately, a substantial proportion of patients will relapse after immunosuppression is tapered or discontinued, regardless of what drug regimen is initially employed.^{13, 34, 99, 290, 291} Even in WG, where relapse rates are highest, the traditional NIH regimen based on daily oral cyclophosphamide (2 mg/kg/day maintained for one year after disease remission) can no longer be recommended.²⁹² That aggressive treatment results in complete disease remission in 75% to 90% of patients, but as many as one half will still relapse, and major

side effects of therapy are seen in over 40%.^{34, 40} Treatment-related deaths may be related to infection, hemorrhagic cystitis, bladder cancer or hematologic malignancy.^{34, 293-297}

The presence of generalized vasculitis or serious organ dysfunction (e.g., diffuse alveolar hemorrhage, severe GN, CNS, cardiac or GI involvement) at presentation probably warrants 3-6 months of daily, oral cyclophosphamide (CYC) before switching to IV CYC, methotrexate (MTX) or azathioprine (AZA).^{4, 33, 298, 299} Methotrexate (20-25 mg/week) has been shown to induce remission in about three quarters of WG patients who did not have life-threatening disease manifestations at the time of presentation.^{300, 301} It is effective in patients with active GN as well, but should not be used if the serum creatinine level is above 2 mg/dl, or if other serious complications such as DAH or CNS involvement are present.^{112, 302} Relapses appear to occur most often when patients are taking ≤ 15 mg/week.³⁰³ Both MTX and azathioprine (AZA) seem capable of maintaining CYC-induced remission.^{33, 304} Longer initial courses of oral CYC do not appear to reduce the risk of relapse when immunosuppressive drugs are finally withdrawn, but are associated with more toxicity.²⁹¹ The optimal duration of maintenance therapy, unfortunately, has not been established.³³

The debate over the relative efficacy of pulse, intravenous CYC (0.5-0.75 gm/m² every month) continues. Compared to daily, oral administration, pulse CYC is generally better tolerated, and associated with significantly less cumulative drug exposure.^{305, 306} One study, however, suggested that WG patients with widespread disease and high ANCA titers ($\geq 1:64$) do not respond well to intermittent dosing.³⁰⁷ The relapse rate has also been reported to be higher in some WG treatment trials utilizing pulse CYC.^{305, 307, 308} In other studies, favorable responses were seen when patients who failed to achieve remission or relapsed on pulse CYC were converted to a daily, oral regimen.^{305, 309} Notwithstanding, Haubitz and colleagues prospectively evaluated pulse, intravenous CYC versus daily, oral CYC in 47 WG and MPA patients with renal involvement and did not find a significant difference in relapse rate, renal outcome or patient survival.³⁰⁶ In fact, 100% of patients in the pulse CYC group achieved remission compared to 84% in the daily, oral CYC group. One of the studies showing a higher relapse rate with pulse CYC also showed equivalent efficacy in remission induction, and actually a lower mortality rate (33%) in the IV group compared to the daily, oral CYC group (44%).³⁰⁵ One study showed a much higher relapse rate for pulse CYC,³⁰⁸ but many of the patients included had previously failed daily, oral CYC therapy. In a meta-analysis of 11 non-randomized treatment trials in AASV, pulse CYC was actually less likely to fail to induce remission when compared to daily, oral CYC.³¹⁰ Patients receiving pulse CYC had a significantly lower risk of developing leukopenia and infection. Mortality, the risk of developing end stage renal disease, and relapse rate were not statistically different in the two groups, although there was a trend toward a higher relapse rate in patients treated with pulse CYC.³¹⁰ For patients with CSS who require cytotoxic therapy, and for most patients with MPA, pulse CYC is generally preferred unless immediately life-threatening disease is present.³¹¹

Corticosteroids are usually employed at an initial dose of 1 mg/kg/day along with cytotoxic therapy until significant clinical improvement is seen. For patients with advanced disease (e.g., DAH or severe GN), pulse methylprednisolone (250 mg IV every six hours) may be given for three days. Steroids alone may control upper airway disease and improve systemic symptoms, but they are not sufficient to prevent progression of pulmonary or renal disease.^{34, 40, 87, 312-314} In most cases, prednisone can be tapered to 20-30 mg/day by two to three months, and it can often be discontinued by six to twelve months. Some authors recommend tapering glucocorticoids on an alternate day regimen to limit side effects (including infection risk).^{1, 289, 292, 315}

While corticosteroids are the cornerstone of therapy for CSS, concomitant cytotoxic therapy is warranted in patients who present with cardiac involvement, significant mesenteric ischemia, CNS involvement or RPGN, and for others who respond poorly to corticosteroids alone, or relapse.¹¹² In patients that can be managed with corticosteroids as monotherapy, the

response is often brisk, and tapering can begin within one month.^{11, 102} However, in Solans' series of 32 patients, only 40% could be managed with corticosteroids alone.³¹⁶

Upper airway relapse can be significantly reduced by the addition of trimethoprim/sulfamethoxazole (TMP/SMX), but major organ system flares cannot.^{24, 304, 317-319} Some authors have recommended an aggressive regimen of nasal irrigation, nasal steroids and antibiotics (TMP/SMX or other) for isolated upper airway flares, followed by oral steroids or MTX if that is ineffective.⁸⁵

Intravenous immunoglobulin (IVIG) has shown benefit in some patients with AASV.³²⁰⁻³²⁴ In the only randomized, placebo controlled study, a single course of IVIG resulted in decreased disease activity among persistently active (despite conventional therapy) AASV patients, but not beyond 3 months.³²² No patients, however, received repeated dosing of IVIG. An immunosuppressant drug-sparing effect was not identified, and the IVIG group experienced more adverse events.³²² The authors suggested that the use of IVIG was, therefore, not supported. However, when 26 patients with active systemic vasculitis were given open label treatment with IVIG, 13 patients entered full remission, and all patients experienced clinical improvement.³²⁵ A number of these patients had disease refractory to steroids and cytotoxic agents, and others were previously untreated. After 12 months, clinical benefit was maintained in 18 of the patients. In another study, four of six new AASV patients with early disease entered full clinical remission lasting at least one year after a single course of IVIG while receiving no immunosuppressive treatment.³²⁰ IVIG has also shown a marked clinical benefit in a handful of patients with CSS, and can be considered in patients refractory to, or intolerant of, the usual therapies.³²⁶ The remission of systemic vasculitis may be associated with the generation of anti-idiotypic antibodies against ANCA.^{327, 328} Stimulation of idiotype regulation by IVIG may be the most important mechanism by which this therapy aids in the restoration of self-tolerance and clinical remission of the vasculitis.^{329, 330} IVIG may cause reversible impairment of renal function in patients with glomerulonephritis.³³¹ It carries some risk of viral transmission and has a number of potential infusion-related side effects, but it is generally safe.³³¹ It is currently utilized mostly in individuals with fulminant or refractory disease, but further study may reveal a broader role for IVIG in the management of AASV.

The precise role of plasma exchange in the management of AASV is similarly ill-defined. Its use has largely been limited to patients with fulminant presentations as well.^{17, 332} The purpose of plasma exchange is to remove pathogenic ANCA from the circulation. Other desirable effects may be linked to the removal of proinflammatory cytokines, complement and coagulation factors.²⁸⁶ Plasma exchange has been reported to improve outcomes in patients with AASV and either DAH or dialysis-dependent renal failure.^{17, 333-337} Two studies, however, failed to show additional benefit to steroid or combination steroid/cytotoxic therapy in other patients with MPA or CSS.^{338, 339} It has been suggested that initiation of plasma exchange in patients with severe pulmonary-renal syndromes or rapidly progressive GN may be wise while awaiting the completion of the laboratory and histopathologic assessment.¹⁷

Although no data exist to confirm their benefit, a variety of adjunctive measures may prove to be useful in dealing with AASV patients. The association in MPO-ANCA patients between proteinuria and poor long-term renal outcome argues for an aggressive strategy of blood pressure control and angiotensin converting enzyme inhibitor use in these patients.⁸⁸ Prophylactic antibiotics may be useful to prevent opportunistic infection, and to reduce nasal carriage of staphylococcal organisms that have been linked to disease relapse in WG.^{292, 340, 341} Local therapies like intranasal mupirocin (which has excellent anti-staphylococcal activity) or antibiotics with both direct anti-inflammatory, immunomodulating and antimicrobial effects (i.e., macrolides or tetracyclines) could prove useful.

For patients with refractory, life-threatening disease, immune ablation and stem cell rescue can be considered.³⁴² This allows for the deletion of autoreactive T cell clones, and is

being used with increasing frequency, in an experimental setting, for vasculitis and collagen vascular disease.³⁴²

Ultimately, targeted biological therapies should allow for control of these vasculitic illnesses, saving patients from the severe untoward effects associated with months or years of cytotoxic and glucocorticoid treatment.^{24, 272, 343} For example, interferon- α has demonstrated efficacy in diseases characterized by excess Th2 responses and hypereosinophilia, and has been useful in patients with CSS.³⁴³⁻³⁴⁷ Unfortunately, this drug has been associated with cardiac toxicity that could limit its use in this setting.³⁴⁸ Monoclonal antibodies directed at T lymphocytes have shown promise, anecdotally, in the treatment of various vasculitides, including some otherwise refractory cases.^{6, 349, 350} Rabbit anti-thymocyte globulin infusion managed to effect a partial or complete remission in four cases of WG refractory to conventional treatment.³⁵¹ Eterncept, an anti-tumor necrosis factor agent, has been associated with improvement in disease activity scores and few adverse events in a preliminary trial to assess safety in WG.³⁵² A randomized treatment trial in WG is underway. Interferon- γ is responsible for macrophage activation leading to increased production of reactive oxygen species and metalloproteinases, increased adhesion molecule expression and growth factor elaboration that promotes intimal hyperplasia.^{272, 353-355} Blocking its effects could also prove useful in patients with AASV. Inhibition of adhesion molecule function may provide another important therapeutic option in patients with systemic vasculitis.²⁷⁸ Treatment with an antibody to the integrin VLA-1 in a rat model of crescentic GN led to improved serum creatinine, improved renal survival, and decreased fibronectin and type IV collagen deposition.³⁵⁶

The Natural History of AASV

The natural history of AASV is variable. Some patients with AASV have several months of prodromal symptoms, while others present with fulminant, life-threatening disease manifestations with little forewarning.^{87, 286, 332} Overall, relapses occur in 25-50% of patients.^{13, 34, 99, 158, 291, 294, 296, 357} Patients with PR3-ANCA have a higher risk of relapse compared to those with MPO-ANCA.³⁵⁸ Patients with sinus involvement and elevated serum creatinine levels are also more likely to relapse.^{317, 359} CSS appears to have slightly higher remission rates (80-90%) and slightly lower relapse rates (25%) relative to other ANCA-associated systemic vasculitis patients.^{11, 99, 102, 175, 294, 316} For the most part, relapses are not as severe as the initial presentation. When they do occur, relapses tend to affect the same organ systems that were involved at presentation.³⁶⁰

With combination cytotoxic and corticosteroid therapy, the five year mortality for WG and MPA has been reduced to around 20%.⁴ In a follow-up study of 96 CSS patients, the survival at 6.5 years was 72%.⁹⁹ In Solans' analysis of 32 CSS patients, however, the long term survival was 90% at 5 years and beyond.³¹⁶ In this group, additional immunosuppressive agents were appropriately added to patients with severe neurological, cardiac or gastrointestinal involvement. Mortality in AASV is particularly high among elderly patients and those with renal disease (creatinine > 1.6 mg/dl, or proteinuria > 1 gm/day).^{123, 286, 359, 361} Other factors identifying patients at increased risk for dying include diffuse alveolar hemorrhage, cardiomyopathy, gastrointestinal or central nervous system involvement.^{58, 123} Mortality is higher in patients with c-ANCA, and in those who do not receive cyclophosphamide in their treatment regimen.³⁵⁹

It appears that much of the organ damage associated with AASV occurs early.³⁶² Part of this may be attributed to long average delays of 5-17 months from the onset of symptoms until diagnosis and initiation of treatment.^{34, 37, 363} The most important determinant of outcome in AASV is the presence of renal disease, and the serum creatinine level at presentation seems to be the best predictor of renal outcome.^{296, 359, 360} The presence of higher amounts of proteinuria at presentation is also indicative of a worse long-term renal prognosis.¹⁵⁴ Patients who do require renal replacement therapy at the time of presentation can recover sufficient function to have a

substantial dialysis-free interval.^{86, 335, 364, 365} In a retrospective study of 246 new patients with ANCA-associated renal vasculitis, 56 patients presented with a creatinine clearance above 6 mg/dl, and 55% of these were able to achieve dialysis independence.⁸⁶ The long term renal prognosis, however, is poor for these individuals.^{357, 366} The renal outcome of AASV patients after kidney transplantation, interestingly, is not different from other causes of end stage renal disease.³⁶⁷ Relapses of vasculitis are, in fact, uncommon in this setting.³⁶⁸

In some cases, it is hard to distinguish a disease flare from an opportunistic infection or other drug-related untoward effect. When patients experience clinical deterioration after initial improvement on a stable immunosuppressive regimen, they must be presumed to have an infection or other treatment related side effect until proven otherwise.¹¹² In a study of infectious complications among 207 vasculitis patients receiving cyclophosphamide where leukopenia was specifically avoided, 10% were still found to develop serious infections, often within the first three months of therapy.³⁶⁹ Glucocorticoid use further increases this risk.

Many patients will suffer from some type of permanent morbidity in the form of chronic renal insufficiency, visual or hearing impairment, chronic rhinosinusitis, long-term side effects of drug therapy, or, in the case of CSS, chronic steroid-dependent asthma.^{2, 34, 270} Hoffman and colleagues found that 80% of WG patients reported long-term limitations with regard to activities of daily living, with one third being forced to accept total disability.³⁶³

Summary

The ANCA-associated vasculitides are a group of closely related systemic, small vessel inflammatory disorders with protean clinical manifestations. Wegener's granulomatosis is best known for necrotizing, granulomatous upper and lower respiratory tract disease, kidney involvement and c-ANCA (anti-PR3) positivity. Churg Strauss syndrome, another granulomatous vasculitis, is strongly associated with asthma and eosinophilia. Major organs involved in CSS include the lung, skin, heart and nervous system. It is usually associated with p-ANCA (anti-MPO) positivity. Microscopic polyangiitis is essentially a diagnosis of exclusion. It does not have specific clinical or histopathologic findings that distinguish it from other AASV. Glomerulonephritis is almost always seen in this patient group, and it is usually associated with MPO-ANCA. Skin, GI and lung involvement are fairly common, but without granulomas. MPA is the most common cause of the classic pulmonary-renal syndrome with glomerulonephritis and diffuse alveolar hemorrhage.

Heritable factors predisposing individuals to the development of AASV (or affecting disease phenotype) may include allelic variations in MPO and PR3 (the ANCA target antigens), Fc receptors on leukocytes, adhesion molecules, α -1 antitrypsin (or other anti-proteases), and HLA antigens. Defective clearance of apoptotic cells may augment the inflammatory process in AASV, and play an important role in the development of ANCA and autoreactive T cell clones. Disease expression likely depends on the complex interaction of several endogenous and exogenous factors. A certain genetic makeup may be required for the development of systemic vasculitis, but infection or other environmental factors may provide the inflammatory stimulus needed to prime leukocytes and trigger vascular inflammation in ANCA-positive patients.

Rapid recognition of these disorders is key, preferably prior to the onset of widespread organ damage. The probability of suffering permanent morbidity is high when patients present with multisystem disease, even if the initial treatment is successful. Rapid induction of disease remission is important, particularly when vital organs are at risk; but initial treatment should be customized to the severity of the presentation, and may include cytotoxics, corticosteroids, plasma exchange and IVIG. Once remission is achieved, less toxic maintenance regimens are employed to reduce the chance of relapse. Relapse is, nevertheless, a common occurrence, and the optimal duration of maintenance therapy is simply not known. Ultimately, targeted biological therapies are expected to assume a major role in the control of AASV.

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