

“Dietary Protein in Bone Health: Friend and Foe”

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His interests include clinical evaluation and research in the areas of calcium, phosphorus and uric acid metabolism, with a special emphasis on the pathogenetic mechanisms of osteoporosis and kidney stone formation.

INTRODUCTION

Osteoporosis is a complex disorder characterized by an imbalance between bone resorption and bone formation, which results in deterioration of bone mass and of the microstructural integrity of bone. It is a major health issue with a significant impact on the economic burden in the nation. The number of physician visits for osteoporosis has increased significantly over the past decade (1). Moreover, the increased awareness of osteoporosis has been associated with a significant rise in the number of prescriptions for anti-osteoporosis medications. Pathophysiological mechanisms of osteoporosis are diverse and include a strong genetic predisposition (2-7), environmental risk factors, and the dysregulation of hormonal metabolism (8-15). Although new medications for osteoporosis are now available, the specific role of diet as a modifier should not be ignored.

The NIH consensus panel on optimal calcium intake (9) has acknowledged the importance of dietary protein intake in the development of an optimal calcium balance. However, guidelines for an optimum protein intake were not provided due to the assumption that such dietary modifications may cause a great deal of confusion in the public domain. The objective of this review is to explore the potential pathophysiological associations between dietary protein intake and the development of skeletal bone loss.

Dietary Proteins and Bone Health

The recommended dietary allowance (RDA) for protein is 0.8 g/kg/day (16). The typical American diet provides 1.2 g protein/kg/day (17, 18), exceeding the established RDA. Dietary protein intake greater than 1.6 g/kg/day is considered to be high (19, 20). The widespread use of low carbohydrate high fat-diets including the Atkins' Diet and the South Beach Diet emphasizes the importance of long-term consideration of the impact of high dietary protein intake on bone health and kidney stone disease (21).

Epidemiology

Epidemiologic studies have provided conflicting results on the role of dietary protein intake on bone health (22-27). The population-based Framingham osteoporosis study showed a protective role of high protein intake on bone mineral density in elderly men and women (25). In this observational study, subjects in the lowest quartile of protein consumption demonstrated the largest loss in bone mineral density at the vertebral spine and femoral neck over 4 years of follow-up (Figure 1). Moreover, few studies have also shown that hip fractures occur with an increased frequency in malnourished elderly (26-28). Furthermore, most epidemiological studies have demonstrated a positive relationship between protein intake and bone mineral density (25, 29-31).

The results of these population-based studies differ from a cross-cultural comparison, showing the incidence of non-traumatic hip fractures to be directly associated with the amount of protein consumption. The incidence of non-traumatic hip bone fractures was found to be related to per person consumption of protein of an individual country, with a higher incidence in

industrialized compared to less industrialized societies (23). However, it has been argued that the differences in hip fracture incidence in this study may have been related to the ethnic differences in the studied population (32, 33) as lower rates were detected in countries inhabited by Blacks

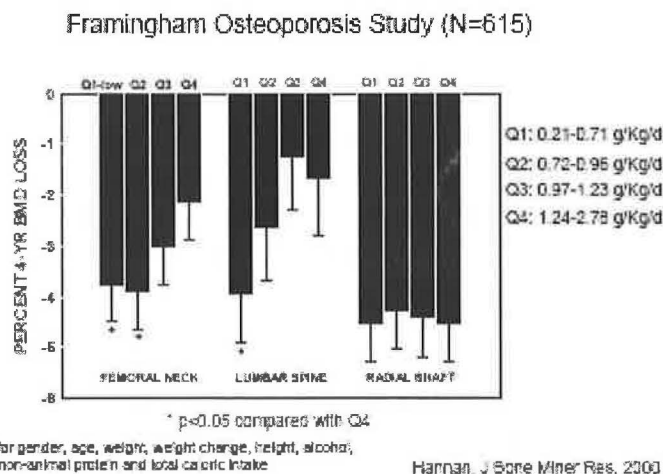
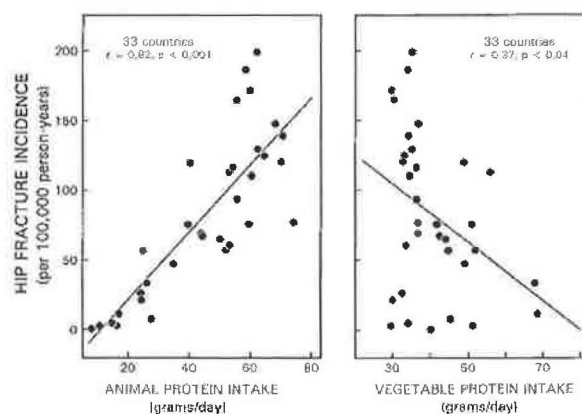


Figure 1. BMD according to quartile of protein intake

(Figure 2). Furthermore, the incidence of hip fractures was demonstrated to be significantly reduced in relation to an increased consumption of vegetable foods. This study suggests that vegetable foods, by provision of alkali, may possibly mitigate the deleterious effect of acid generating animal protein intake on bone.

The results of these cross-cultural reports are consistent with another epidemiological study in a cohort of 85,900 women, age 35-59 years, who were participating in Nurse's Health Study (25). The risk of forearm bone fractures was found to be significantly higher in those women with a protein consumption exceeding 95 g/day compared with those who consumed less than 68 g/day. In this study no such relationship was detected between the consumption of vegetable protein and the risk of forearm bone fractures.

The differences in the results of these epidemiologic studies underscore the difficulties involved in estimating the exact amount of protein intake using food frequency questionnaires, the baseline protein intake and the type of protein consumed (such as whole meal protein or purified protein). Furthermore, contributions of other nutrients to calcium balance including sodium, calcium, and phosphorus have not been examined closely. In addition, a potential for bias exists assuming food consumption for whole population is also reflective of



Frassetto, J Gerontol, 2000

Figure 2. Cross-Cultural Relationship Between Hip Fracture and Protein Intake

dietary consumption of the studied population. However, population and epidemiological studies have several strengths including large sample size and lack of selection bias.

Pathogenetic mechanisms of protein-induced bone loss

Extracellular calcium is in constant equilibrium between intestinal calcium absorption, renal reabsorption of calcium, and bone resorption. In a steady state, when an individual is in balance, net intestinal absorption of calcium is matched by urinary calcium excretion, and calcium exchanges across the bone are equal. Urinary calcium excretion plays a significant role in calcium homeostasis. The importance of urinary calcium excretion in overall calcium balance is

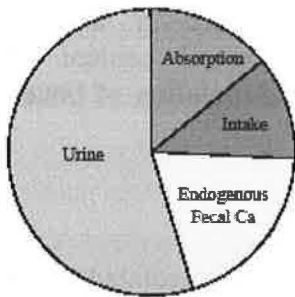


Figure 3. Determinants of Calcium Balance (35)

evident from 560 balance studies performed in healthy middle-aged women (35). Calcium intake and absorption accounted for 25% and urinary calcium for 50% of the changes in calcium balance (Figure 3). The current scheme for the development of bone loss with a high protein intake has centered on the effects of high protein diet on increasing urinary calcium excretion.

The metabolic processes that contribute to the endogenous acid load with consumption of animal protein play a key pathogenetic role in protein-induced bone loss. These processes involve oxidation of the sulfhydryl groups of the aminoacids cystine and methionine found in protein to form sulfuric acid. In this model, protons generated from increased animal protein intake elevate urinary calcium excretion by involving the three target organs regulating calcium homeostasis.

Role of bone in protein-induced acidosis

Metabolic acidosis primarily promotes bone resorption through three different mechanisms: (a) physicochemical mechanism involving dissolution of bone mineral, (b) cellular mechanisms mediated by osteoclastic bone resorption, and (c) hormonal mechanism through stimulation of PTH secretion secondary to inhibition of renal tubular calcium reabsorption.

Physicochemical Mechanism

Bone is major source of calcium in the body as 99% of body calcium is contained in the bone. Bone also contains 35% of total body sodium and 60% of total body magnesium (36). In addition, bone is a major reservoir of alkali and contains 80% of total carbon dioxide in the body (including carbonate, bicarbonate and CO₂) and 80% of body citrate (37) (Table 1). Thus, bone

Table 1. Ionic Composition of Bone Mineral

Cations	Anions
Calcium (6.66)	Phosphate (4.02)
Sodium (0.32)	Carbonate (0.79)
Magnesium (0.18)	Citrate (0.05)
Potassium (0.02)	Chloride (0.02)

Values are indicated as mmol/g of dry fat-free bone

acts as a major extracellular buffer by releasing alkali to the surrounding extracellular environment to support acid-base homeostasis. In vitro studies in fetal mouse calvaria have shown that acute metabolic acidosis stimulates calcium release from both dead bone and live bone into the surrounding medium (38, 39). This suggests that with acute acid exposure, physiochemical mechanisms are operating to resorb bone and release calcium. The initial physiochemical reaction to acute metabolic acidosis involves an exchange of sodium for hydrogen and release of bone bicarbonate from the mineral surface (40). This process is

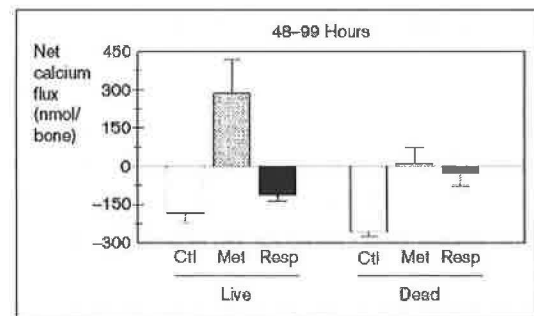
Table 2. Ionic Fluxes in Metabolic Acidosis

	Metabolic Acidosis	
	Acute	Chronic
H ⁺ Influx	↑ ↑ ↑	↑ ↑ ↑
Ca ²⁺ Efflux	↑ ↑	↑ ↑ ↑
Efflux of ions other than Ca ²⁺	HCO ₃ ⁻ , Na ⁺ , K ⁺	CO ₃ ⁻² , PO ₄ ⁻³

accompanied by a small release of calcium from bone (Table 2). In chronic metabolic acidosis, release of calcium carbonate comprises the main buffering mechanism. Thus, in chronic metabolic acidosis, in contrast to acute metabolic acidosis, there is a significant reduction in total mineral content of bone accompanied by the dissolution of bone crystal (Table 2).

Cell-mediated mechanism

It has been demonstrated that with chronic metabolic acidosis, cell-mediated osteoclastic bone resorption predominates over the physicochemical mechanisms (41) (Figure 4). With chronic metabolic acidosis, calcium efflux was shown to occur only with live bone, indicative of the predominance of a cell-mediated pathway (Figure 4). Moreover, chronic metabolic acidosis-induced release of calcium was shown to be associated with an increased release of the osteoclastic enzyme β -glucuronidase into the culture medium (42), suggestive of an increase in osteoclastic cell activity. This was accompanied by inhibition of osteoblastic collagen synthesis and consequently reduction of alkaline phosphatase release from osteoblasts (42).



Bushinsky, Am J Physiol, 1989

Figure 4. Net Calcium Flux From Live and Dead Bone During Chronic Acidosis

The molecular mechanisms by which metabolic acidosis modulates cell-mediated bone remodeling has recently been elucidated (43, 44). Metabolic acidosis inhibits specific osteoblastic matrix protein synthesis and alkaline phosphatase activity and stimulates production of prostaglandin E₂ (PGE₂) by osteoblasts (45, 46). The increased osteoblastic production of PGE₂ increases the osteoblastic expression of receptor activator of nuclear factor Kappa B ligand (RANKL). RANKL is the major downstream cytokine that stimulates osteoclastogenesis by binding to its receptor (receptor activator of nuclear factor Kappa B; RANK) on osteoclastic precursor cells, transforming them into activated bone-resorbing osteoclasts, and thereby buffering the proton load to regulate acid-base homeostasis.

Despite extensive in vitro experiments, the role of chronic metabolic acidosis on bone

histomorphometry in vivo using animals and human subjects has not been fully investigated. Recently, a high casein diet (a model of high animal protein intake) compared to low casein diet (a model of low animal protein intake) provided for two months in pair-fed rats caused a higher urinary calcium excretion and increased the propensity for kidney stone formation in the high casein group (47). Moreover, histomorphometric analysis of femur after 59 days on the diet showed a marked increase in bone resorption in the high casein group.

Hormonal Mechanism

The effects of chronic metabolic acidosis on calcium metabolism are complex and depend on the response of calcitropic hormones to calcium and phosphate fluxes in the gut, the skeleton and the kidney. Parathyroid hormone (PTH) secretion could increase or decrease depending on the prevailing serum ionized calcium concentration which is altered by renal calcium wasting or calcium flux from the bone respectively. The changes in PTH secretion directly influences skeletal bone turnover independent of the prevailing acid-base status (38).

Role of kidney in protein-induced hypercalciuria

Another potential mechanism of hypercalciuria with increased animal protein intake includes the inhibition of the renal tubular reabsorption of calcium. This inhibition of renal tubular calcium reabsorption has been identified both in human and animal studies (48-50) using animal protein or ammonium chloride-induced metabolic acidosis. However, until recently the molecular mechanism for this inhibitory effect was not fully explored.

The kidney plays a key role in regulation of calcium homeostasis and approximately 98% of calcium filtered at the glomerulus is reabsorbed by the nephrons. In the kidney, paracellular reabsorption of calcium is responsible for 80-85% across the proximal tubules and the thick ascending limb of Henle (51, 52). The transcellular reabsorption of calcium occurs principally in the distal convoluted tubule (DCT) (Figure 5). This transcellular calcium reabsorption starts with

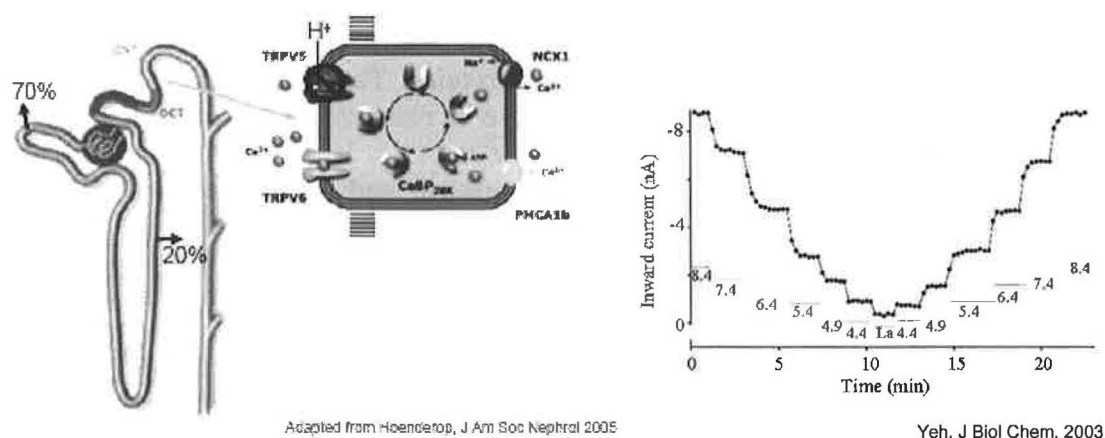


Figure 5. Renal Handling of Calcium and Effect of Extracellular pH on TRPV-5 (Calcium Channel)

passive entry of calcium through calcium channels in the apical membrane, is followed by diffusion through the cytosol facilitated by binding to calcium-binding protein calbindin-28K, and finally extrusion through the basolateral membrane. The calcium exit from the basolateral

membrane requires energy and is mediated by sodium/calcium exchanger and calcium-ATPase. The first step of passive entry through calcium channel in the apical membrane is the rate-limiting step in transepithelial calcium reabsorption in the distal nephron (53).

Recently, the transient receptor potential Type 5 (TRPV-5) channel present in kidney and intestine was shown to play a key role in transepithelial reabsorption of calcium in these target organs (54, 55). In a whole cell patch clamp study, extracellular protons inhibited TRPV-5 by titrating glutamate 522 in the extracellular loop between the putative fifth transmembrane domain and the pore region (Figure 5). Thus, direct proton sensing via titration of glutamate 522 appears to mediate acid-induced inhibition of TRPV-5 possibly by changing the protein conformation. Another mechanism may be through an increase in the abundance of TRPV-5 and TRPV-6 in chronic metabolic acidosis (56).

Role of intestine in protein-induced hypercalciuria

The potential role of intestine and its relationship to hypercalciuria in metabolic acidosis has not been fully elucidated. Decreased PTH secretion from a transient rise in serum ionized calcium concentration due to acidosis-induced excessive bone resorption may lower serum circulating 1,25-(OH)₂-D or directly impair conversion of 25 hydroxyvitamin D to 1,25-(OH)₂-D (57) and consequently lower intestinal calcium absorption (58). On the other hand, hypercalciuria from inhibition of renal tubular calcium reabsorption due to metabolic acidosis would transiently lower serum ionized calcium concentration which in turn raises serum circulating concentration of serum PTH and 1,25-(OH)₂-D (59, 60) and ultimately raises intestinal calcium absorption. Alternatively, a low serum phosphorous and an increase in renal mass caused by metabolic acidosis have been shown to increase 1,25-(OH)₂ D levels in human subjects (61, 62), which may increase intestinal calcium absorption. These opposing mechanisms may mitigate the effect of metabolic acidosis on intestinal calcium absorption. Indeed, most clinical balance studies have shown the lack of effects of metabolic acidosis on intestinal calcium absorption (63, 64).

Recently, two short-term metabolic studies (65, 66) have suggested that intestinal calcium absorption increases with consumption of high dietary proteins. The rise of intestinal calcium absorption during high protein diet was suggested in part to be responsible for the significant rise in urinary calcium excretion (66) with high protein consumption. It has been speculated that dietary protein and aromatic aminoacids may increase gastrin and gastric acid secretion, by interacting with calcium sensing receptors on antral G cells and parietal respectively (67, 68). In fact, the calcium sensor agonist Cinacalcet® which is currently used in the treatment of hyperparathyroidism has structural similarity to phenylalanine and tyrosine. It is plausible to suggest that increased gastric acid secretion perhaps by improving calcium solubility may increase intestinal calcium absorption.

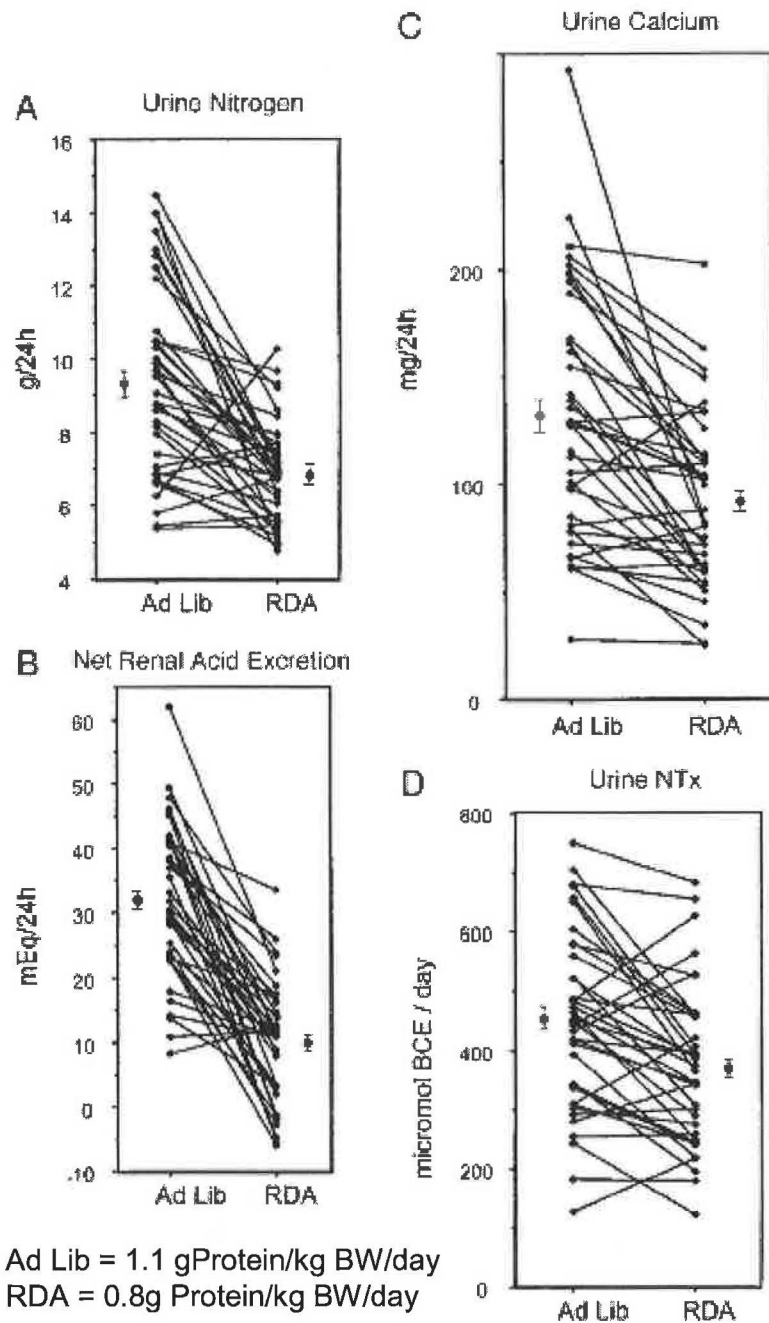
The effects of high dietary protein: short-term clinical studies

The exact causal relationship between high protein consumption and bone disease could not be derived from observational studies. Furthermore, to date, there is no large scale controlled study that has investigated the pathogenic role of high animal protein intake on bone. Our

knowledge of a causal relationship between high protein consumption and bone disease is limited to short-term physiological studies.

In one 2-week study in 39 healthy premenopausal women, dietary protein was reduced from a habitual intake of 1.1 g protein/kg body weight/day to the U.S. recommended dietary allowance (RDA) of protein of 0.8 g protein/kg body weight/day (69). This change significantly decreased urinary nitrogen (a marker of protein intake) and significantly reduced net acid excretion. On the diet with RDA protein content, urinary calcium decreased significantly by 1 mmol/day (42 mg/day) and urinary N-telopeptide (a marker of bone resorption) decreased significantly (Figure 6). These changes were achieved despite similar caloric intake and constant dietary compositions of sodium, potassium, and phosphorus between the two diets.

In another study, consumption by 10 healthy subjects of a low-carbohydrate high-protein diet (70) for six weeks delivered a significant acid load of 50 meq/day and increased urinary calcium by 2.5 mmol/day (100 mg/day). The increase in urinary calcium was not compensated by an increase in fractional intestinal calcium absorption. As a result, the estimated



Ince, J Clin Endocrinol Metab, 2004

Figure 6. Changes in Urinary Parameters with Reduction in Protein Intake to the Recommended Daily Allowance (0.8g of Protein/kg Body Weight/day).

calcium balance decreased significantly by 90 to 130 mg/day (2.25 to 3.24 mmol/day). The serum total carbon dioxide concentration did not change significantly, despite the delivery of a markedly high acid load. Thus, it is plausible to suggest a potential role of bone as an extracellular buffer to maintain normal acid-base homeostasis (71), despite marked endogenous acid production.

No definitive conclusion could be derived from the results of the short-term physiological investigations to the effect of chronic protein load on skeletal bone health. Further prospective investigation of the potential increase in risk of bone loss with a high protein intake is warranted.

Pathogenetic mechanisms of high dietary protein and aging-induced bone loss

In healthy adult subjects in a steady state, the endogenous acid production (EAP) matches net acid excretion (NAE) (72). The homeostatic mechanism regulating acid-base balance is finely tuned. It has been shown that changes within a wide range of EAP induced by diet ranging from 0 to 150 meq/day (73) resulted in minimal changes in serum extracellular bicarbonate concentration. However, the stimulation of this homeostatic mechanism has a “trade off” similar to the adaptation mechanism previously described in the regulation of calcium, phosphorus and PTH homeostasis during the progression of chronic renal failure. The adaptation mechanism with chronic metabolic acidosis in response to the increase in EAP involves the release of alkali salts from the bone in addition to the increase in the urinary ammonium (NH_4^+) and titratable acidity excretion by the kidney to neutralize an excess acid load and to maintain a near normal serum bicarbonate concentration (Figure 7).

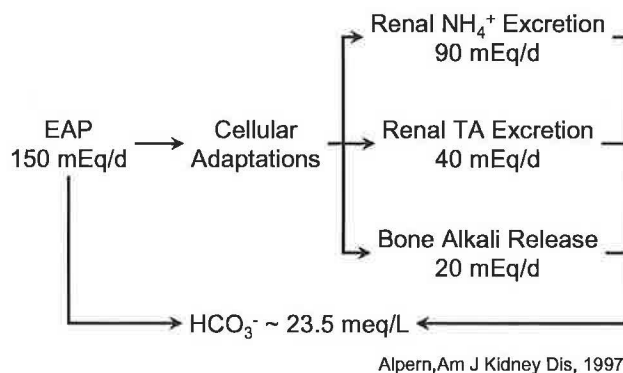


Figure 7. Acid-Base Homeostasis in Elderly Subjects and Individuals on “High Protein” Diets

Two commonly unrecognized clinical conditions associated with chronic metabolic acidosis are aging and excessive meat intake. Several investigators have provided evidence of a progressive decline in plasma bicarbonate concentration and blood pH with aging (74). These changes were shown to occur despite a habitual dietary intake. Aging is associated with a progressive loss of nephron mass, which results in progressive decrease in glomerular filtration rate (75). It has been demonstrated that the renal capacity to excrete an acid load is diminished in older individuals (74). Therefore, it is conceivable that a low grade chronic metabolic acidosis is created in the elderly consuming a habitual Westernized acid ash diet. The impact of this condition will be grave in this population due to its high risk of osteoporosis and bone fractures.

Pathogenesis of the protective role of protein in bone health

Over 6 decades ago, Fuller Albright suggested that osteoporosis may not be due to “lack of calcium and phosphorus in the diet, but may be really due to protein starvation” (76). Additional data has emerged in the recent years to support Albright’s hypothesis of a protective role of protein in bone health.

Pathogenetic mechanisms of protein-reduced bone loss

Dietary proteins play a key regulatory role in the production and the metabolism of insulin-like growth factor 1 (IGF-1) (Figure 8) (77). IGF-1 has been shown to stimulate both proliferation and differentiation of osteoblasts, and thereby increases bone collagen synthesis (78-80). The serum circulating concentrations of IGF-1 and IGF-1 binding protein 3 have been shown to be lower in osteoporotic patients with vertebral fractures than in osteoporotic individuals without fractures (Figure 9) (81). A defect in osteoid mineralization has also been recently demonstrated in the selective knockout of IGF-1 receptor gene in mouse osteoblasts (82). Moreover, protein intake may directly affect bone collagen structure as abnormalities in lysine residues in the $\alpha 1$ Type 1 collagen of bone have been shown in protein-restricted rats (83). Lysine plays a

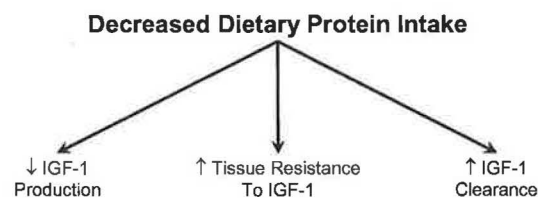


Figure 8. Impaired IGF-1 Action associated with Decreased Protein Intake

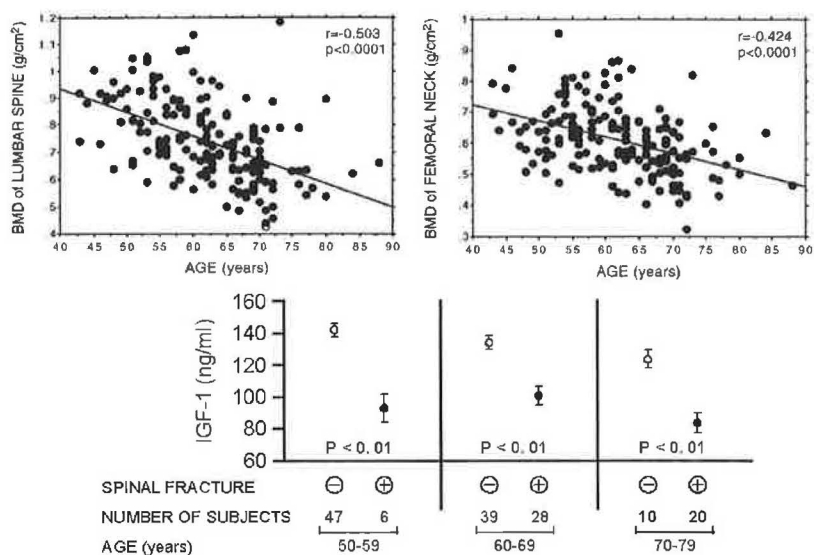


Figure 9. Correlation Between Serum IGF-1 Levels, BMD, and Fractures

significant role in Type 1 collagen cross linking which is predominantly found in bone. It has not been clearly elucidated whether lysine directly affects calcium homeostasis.

Furthermore, protein malnutrition due to an impaired anabolic effect (27) has been shown to reduce muscle strength and perhaps increase frailty and fall in older subjects, which in turn increases the risk of bone fractures.

Protein deficiency may also enhance skeletal bone resorption. Bone resorption is mediated by the upregulation of cytokine release, as well as by the dysregulation of gonadal hormonal metabolism (84, 85). The impact of protein deficiency has been recently examined in an experimental animal model, by inserting titanium rod implants into tibial bone (85). Low protein

(2.5%) isocaloric diet compared to a normal protein isocaloric diet (15%) was shown to induce defective bone microarchitecture, and to significantly diminish resistance to implant pull-out (85).

Protein supplementation and reduced bone loss-clinical studies

There is substantial observational evidence to suggest that protein supplementation influences the outcome of hip fractures. Moreover, a randomized double-blind study conducted in 82 elderly patients with recent hip fractures demonstrated that protein supplementation compared to isocaloric placebo (86) resulted in a significantly higher serum circulating concentration of IGF-1, significantly lower proximal femoral bone mineral density loss, and shortened recovery period. This study did not conclusively exclude the possibility that IGF-1 induced increased muscle strength that in turn lowered the loss of bone mineral density at the proximal femur.

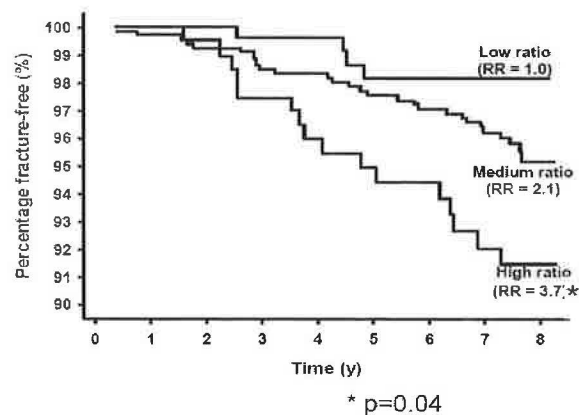
However, there has been disagreement on the effects of protein consumption and its impact on bone fractures (11). High protein consumption has been shown to be associated with an increased incidence of bone fractures in observational cross-cultural studies (22, 23, 34), as well as within population studies. The low bone mineral density and hip fractures in subjects with underlying protein malnutrition does not totally preclude the association of high protein consumption with an increased risk of bone fractures, in an otherwise healthy protein-replete population.

Countermeasures to dietary protein induced-bone loss

Alkali therapy is believed to prevent bone loss by several mechanisms. It may inhibit osteoclastic bone resorption by the direct action of alkali (87). Furthermore, alkali treatment may indirectly prevent bone resorption by reducing urinary calcium (88, 89). Finally, there is also some evidence that alkali treatment may stimulate bone formation (87).

Dietary measures

Animal proteins are a dietary source of acid ash, while vegetable proteins are metabolized without significant production of endogenous acid. In a recent longitudinal prospective study, the contribution of a high ratio of dietary animal to vegetable protein on bone mineral density and the risk of hip bone fractures in 1035 Caucasian elderly women was examined (90). The women



Sellmeyer, Am J Clin Nutr, 2001

Figure 10. Animal - Vegetable Protein Ratio and Probability of Hip Fracture

with a high ratio of animal to vegetable protein consumption had a higher rate of femoral neck bone mineral density loss than those with a low ratio. A proportional hazard model adjusting for age, weight, calcium intake, smoking habit, alcohol consumption, total protein consumption, physical activity and estrogen intake was used. Survival free of hip fracture was significantly lower in those with the high ratio of animal to vegetable protein intake (RR = 3.7, $p=0.04$) than those with low ratio (RR = 1.0) and subjects with medium ratio (RR = 2.1) (Figure 10). These results suggest that the effect of protein intake on bone is related to endogenous acid production.

Pharmacological countermeasures

In a recent study, calcium and phosphorus balances were examined in postmenopausal women with a high protein intake (89). Potassium bicarbonate treatment at the dosage equivalent to neutralize endogenous acid production caused a significant fall in urinary calcium excretion and a net positive calcium balance. This resulted in a significant fall in bone resorption and a rise in bone formation as assessed by the fall in urinary hydroxyproline excretion and the rise in serum osteocalcin concentration, respectively.

The above findings are in part consistent with our study in 18 healthy postmenopausal women with dietary protein intake equivalent to the recommended dietary allowance (RDA) of 0.8 g protein/kg/d for two weeks (91). Potassium citrate at a much smaller dose of 40 meq/day caused a significant reduction in urinary calcium excretion and a significant rise in estimated calcium balance (Figure 11).

The results of the above studies were also confirmed in healthy young subjects who were in a mild metabolic acidosis while maintained on habitual Westernized diet. An equimolar amount of sodium bicarbonate and potassium bicarbonate was substituted for sodium chloride and potassium chloride (92). Lowered urinary calcium excretion and urinary markers of bone resorption occurred independent of potassium intake. The result was suggestive of a protective effect of alkali treatment neutralizing the relatively high acid ash diet.

The hypocalciuric effect of alkali therapy has been shown to be sustained during chronic alkali treatment (93). This effect has been demonstrated to be dose-dependent and is greatest in those subjects with the highest baseline urinary calcium excretion. It has been inferred that with chronic alkali treatment and sustained hypocalciuria, a substantial amount of bone calcium content may be spared (93). However, it has also been argued (94, 95) that a reduction in intestinal calcium absorption from alkali-induced hypocalciuria may negate the bone sparing effect of alkali treatment. Therefore, it is necessary to design long-term clinical trials to examine whether chronic alkali treatment improves bone mineral density and reduces the incidence of bone fractures.

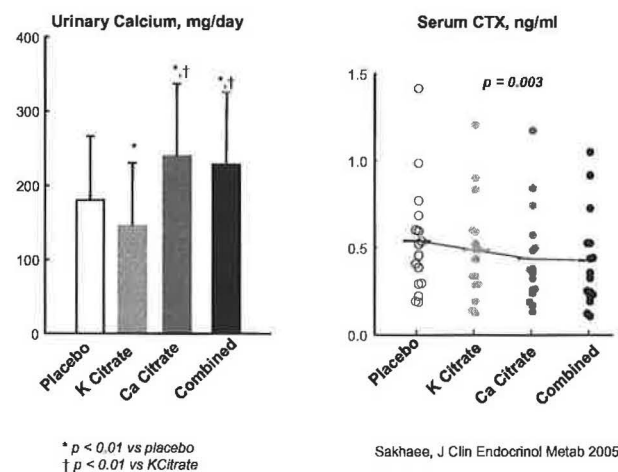


Figure 11. Change in Urinary Calcium and serum CTX (a marker of bone resorption) in healthy postmenopausal women treated with K Citrate, Ca Citrate or their Combination

CONCLUSIONS

Protein malnutrition is associated with low bone mineral density and increased fall and fracture risk. On the other hand, there is increasing evidence that chronic subtle metabolic acidosis caused by aging and increased dietary protein consumption potentially contributes to bone loss and osteoporosis. However, due to the subtle nature of this effect, these conditions are generally unnoticed by most clinicians. Regardless of the deficiencies in our current knowledge, physicians must be reminded to provide appropriate care to patients to overcome the consequence of chronic subtle metabolic acidosis on bone. Alkali treatment is a logical choice to maintain the anabolic effect of protein on bone, while mitigating its deleterious influence. One major question is whether the hypocalciuric effect of long-term alkali treatment translates into a positive calcium balance or is due to an adaptive decline in intestinal calcium absorption. Further research must focus on innovative approaches that will help improve our current knowledge into the long-term effect of protein on bone.

References

1. Stafford RS, Dreiling RL and Hersh AL. National trends in osteoporosis visits and osteoporosis treatment, 1988-2003. *Arch Intern Med*, 2004; 164:1525-1530.
2. Pocock NA, Eisman JL, Hopper GM, Yeats PN and Ebert S. Genetic determinants of bone mass in adults: A twin study. *J Clin Invest*, 1987; 80:706-710.
3. Flicker L, Hooper JL, Rodgers L, Kaymakci B, Green RM and Wark JD. Bone density determinants in elderly women: A twin study. *J Bone Miner Res*, 1995; 10:1607-1613.
4. Johnson ML, Gong G, Kimberling W, Recker SM, Kimmel DB and Recker RR. Linkage of a gene causing high bone mass to human chromosome 11 (11q 12-13). *Am J Hum Genet*, 1997; 60:1326-1332.
5. Morrison NA, Qi JC, Tokita A, Kelly PJ, Crofts L, Nguyen TV, Sambrook PM and Eisman JA. Prediction of bone density from vitamin D receptor alleles. *Nature*, 1994; 367:284-287.
6. Uitterlinden AG, Burger H, Huang Q, Yue F, McGuigan FEA, Grant SFA, Hofman A, Van Leeuwen JPTM, Pols HAP and Ralston SH. Relation of alleles of the collagen Type I $\alpha 1$ gene to bone density and the risk of osteoporotic fractures in postmenopausal women. *New Engl J Med*, 1998; 338:1016-1021.
7. Raiz LG. Pathogenesis of osteoporosis: Concepts, conflicts and prospects. *J Clin Invest*, 2005; 115:3318-3325.
8. Cummings SR, Nevitt MC, Browner WS, Stone K, Fox KM, Ensurd KE, Cauley J, Black D and Vogt TM. Risk factors for hip fracture in white women. *New Engl J Med*, 1995; 332:767-773.
9. Optimal calcium intake. NIH consensus conference. *JAMA*, 1994; 272:1942-1948.
10. Ginty F. Dietary protein and bone health. *Proc Nutr Soc*, 2003; 62:867-876.
11. Rizzoli R and Bonjour JP. Dietary protein and bone health. *J Bone Miner Res*, 2004; 19:527-531.
12. Khosla S, Melton LJ, 3rd, Atkinson EJ and O'Fallon WM. Relationship of serum sex steroid levels to longitudinal changes in bone density in young versus elderly men. *J Clin Endocrinol Metab*, 2001; 86:3555-3561.
13. Eastell R, Yergey AL, Vieira NE, Cedel SL, Kumar R and Riggs BL. Interrelationship among vitamin D metabolism, true calcium absorption, parathyroid function, and age in women: evidence of an age-related intestinal resistance to 1,25-dihydroxyvitamin D action. *J Bone Miner Res*, 1991; 6:125-132.
14. Bilezikian JP. Estrogens and postmenopausal osteoporosis: was albright right after all? *J Bone Miner Res*, 1998; 13:774-776.
15. Ebeling P, Jones J, Burritt M, Duerson C, Lane A, Hassager C, Kumar R and Riggs BL. Skeletal responsiveness in postmenopausal osteoporosis. *J Clin Endocrinol Metab*, 1992; 75:1033-1038.
16. 2002 Dietary reference intakes for energy, carbohydrate, fiber, fat, fatty acids, cholesterol, protein, and amino acids (micronutrients). Washington, DC: National Academies Press (<http://books.nap.edu/catalog/10490.html>).
17. U.S. Department of Agriculture Nationwide food consumption survey. Nutrient intakes: individuals in 48 states, year 1977-1978. Hyattsville, MD:USDA, consumer Nutrition Division, Human Nutrition Information Service. Report No 1-2. 1984.

18. St. Jeor ST, Howard BV, Prewitt TE, Bovee V, Bazzarre T and Eckel RH. Dietary protein and weight reduction: a statement for healthcare professionals from the Nutrition Committee of the Council on Nutrition, Physical Activity, and Metabolism of the American Heart Association. *Circulation*, 2001; 104:1869-1874.
19. Eisenstein J, Roberts SB, Dallal G and Saltzman E. High-protein weight-loss diets: are they safe and do they work? A review of the experimental and epidemiologic data. *Nutr Rev*, 2002; 60:189-200.
20. Kerstetter JE and Allen LH. Dietary protein increases urinary calcium. *J Nutr*, 1989; 120:134-136.
21. Bonow RO and Eckel RH. Diet, Obesity, and Cardiovascular Risk. *N Engl J Med*, 2003; 348:2057-2058.
22. Feskanich D, Willett WC, Stampfer MJ and Calditz GA. Protein consumption and bone fractures in women. *Am J Epidemiol*, 1996; 143:472-479.
23. Abelow BJ, Hofford TR and Insogna KL. Cross-cultural association between dietary animal protein and hip fracture: a hypothesis. *Calcif Tissue Int*, 1992; 50:14-18.
24. Mazess RB and Mather W. Bone mineral content of North Alaskan Eskimos ^{1, 2, 3}. *Am J Clin Nutr*, 1974; 27:916-925.
25. Hannan MT, Tucker KL, Dawson-Huges B, Cupples LA, Felson DT and Kiel DP. Effect of dietary protein on bone loss in elderly men and women: The Framingham osteoporosis study. *J Bone Miner Res*, 2000; 15:2504-2012.
26. Munger RG, Cerhan JR and Chiv BC. Prospective study of dietary protein intake and risk of hip fracture in postmenopausal women. *Am J Clin Nutr*, 1999; 69:147-152.
27. Wengreen HJ, Munger RG, West NA, Cutler DR, Corcoran CD, Zhang J and Sassano NE. Dietary protein intake and risk of osteoporotic hip fracture in elderly residents of Utah. *J Bone Miner Res*, 2004; 19:537-545.
28. Bastow MD, Rawlings J and Allison SP. Undernutrition, hypothermia, and injury in elderly women with fractured femur: an injury response to altered metabolism? *Lancet*, 1983; 1:143-146.
29. Promislow JH, Goodman-Gruen D, Slymen DJ and Barnett-Connor E. Protein consumption and bone mineral density in the elderly: The Rancho Bernardo Study. *Am J Epidemiol*, 2002; 155:636-644.
30. Geinzo G, Rapin CH, Rizzoki R, Kraemer R, Buchs B, Slosman D, Michel JP and Bonjour JP. Relationship between bone mineral density and dietary intakes in the elderly. *Osteoporosis Int*, 1993; 3:242-248.
31. Cooper C, Atkinson EJ, Hensrud DD, Wahner HW, O'Fallon WM, Riggs BL and Melton LJ, 3rd. Dietary protein intake and bone mass in women. *Calcif Tissue Int*, 1996; 58:320-325.
32. Heaney RP. Protein and calcium: antagonists or synergists? *Am J Clin Nutr*, 2002; 75:609-610. Heaney RP. Ethnicity, bone status, and the calcium requirement. *Nutr Res*, 2002; 22:153-178.
33. Heaney RP. Ethnicity, bone status and the calcium requirement. *Nutr Res*, 2002; 22:153-178.
34. Frassetto LA, Todd KM, Morris JRC and Sebastian A. Worldwide incidence of hip fracture in elderly women: relation to consumption of animal and vegetable foods. *J Gerontol*, 2000; 55:M585-M592.

35. Barger-Lux MJ, Heaney RP, Pachard PT, Lappe JM and Recker RR. Nutritional correlates of low calcium intake. *Clin Appl Nutr*, 1992; 2:39-44.
36. Armstrong WD and Singer L. Composition and constitution of the mineral phase of bone. *Clin Orthop*, 1965; 38:179-190.
37. Pasquale SM, Messier AA, Shea ML and Schaefer KE. Bone CO₂-titration curves in acute hypercapnia obtained with a modified titration technique. *J Appl Physiol*, 1980; 48:197-201.
38. Krieger NS, Frick KK and Bushinsky DA. Mechanism of acid-induced bone resorption. *Curr Opin Nephrol Hypertens*, 2004; 13:423-436.
39. Bushinsky DA, Sessler NE, Glena RE and Featherstone JD. Proton-induced physiochemical calcium release from ceramic apatite discs. *J Bone Miner Res*, 1994; 9:213-220.
40. Bushinsky DA, Levi-Setti R and Coe FL. Ion microprobe determination of bone surface elements: effects of reduced medium pH. *Am J Physiol*, 1986; 250:F1090-F1097.
41. Bushinsky DA. Net calcium efflux from live bone during chronic metabolic, but not respiratory, acidosis. *Am J Physiol*, 1989; 256:F836-F842.
42. Krieger NS, Sessler NE and Bushinsky DA. Acidosis inhibits osteoblastic and stimulates osteoclastic activity in vitro. *Am J Physiol*, 1992; 262:F442-F448.
43. Frick KK, Jiang L and Bushinsky DA. Acute metabolic acidosis inhibits the induction of osteoblastic egr-1 and type 1 collagen. *Am J Physiol*, 1997; 272:C1450-C1456.
44. Frick KK and Bushinsky DA. Chronic metabolic acidosis reversibly inhibits extracellular matrix gene expression in mouse osteoblasts. *Am J Physiol*, 1998; 275:F840-F847.
45. Frick KK and Bushinsky DA. Metabolic acidosis stimulates RANKL RNA expression in bone through a cyclo-oxygenase-dependent mechanism. *J Bone Miner Res*, 2003; 18:1317-1325.
46. Krieger NS, Parker WR, Alexander KM and Bushinsky DA. Prostaglandins regulate acid-induced cell-mediated bone resorption. *Am J Physiol*, 2000; 279:F1077-F1082.
47. Amanzadeh J, Gitomer WL, Zerwekh JE, Preisig PA, Moe OW, Pak CYC and Levi M. Effect of high protein diet on stone-forming propensity and bone loss on rats. *Kidney Int*, 2003; 64:2142-2149.
48. Lemann J Jr. Relationship between urinary calcium and net acid excretion as determined by dietary protein and potassium: a review. *Nephron*, 1999; 81(Suppl 1):18-25.
49. Lemann J Jr, Litzow JR and Lennon EJ. Studies of the mechanism by which chronic metabolic acidosis augments urinary calcium excretion in man. *J Clin Invest*, 1997; 46:1318-1328.
50. Sutton RA, Wang NL and Dirks JH. Effects of metabolic acidosis and alkalosis on sodium and calcium transport in the dog kidney. *Kidney Int*, 1999; 15:520-533.
51. Friedman PA and Gesek FA. Calcium transport in renal epithelial cells. *Am J Physiol*, 1993; 264:F181-F198.
52. Sakhaee K. The effect of diuretics on calcium metabolism: physiologic and clinical effects. In: *Diuretic Agents: Clinical Physiology and Pharmacology*: Seldin DW, Geibisch G eds, 1st ed. 1997; pp 595-609. Academic Press, California.
53. Hoenderop JG, Nilius B and Bindles RJ. Calcium absorption across epithelia. *Physiol Rev*, 2005; 85:373-422.

54. Yeh B-Il, Sun T-J, Lee JZ, Chen H-H and Huang C-L. Mechanism and molecular determinant for regulation of rabbit transient receptor potential type 5 (TRPV5) channel by extracellular pH. *J Biol Chem*, 2003; 278:51044-51052.
55. Yeh B-Il, Kim YK, Jabbar W and Huang C-L. Conformational changes of port helix coupled to gating of TRPV5 by protons. *EMBO J*, 2005; 24:3224-3234.
56. Nijenhuis T, Renkema KY, Hoenderop JG and Bindels RJ. Acid-base status determines the renal expression of Ca²⁺ and Mg²⁺ transport proteins. *J Am Soc Nephrol*. 2006; Jan 18; [Epub ahead of print]
57. Lee SW, Russell J and Avioli LV. 25-hydroxycholecalciferol to 1,25-dihydroxycholecalciferol: conversion impaired by systemic metabolic acidosis. *Science*, 1976; 195:994-996.
58. Greenberg AJ, McNamara H and McCrory WW. Metabolic balance studies in primary renal tubular acidosis: effects of acidosis on external calcium and phosphorus balances. *J Pediatr*, 1966; 69:610-618.
59. Coe FL, Firpo JJ Jr., Hollandworth DL, Segil L, Canterbury JM and Reiss E. Effect of acute and chronic metabolic acidosis on serum immunoreactive parathyroid hormone in man. *Kidney Int*, 1975; 8:262-273.
60. Krout JA, Gordon EM, Ransom JC, Horst R and Slatopolsky E. Effect of chronic metabolic acidosis on vitamin D metabolism in humans. *Kidney Int*, 1983; 24:644-648.
61. Hess B, Ackermann D, Essig M, Takkineneger R and Jaeger P. Renal mass and serum calcitriol in male idiopathic calcium renal stone formers: role of protein intake. *J Clin Endocrinol Metab*, 1995; 80:1916-1921.
62. Krapt R, Vetsch R, Vetsch W and Hulter HN. Chronic metabolic acidosis increases the serum concentration of 1,25-dihydroxyvitamin D in humans by stimulating its production rate. Critical role of acidosis-induced renal hypophosphatemia. *J Clin Invest*, 1992; 90:2456-2463.
63. Lemann J Jr, Litzow JR and Lennon EJ. The effect of chronic acid loads in normal man: further evidence for the participation of bone mineral in the defense against chronic metabolic acidosis. *J Clin Invest*, 1966; 45:1608-1614.
64. Weber HP, Gray RW, Dominguez JH and Lemann J Jr. The lack of effect of chronic metabolic acidosis on 25-OH-vitamin D metabolism and serum parathyroid hormone in humans. *J Clin Endocrinol Metab*, 1976; 43:1047-1055.
65. Kerstetter JE, O'Brien KO and Insogna KL. Dietary protein affects intestinal calcium absorption. *Am J Clin Nutr*, 1998; 68:859-865.
66. Kerstetter JE, O'Brien KO, Caseria DM, Wall DE and Insogna KL. The impact of dietary protein on calcium absorption and kinetic of bone turnover in women. *J Clin Endocrinol Metab*, 2005; 90:26-31.
67. Geibel JP, Wagner CA, Caroppo R, Qureshi I, Gloeckner J, Manuelidis L, Kirchhoff P and Radebold K. The stomach divalent ion-sensing receptor SCAR is a modulator of gastric acid secretion. *J Biol Chem*, 2001; 276:39549-39552.
68. Ray JM, Squires PE, Curtis SB, Meloche MR and Buchan AM. Expression of calcium-sensing receptor on human antral gastrin cells in culture. *J Clin Invest*, 1997; 99:2328-2333.
69. Avery I, Anderson EJ and Neer RM. Lowering dietary protein to U.S. recommended dietary allowance levels reduces urinary calcium excretion and bone resorption in young women. *J Clin Endocrinol Metab*, 2004; 89:3801-3807.

70. Reddy ST, Wang CY, Sakhaee K, Brinkley L and Pak CYC. Effect of low-carbohydrate high-protein diets on acid-base balance, stone-forming propensity and calcium metabolism. *Am J Kidney Dis*, 2002; 40:265-274.
71. Alpern RJ and Sakhaee K. The clinical spectrum of chronic metabolic acidosis: homeostatic mechanisms produce significant morbidity. *Am J Kidney Dis*, 1997; 29:291-302.
72. Relman AS, Lennon EJ and Lemann J Jr. Endogenous production of fixed acid and the measurement of the net balance of acid in normal subjects. *J Clin Invest*, 1961; 40:1621-1631.
73. Kurtz I, Maher T, Hulter HN, Schambelan M and Sebastian A. Effect of diet on plasma acid-base composition in normal humans. *Kidney Int*, 1983; 24:670-680.
74. Frassetto LA, Morris RC Jr. and Sebastian A. Effect of age on blood acid-base composition in adult humans: role of age-related renal functional decline. *Am J Physiol*, 1996; F1114-F1122.
75. Levi M and Rowe JW. Renal function and dysfunction in aging. In: Seldin DW Geibisch G (eds): *The Kidney: Physiology and Pathophysiology* (ed 2). New York, NY, Raven, 1992; pp 3433-3456.
76. Albright F, Smith PH and Richardson AM. Postmenopausal osteoporosis. *JAMA*, 1941; 116:2456-2474.
77. Thissen JP, Ketelslegers JM and Underwood LE. Nutritional regulation of the insulin-like growth factors. *Endocr Rev*, 1994; 15:80-101.
78. Langdahl BL, Kassem M, Moller MK and Eriksen EF. The effects of IGF-I and IGF-II on proliferation and differentiation of human osteoblasts and interactions with growth hormone. *Europ J Clin Invest*, 1998; 28:176-183.
79. McCarthy TL, Centrella M and Canalis E. Insulin-like growth factor (IGF) and bone. *Connective Tissue Res*, 1989; 20:277-282.
80. Mohan S, Strong DD, Lempert UG, Tremollieres F, Wergedal JE and Baylink DJ. Studies on regulation of insulin-like growth factor binding protein (IGFBP)-3 and IGFBP-4 production in human bone cells. *Acta Endocrinol*, 1992; 127:555-564.
81. Sugimoto T, Nishiyama K, Kuribayashi F and Chihara K. Serum levels of insulin-like growth factor (IGF) I, IGF-binding protein (IGFBP)-2 and IGFBP-3 in osteoporotic patients with and without spinal fractures. *J Bone Miner Res*, 1997; 12:1272-1279.
82. Zhang M, Xuan S, Bouxsien ML, Von Stechow D, Akeno N, Faugere MC, Malluche H, Zhaoc S, Rosen CJ, Efstratiadis A and Clemens TL. Osteoblast-specific knockout of insulin growth factor (IGF) receptor gene reveals an essential role of IGF signaling. *J Biol Chem*, 2002; 277:44005-44012.
83. Oxlund H, Barckman M, Ortaft G and Andreassen TT. Reduced concentrations of collagen cross-links are associated with reduced strength of bone. *Bone*, 1995; 17 (suppl 4):365-371.
84. Ammann P, Bourrin S, Bonjour J-P, Meyer J-M and Rizzoli R. Protein undernutrition-induced bone loss is associated with decreased IGF-I levels and estrogen deficiency. *J Bone Miner Res*, 2000; 15:683-690.
85. Dayer R, Rizzoli R, Kaelin A and Ammann P. Low protein intake is associated with impaired titanium implant osseointegration. *J Bone Miner Res*, 2006; 21:258-264.

86. Schurch MA, Rizzali R, Slosman D, Vadas L, Vergraude P and Bonjour JP. Protein supplements increase serum insulin growth factor-I levels and attenuate proximal femoral bone loss in patients with recent hip fracture. *Ann Intern Med*, 1998; 128:801-809.
87. Bushinsky DA. Metabolic alkalosis decreases bone calcium efflux by suppressing osteoclasts and stimulating osteoblasts. *Am J Physiol*, 1996; 271:F216-F222.
88. Sakhaee K, Alpern RJ, Jacobson HR and Pak CYC. Contrasting effects of various potassium salts on renal citrate excretion. *J Clin Endocrinol Metab*, 1991; 72:396-400.
89. Sebastian A, Harris ST, Ottaway JH, Todd KM and Morris C Jr. Improved mineral balance and skeletal metabolism in postmenopausal women treated with potassium bicarbonate. *N Engl J Med*, 1994; 330:1776-1781.
90. Sellmeyer DE, Stone KL, Sebastian A and Cummings SR for the Study of Osteoporotic Fractures Research Group. A high ratio of dietary animal to vegetable protein increases the rate of bone loss and the risk of fracture in postmenopausal women¹⁻³. *Am J Clin Nutr*, 2001; 73:118-122.
91. Sakhaee K, Maalouf NM, Abrams SA and Pak CYC. Effects of potassium alkali and calcium supplementation on bone turnover in postmenopausal women. *J Clin Endocrinol Metab*, 2005; 90:3528-3533.
92. Maurer M, Riesen W, Muser J, Hulter HN and Karpp R. Neutralization of western diet inhibits bone resorption independently of K intake and reduces cortisol secretion in humans. *Am J Physiol*, 2003; 284:F32-F40.
93. Frassetto L, Morris C Jr. and Sebastian A. Long-term persistence of the urine calcium-lowering effect of potassium bicarbonate in postmenopausal women. *J Clin Endocrinol Metab*, 2005; 90:831-834.
94. Rafferty K, Davies KM and Heaney RP. Potassium intake and the calcium economy. *J Am Coll Nutr*, 2005; 24:99-106.
95. Sakhaee K, Nicar MJ, Glass K, Zerwekh JE and Pak CYC. Reduction in intestinal calcium absorption by hydrochlorothiazide in postmenopausal osteoporosis. *J Clin Endocrinol Metab*, 1984; 59:1037-1043.