MAST CELLS AND MASTOCYTOSIS:

Developments Over The Last Decade

Southwestern Medical Center Internal Medicine Grand Rounds November 11, 1993

Rebecca S. Gruchalla, M.D., Ph.D.

INTRODUCTION

Mastocytosis is an uncommon clinical disorder that is characterized by an increased number of tissue mast cells. While the cutaneous manifestations of this disorder have been recognized for over a century it has been only over the last decade, with our increased awareness of the importance of the mast cell in health and disease, that interest in this disorder has heightened.

The effector role of mast cells in allergic diseases is well-recognized. In addition, however, it is now known that these cells participate in a variety of processes, both physiologic and pathophysiologic. While most considerations of mast cell function have focussed on the pathologic role of the inflammatory mediators contained within the mast cell granules, the recent demonstration of the production, upon activation, of cytokines and growth factors suggests a role for mast cells in the growth and development of a variety of cells. Also, the strategic location of mast cells - in the lymphoid tissues, around blood vessels and nerves, throughout connective tissues and in tissues that interface with the environment - adds further evidence that these cells fulfill a number of regulatory functions. Indeed, evidence exists to suggest that mast cells not only provide a critical defense against certain parasites, but in addition, they also have been shown to play a role in the regulation of lymphoid responses, stimulation of connective tissue repair and maintenance of the vasculature. While much of our knowledge over the last decade has stemmed from work done in animals, studies of patients with mastocytosis has allowed us to gain enormous insight into the biologic properties and functions of the mast cell and mast cell disease.

HISTORICAL PERSPECTIVE

Nettleship and Tay (1) are credited with the first description of mastocytosis. In 1869 they reported a two year old girl with persistent, hyperpigmented lesions that spontaneously urticated. It was not until 1877, however, that mast cells actually were described. Upon staining connective tissue with aniline dyes, Paul Ehrlich (2) found that certain cells possessing cytoplasmic granules stained metachromatically. He chose the term mastzellen, derived from the German word "to chew", to describe these cells since he believed the presence of the granules resulted from overfeeding. In addition to being credited with their initial description, Ehrlich also described the process of degranulation and noted the association of mast cells with blood vessels, inflamed tissues, nerves and neoplastic tissue. Subsequently, in 1878 Sangster (3) described a patient with a pruritic, pigmented urticarial rash and he labeled this cutaneous eruption urticaria pigmentosa. The importance of mast cells in this disorder was not discovered, however, until 1887 when Unna and Beitrage (4) demonstrated the presence of large numbers of dermal mast cells in the skin lesions of affected patients. It was not until 1936, almost 50 years later, that Sezary and colleagues (5) coined the term "mastocytosis". Subsequently, Ellis (6) who formally showed that other organs, including the liver, spleen, thymus, bone marrow, pancreas and lymph nodes may be affected by this disease process.

THE MAST CELL

Morphology and Distribution

Mature human mast cells may be round, spindle-shaped or spiderlike and are usually 9 to 12 μm in diameter. The plasma membrane contains elongated folds and may appear ruffled by transmission electron microscopy. Unlike the lobulated nucleus of the basophil, the mast cell nucleus is round or oval and is elliptically situated in the cell. The most characteristic feature of the mast cell is the beautiful array of metachromatic granules. These may comprise a large amount of the dry weight of the cell (7) and as much as half its volume (8). Ultrastructurally, there are differences in granule morphology among species. While rodent mast cell granules have an amorphous appearance, human mast cell granules demonstrate a variety of patterns.

Mast cells are positioned in connective tissue and mucosal surfaces at potential sites of entry of noxious substances. The tissues in which they predominate include those that interface with the outside environment where allergic reactions most often occur and include the skin, gut, conjunctivae and respiratory tract. While bone and cartilage have few mast cell numbers, large numbers are found around blood vessels and in the alveolar wall, dermis and mucosa of the nose, bowel and conjunctivae.

Origin and Development

Only recently has the origin of mast cells been clearly delineated. Due to their connective tissue location there was a great deal of confusion regarding their derivation. After their discovery, a number of proposed cells of origin were suggested: T cells, fibroblasts, mesenchymal cells, plasma cells, histiocytes, endothelial cells, and degenerate cells (9-11, reviewed in 7). It was also thought that since mast cells and basophils both contain metachromatic granules, histamine, and high affinity IgE receptors, these two cells were intimately related in lineage. We now know that while these cells are similar in some respects, they can easily be distinguished by their nuclear morphology, mediator content, location and response to activating agents. Moreover, basophils are terminally-differentiated cells that, after maturation in the bone marrow, are released into the peripheral blood. In contrast, it is now clear that mast cells are released from the bone marrow as progenitors to complete their differentiation in the peripheral tissues.

The bone marrow origin of mast cells was first clearly demonstrated by Kitamura and colleagues (12,13) in a series of *in vivo* reconstitution experiments using genetically mast cell-deficient mutant mice and their genetically normal littermates. Mutations at the dominant white-spotting (W) locus on mouse chromosome 5, and the steel (SI) locus on chromosome 10 affect several critical developmental processes including gametogenesis, pigmentation, and hematopoiesis. In addition, Kitamura et al. (12,13) demonstrated that mutations at either of these loci also dramatically affected mast cell

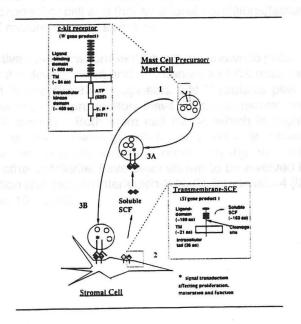
development. They found that transplantation of congenic +/+ bone marrow cells corrected the mast cell defect of W/W' mice but not SI/SI^d mice (12). The mast cell defect of W/W' mice was also corrected by transplanted bone marrow from SI/SI^d mice (13). Taken together, these findings demonstrate that the W/W' mutants have a deficiency within the mast cell precursor population, whereas the SI/SI^d mutants have normal bone marrow progenitors for mast cells but a defective microenvironment which is necessary for normal mast cell differentiation.

The explanation on a molecular level for these mast cell deficiencies has been provided within the last few years when the gene products of the *W* and the *SI* loci were cloned (14-20). It was demonstrated that the c-*kit* proto-oncogene, which encodes c-*kit*, a tyrosine kinase receptor expressed on mast cells, maps to the *W* region. Identification of the c-*kit* receptor as the *W* gene product subsequently set the stage for the elucidation of its ligand. Within a short time, three groups reported simultaneously the cloning and characterization of a new hematopoietic growth factor that represented the product of the *SI* locus and a ligand for the c-*kit* receptor (17,18,20-24). This factor which has been termed stem cell factor (SCF) (23), kit ligand (KL) (25), mast cell growth factor (MGF) (21), and Steel factor (26) is highly expressed on fibroblasts and stromal cells and has been found to play a major role in mast cell proliferation and maturation.

Figure 1

c-kit Receptor/c-kit Receptor Ligand (SCF) Interactions in Mast Cell Development

(1) Both mast cell precursors and cells in the mast cell lineage express the c-kit receptor on their surface. The structure of the receptor includes a transmembrane portion (TM), an ATP binding site, and the tyrosine autophosphorylation (T-P*) site. (2) A stromal cell expresses transmembrane SCF, including the cleavage site for generation of soluble SCF. (3) Interaction of the mast cell precursor or mast cell with the soluble (A) or transmembrane (B) SCF induces signal transduction in the mast cell precursor/mast cell (Ref 27).



While *in vitro* studies have expanded our knowledge regarding the growth and differentiation of murine mast cells, similar studies of human mast cells have been hampered by the inability to culture sufficient mast cell numbers from human blood and bone marrow. Recently, however, small numbers of mast cells were cultured and examined using a technique whereby human bone marrow-derived mononuclear cells were suspended over agar (28) or agarose (29) in the presence of human recombinant interleukin-3 (rhlL-3). The cells produced stained histochemically like mast cells and possessed high affinity IgE receptors and contained the neutral protease tryptase within cytoplasmic granules, all characteristic features of human mast cells (30).

Despite the demonstration that human mast cells could arise from bone marrow-derived mononuclear cells, their derivation remained unclear. Reports were conflicting. While human skin, colon, and spleen mast cells were not found to possess T-cell, B-cell, or monocyte markers (21) they were found to possess surface markers associated with a late stage of monocyte/macrophage but not basophil differentiation (32). In experiments designed to remove committed T-cell (CD2), B-cell (CD19 and CD20), eosinophil and macrophage (CD14) precursors from human bone marrow, bone marrow-derived mononuclear cells continued to give rise to mast cells. However, when CD34+pluripotent progenitor cells were removed from the culture mast cells did not arise. Additional experiments, using highly enriched CD34+ cells cultured in the presence of rhIL-3 over agarose, yielded mast cells in small numbers (33). The granules that were produced contained homogenous material, not the scroll or lattice patterns characteristic of mature mast cells. Taken together, these studies demonstrate that human mast cells originate from a pluripotent CD34+ progenitor cell and that additional conditions/factors are required for the development of mature granule structure.

Both fibroblasts and a connective tissue environment have been shown to provide the additional conditions needed for the development and maintenance of the mast cell phenotype both *in vitro* (34-36) and *in vivo* (37-39). It appears that fibroblasts play a critical role late in mast cell differentiation, after progenitors leave the bone marrow and become committed to the mast cell lineage. Both stem cell factor, which is highly expressed on the fibroblast surface, as well as nerve growth factor, another fibroblast-derived growth factor, have been shown to be important in promoting the mast cell phenotype. In addition, a variety of other cytokines have been shown to be involved in mast cell proliferation and differentiation and include interleukin-3 (IL-3), interleukin-4 (IL-4), interleukin-9 (IL-9) and interleukin-10 (IL-10).

sory granules of the MC, cells have variable shapes and are smaller than tribus of

MAST CELL PROLIFERATION AND DIFFERENTIATION

Confirmation of bone marrow origin: CD34+ progenitor cell

Mast cell growth factors:

Interleukin-3
Interleukin-4
Interleukin-9
Interleukin-10
Nerve growth factor
Stem cell factor (c-kit Ligand)

Heterogeneity

Two major types of mast cells have been identified in both rodents and humans and, as stated in a recent review by Schwartz and Huff (40), "recognition of this complexity is crucial for understanding mast cell biology and, potentially, for treating mast cell-associated disease" (41). "Mucosal" and "connective tissue", the most common terms used to describe the two mast cell types, are phenotypic descriptions. Unlike rodent mast cells, human mast cell types can not be differentiated from each other by their histamine or heparin proteoglycan concentration. However, they can be distinguished by their neutral protease composition (42,43): MC_T contain tryptase alone and correspond to the mucosal mast cell phenotype (MMC) while MC_{TC} contain tryptase, chymase, carboxypeptidase and a cathepsin G-like enzyme and correspond to the connective tissue phenotype (CTMC).

In histologically normal tissues MC_T cells are found predominantly in the alveoli of the lung and in the intestinal mucosa, while MC_{TC} cells are found mainly in the skin and intestinal submucosa. Despite the fact that one particular mast cell type may predominate in a tissue, a mixture of these two cell types is usually seen. For this reason, MC_T and MC_{TC} cells can not be differentiated from each other based upon their location alone.

Ultrastructurally, these two cell types differ as well. Studies have shown that the secretory granules of the MC_T cells have variable shapes and are smaller than those of the MC_{TC} cells. The majority of the MC_T cells possess granules with discrete scroll-like structures whereas grating and lattice structures predominate in the MC_{TC} cells.

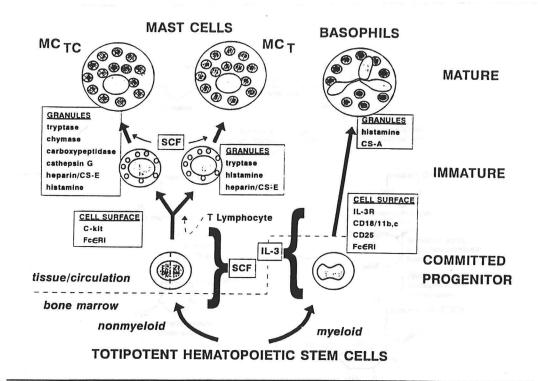
Characteristics of Human Mast Cell Types*

Characteristic	MC _{TC} Cell	MC _T Cell
Tissue Distribution		
Skin	++	
Intestinal submucosa	++	+
Intestinal mucosa	+	++
Alveolar wall	•	++
Bronchi/bronchioles	+	++
Nasal mucosa	++	++
Conjunctiva	++	+
Synovium	++	
Protease content	Tryptase, chymase cathepsin G-like protease, carboxypeptidase	Tryptase
T lymphocyte dependency	No	Yes
Granule morphology	Grating/lattice	Complete
	Complete scroll	scroll rich
	poor	

The issue of whether or not two distinct mast cell lineages exist for human MC_T and MC_{TC} types is still unresolved. However, evidence exists to suggest that these cell types may develop along distinct pathways. Both humans with inherited combined immunodeficiency and those with the acquired immunodeficiency syndrome have marked decreases in their MC_T cell concentrations in the bowel, while the MC_{TC} cell numbers and distribution are unchanged (30). This finding suggests that MC_T cells develop along a pathway that is dependent upon the existence of functional T lymphocytes while MC_{TC} cell development is T lymphocyte-independent.

Figure 4

Developmental Pathways for Human Mast Cells and Basophils*

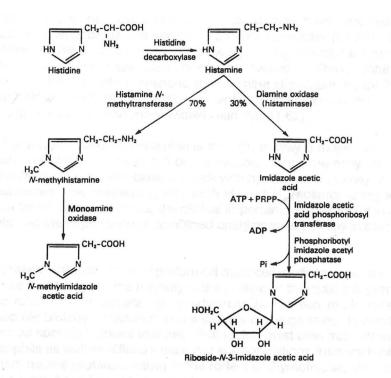


From Schwartz and Huff, Biology of Mast Cells and Basophils in Allergy-Priniciples and Practice, 1993.

Preformed Mast Cell Mediators

Histamine, the sole, major biogenic amine in human mast cells, is formed from histidine (45) by histidine decarboxylase (46). It is present in concentrations of 4-10 pg/cell and is the only preformed mediator of human mast cells that has potent, direct vasoactive and spasmogenic effects. Histamine is released within 30-45 seconds after activation and produces its effects on the vasculature within 1-5 minutes (47). After it is released, histamine is metabolized by one of two pathways (48,49): methylation (70%) or oxidation (30%). Metabolism by diamine oxidase (histaminase) yields imidazole acetic acid, while metabolism by *N*-methyltransferase yields *N*-methylhistamine and *N*-methylimidazole acetic acid. The methylation products are excreted by the kidney and may be measured in the urine as a measure of endogenous histamine release. Metabolism occurs within minutes of release suggesting that histamine acts locally and not on distant target tissues.

HISTAMINE SYNTHESIS AND METABOLISM



The biologic effects of histamine are wide-ranging and are mediated through the activation of three receptor subtypes, termed H_1 and H_2 and H_3 receptors. These receptors are present on a variety of tissues and circulating cells, and their activation results in a variety of biologic responses. H_1 receptor activation results in airway and gastrointestinal smooth muscle contraction, increased capillary permeability and prostaglandin and thromboxane production. Activation of H_2 receptors also causes increased vasopermeability but, in addition, causes mucous production and gastric acid secretion. Other H_2 -mediated effects include inhibition of: basophil histamine release (50), delayed hypersensitivity skin test responses (51) and *in vitro* generation of cytotoxic T lymphocytes (52). Stimulation of H_2 receptors on neutrophils and eosinophils results

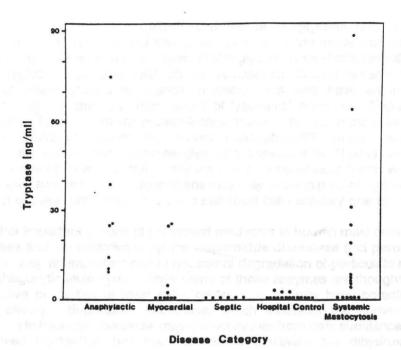
in the inhibition of lysosomal enzyme release from neutrophils and modulation of migration of both cell types (53). H_2 receptor activation of lymphocytes has also been demonstrated to inhibit several B-cell responses including: antigen-stimulated B-cell maturation (54), secretion of antibody by plasma cells (53) and antibody production by human mononuclear cells (55). Activation of H_2 receptors on T lymphocytes leads to the production of cytokines (56).

Recently, both immunochemical and biochemical studies have revealed that histamine is present in nerve endings and serves as a neurotransmitter (57-59). While the postsynaptic receptors have been shown to be the $\rm H_1$ and $\rm H_2$ receptor subtypes, a class of presynaptic receptors have also been discovered. These inhibitory autoreceptors, designated $\rm H_3$ receptors, respond to histamine stimulation by inhibiting both its synthesis and release (59,60). In addition, recent studies have revealed that the functions of this receptor may be even more widespread (59,61,62).

Due to its wide range of effects, histamine is thought to play a major role in the clinical manifestations resulting from mast cell degranulation. Since the early 1900's it has been known that histamine infusion causes shock with hypotension (63,64), as well as flushing and headaches. By pretreating with both $\rm H_1$ and $\rm H_2$ receptor antagonists these symptoms can be inhibited, indicating the clinical importance of histamine in mast cell mediated events and the importance of combined antihistamine therapy in blocking its systemic effects.

Neutral proteases, another group of preformed mast cell mediators, are a group of proteolytic enzymes that comprise the majority of the protein of the mast cell granule. While their biologic roles remain unclear, great advances have been made recently regarding their molecular biology, structures, and substrate cleavage sites. In addition, these enzymes serve as specific markers that help distinguish mast cells from other cell types including basophils as well as different mast cell subpopulations from each other. While the predominant neutral protease activity in the rodent is chymotryptic, the neutral proteases in humans have been found to be tryptic in nature.

Tryptase is the major enzyme accounting for the trypsinlike activity demonstrated by human mast cells (65,66). While the biologic role of tryptase has not yet been clearly demonstrated, it may be important in fibrinogenolysis and latent collagenase activation (67,68). Tryptase is found in both MC_{TC} and MC_{T} cells and is released along with histamine during the degranulation process (69). Negligible amounts of tryptase have been found in basophils while other cells possess none. Thus, this enzyme is a specific marker of human mast cells. Taking advantage of this important and useful finding, Schwartz and colleagues (70-75) recently developed a very sensitive and specific assay to measure tryptase in a variety of biological fluids. By demonstrating that this enzyme is elevated in patients with anaphylaxis and mastocytosis but not those with other systemic disorders, they documented its usefulness as a specific indicator of systemic mast cell activation (70).



From Schwartz et al., Tryptase Levels as an Indicator of Mast-Cell Activation in Systemic Anaphylaxis and Mastocytosis. N Engl J Med 316:1622, 1987 (Ref 70).

Chymase is the major enzyme that accounts for the chymotryptic activity in human cutaneous mast cells. This enzyme has been purified (76) and recently, the gene has been cloned (77). Unlike tryptase, chymase is found in a subpopulation of mast cells, the MC_{TC} subtype, localized to the gastrointestinal submucosa and the skin. Like tryptase, the biologic activities of chymase are not well-understood. Potential interesting findings include the ability of chymase to: efficiently convert angiotensin I to angiotensin II (78), inactivate bradykinin (79) and attack the lamina lucida of the basement membrane at the dermal-epidermal junction of human skin (80). It is thought that this latter function may account for the blister formation seen in infants with systemic mastocytosis.

The most recent neutral protease that has been cloned and characterized from human mast cells is carboxypeptidase (81-90). It, along with cathepsin G-like protease, is present in MC_{TC} cells and helps to extend the compositional differences between the two human mast cell subtypes.

The highly sulfated and negatively charged **proteoglycans** present in the secretory granules of mast cells and basophils account for the metachromasia seen when these cells are stained with basic dyes. Proteoglycans consist of a central protein core from which glycosaminoglycan side chains are attached. While they serve as major constituents of tissue ground substance, proteoglycans also have an important intracellular role as the structural component of lysosomal granules. Heparin and chondroitin sulfate E are the major proteoglycans found in human mast cells (91,92) while chondroitin sulfate A predominates in human basophils (93). At the present time the biologic functions of the mast cell proteoglycans are speculative. Their ability to bind to histamine as well as to the neutral proteases and acid hydrolases found within the secretory granules suggests that proteoglycans may play a role in promoting the uptake and packaging of these preformed mediators into mast cell secretory granules.

Two other important groups of preformed mediators in human mast cells are the acid hydrolases and the oxidative enzymes, superoxide dismutase and peroxidase. These enzymes play an important role in lysosomal degradation of particulate material ingested by phagocytic leukocytes. While some of these enzymes are thought to add to the destructive properties of mast cells, others may actually have potential anti-inflammatory effects. Superoxide dismutase, which facilitates the conversion of superoxide anion to hydrogen peroxide, may protect tissues from toxic substances, while mast-cell derived peroxidase has been shown to inactivate the dihydroxy- and sulfidopeptide leukotrienes (94).

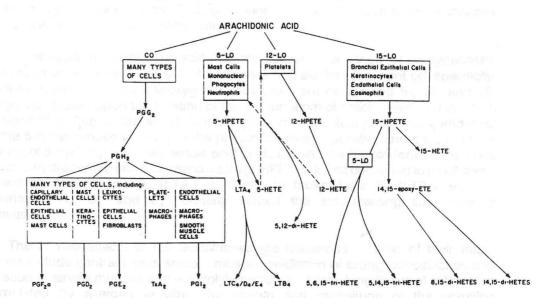
The final group of preformed mast cell mediators are the **chemotactic factors**. A neutrophilic chemotactic factor, thought to arise from mast cells, has been demonstrated in both patients with atopic asthma (95-97) after airway challenge with inhaled allergen and those with physical urticarias (98-100) after provocation of the skin. In addition, eosinophilic chemotactic factors have been demonstrated as well. When stimulated *in vitro* by antihuman IgE human lung tissue has been shown to release an eosinophil chemotactic factor (101). This factor was localized to the mast cell and was termed eosinophil chemotactic factor of anaphylaxis (ECF-A) (102). The potential clinical relevance of this finding is seen in patients with cutaneous mastocytosis where oftentimes the infiltrating cells in the skin include eosinophils (103).

Newly-Generated Mediators

Upon activation arachidonic acid is released from intracellular lipid stores. It may undergo oxidative metabolism via the cyclooxygenase pathway to yield prostaglandins (PGs) and thromboxanes (TXs), or via the lipoxygenase pathway to yield leukotrienes.

Figure 7

ARACHIDONIC ACID METABOLISM



^{*} From Valone, Boggs and Goetzl, Lipid Mediators of Hypersensitivity and Inflammation in Allergy-Principals and Practice, 1993. CO=cyclooxygenase, LO=lipoxygenase (Ref 104).

Cyclooxygenase catalyzes the incorporation of oxygen into arachidonic acid yielding the unstable intermediate, prostaglandin G_2 which is subsequently transformed into prostaglandin H_2 . PGH_2 is then converted into **prostacyclin** (PGI_2), **thromboxane** (TXA_2) and the primary prostaglandins, **prostaglandin** D_2 (PGD_2), **prostaglandin** E_2 (PGE_2), and **prostaglandin** E_2 0 by a variety of enzymatic pathways that are highly cell-specific (105-107).

The most abundant cyclooxygenase product of human mast cells is PGD₂. While it acts similarly to histamine, it acts independently of histamine receptors. In the lung, PGD₂ acts to produce bronchoconstriction and pulmonary artery vasoconstriction while systemic release of this mediator causes marked hypotension (108). A metabolite of PGD₂ has been detected in the urine of patients with systemic mastocytosis but not that of normal persons (108). That PGD₂ played a role in generating hypotension in these patients was suggested when marked clinical improvement was demonstrated after the administration of aspirin, an inhibitor of PGD₂ generation. An interesting, but unexplained observation, is the finding that patients with systemic mastocytosis and high levels of mast cell activation experience rhinorrhea, bronchorrhea and urticaria, but not bronchoconstriction.

 PGD_2 has several other functions, as well. It is chemokinetic for human neutrophils (109), it potentiates LTB_4 -mediated accumulation of neutrophils in the skin (110), and it augments histamine release from human basophils (111). In addition, PGD_2 , like PGI_2 , inhibits the aggregation of human platelets (112). Inhibitors of PGD_2 synthetase and the receptor for PGD_2 are being developed and may prove clinically useful for inhibiting the effects of this mediator *in vivo* (113).

Metabolism of arachidonic acid may also occur by a variety of lipoxygenases. These enzymes are a family of cytosolic enzymes that exhibit prominent cell specificity. The three major mammalian lipoxygenase enzymes are designated 5-, 12- and 15-lipoxygenase based upon the location of the carbon atom of arachidonic acid at which a molecule of oxygen is introduced. In neutrophils, mononuclear phagocytes, and mast cells the 5-lipoxygenase pathway is the primary pathway operative and it leads to the formation of 5-hydroxy-eicosatetraenoic acid (5-HETE) and the **leukotrienes**. The initial product, 5-hydroperoxy-eicosatetraenoic acid (5-HPETE), is first converted to an unstable intermediate, **leukotriene** A_4 (LTA $_4$) and then to **leukotriene** B_4 (LTB $_4$) and the **leukotriene** C_4 , C_4 , and C_4 (formerly termed the slow-reacting substance of anaphylaxis).

The *in vivo* effects of the leukotrienes are numerous. Three of their major functions include contraction of smooth muscle, mediation of bronchoconstriction and stimulation of airway mucous and electrolyte secretion. When lung tissue of asthmatics is stimulated by allergen *in vitro*, the course and magnitude of the resulting bronchoconstriction has been attributed to the large quantities of LTC₄, LTD₄, and LTE₄ that are produced. This effect can be abolished by pretreatment of the tissue with inhibitors of the 5-lipoxygenase pathway but not with inhibitors of cyclooxygenase demonstrating that the leukotrienes are more potent mediators of airway smooth muscle contraction than are the cyclooxygenase products (114). Leukotriene antagonists are in the process of development for human use and should allow a more clear assessment of the role of leukotrienes in airway pathophysiology.

Another potent newly-generated mediator, platelet activating factor (PAF), is an ether-linked phospholipid with the general structure alkylacetyl-glyceryletherphosphorylcholine (AGEPC). While it was initially discovered as a proaggregatory, basophil-derived mediator of anaphylaxis in the rabbit, PAF is now known to be a product of many cell types and has a broad spectrum of activities (115,116) including bronchoconstriction. The role that PAF plays in mast cell mediated disease remains unclear. While purified human lung mast cells do produce PAF with stimulation, little of this is released from cells (117).

Mast Cell Cytokines

Until the last several years, the participation of mast cells in both physiologic and pathophysiologic events was thought to be solely dependent upon their preformed and newly generated mediators. It is now clear that mast cells are also capable of synthesizing cytokines, another class of mediators that play a pivotal role in the pathophysiology of allergic and immunological diseases.

Cytokines are proteins or glycoproteins that are synthesized and secreted by a variety of cells. The first categories of cytokines discovered were the interleukins, the interferons and colony stimulating factors. Generally, cytokine gene induction occurs in response to cellular injury or activation and the cells that produce them are widely distributed in the body. The biological effects are mediated by specific cytokine receptors located on the surface of target cells and include participation in cell growth, repair, inflammation and modulation of the immune response. Each cytokine molecule has multiple functions and often, more than one cytokine may possess a particular bioactivity (118). Importantly, cytokines do not act in isolation. They are involved in an intimate highly regulated network where they influence each other as well as numerous other biological processes (119).

Figure 8

SPECTRUM OF CYTOKINE FUNCTIONS'

Stem cell colony-stimulating factors
Lineage-specific cell growth and/or differentiation
Induction of MHC antigens
Cellular activation
Initiation of the acute-phase response
Enhancement of endothelial/leukocyte/lymphocyte adhesion
Promotion of chemotaxis
Modulation of cellular biological activities
Transcriptional induction
Arachidonic acid metabolism
Protein phosphorylation
Cytotoxic/tumoricidal activity
Antiviral activity
Induction of other cytokines and/or cytokine receptors

*From Costa et al., Mast Cell Cytokines in The Mast Cell in Health and Disease, 1993 (Ref 120).

The initial reports of mast cell cytokine production occurred in 1987 when it was found that rodent mast cells were found to express IL-4 mRNA (121) and a cytolytic factor thought to be tumor necrosis factor (TNF- α) (122). More recently, activated mast cells have been shown to transcribe macrophage inflammatory proteins 1- α and 1- β (MIP 1- α , MIP 1- β), monocyte chemotactic and activating factor (MCAF), IL-1,2,3,4,5,10, granulocyte/macrophage colony stimulating factor (GM-CSF), transforming growth factor- β (TGF- β) and interferon-gamma (IFN- γ) while they constitutively transcribe TNF- α and IL-6 (123). Preliminary studies suggest that rodent mast cell lines also express mRNA for SCF and possess membrane-bound SCF (40). In marked contrast to the immediate release of the preformed mediators and the release within minutes of arachidonic acid metabolites, induction of cytokine synthesis by mast cells occurs hours after mast cell activation.

While most of the cytokine studies have been conducted in rodent mast cells, human mast cells have also been shown to participate in cytokine synthesis and release. Human lung mast cells have been shown to produce IL-4 and human skin mast cells, both TNF- α and IL-4 (124-126). However, the physiologic role played *in vivo* by these mast-cell derived cytokines remains speculative. Treatment of cultured human skin with mast cell degranulating agents has resulted in the appearance of the endothelial adhesion molecule ELAM-1 on the adjacent microvasculature (125). The fact that this response could be blocked by pretreatment of the cultures with mast cell stabilizing drugs, as well as with antibodies to TNF- α , is consistent with a model in which IgE-mediated production and release of TNF- α stimulates the dermal endothelium to express an adhesion molecule that is important for endothelial-leukocyte interactions.

Using skin blister chambers, several investigators have shown that, upon antigen challenge of atopic individuals, IL-18, IL-3, GM-CSF, and IL-6 appear in blister fluid within 12 hours after challenge (127-129). In addition, increased cell numbers expressing mRNA for IL-3, IL-4, IL-5, and GM-CSF have been detected in allergen-challenged skin of atopic individuals (130). While it has not been proven that mast cells are the source of these cytokines, it is highly likely that they are contributors since it is known that the cutaneous late-phase reaction is generated after mast cells have been triggered by IgE-dependent mechanisms.

In light of its ability to produce multifunctional cytokines the mast cell can no longer be thought of as a simple effector cell. Its functions have expanded to include an important regulatory role of both the inflammatory and immune responses. However, it is important to recognize that, to date, most of the studies examining cytokine production by mast cells have been performed using rodent mast cell lines or primary mast cell cultures. The extent to which human mast cells produce and secrete cytokines remains to be determined.

CYTOKINES PRODUCED BY MAST CELLS AND BASOPHILS'

Cell type	<u>Cytok</u>	<u>ines</u>
Human	mRNA	Protein
Mast cells	TNF-α IL-4	TNF-α IL-4
Basophils		TNF-α IL-4
Rodent mast cells	TNF- α GM-CSF TGF- β SCF IFN- γ MIP-1 α , β MCAF	TNF-α
	MCAF IL-1,2,3,4, 5,6,10	IL-1,3,4,6

From Schwartz and Huff, Biology of Mast Cells and Basophils in Allergy-Principles and Practice, 1993 (Ref 40).

Mast Cell Activation

Activation of mast cells immunologically involves the cross-linkage, by multivalent allergen, of IgE that is bound to the high-affinity Fc ϵ receptor (Fc ϵ R1). This receptor contains four polypeptide chains, an α -, a β -, and two γ chains with the α -chain possessing the IgE binding site. Aggregation of as few as 1% of the 10 4 to 10 6 Fc ϵ R1 present per mast cell (131-133) results in exocytosis and the generation of newly-formed mediators. A number of sequential steps occur in exocytotic process. First, the granule swells as a result of the solubilization of the granule contents. This is followed by fusion

of the granular and plasma membrane and finally, expulsion of the granule contents.

Activation may also occur by nonimmunological means. It has been shown that human skin mast cells, unlike those from the lung, adenoid, tonsil or large intestine, release histamine after stimulation with substance P, VIP, somatostatin, compound 48/80, and morphine. Of interest, this reactivity does not parallel the phenotyping based on neutral protease content since intestinal mast cells, consisting of large numbers of the MC_{TC} subpopulation, are unresponsive to neuropeptide stimulation (134).

Recently, it has been shown that there are important differences between IgE-dependent and neuropeptide-induced mast cell activation. Degranulation by an IgE-dependent mechanism is a relatively slow process (occurring over 5 minutes) and results in the release of preformed mediators and the generation of the newly-formed mediators PGD₂ and LTC₄. In contrast, neuropeptide stimulation is rapid (occurring within 15 seconds) and leads to the release of preformed mediators only; PGD₂ and LTC₄ are not produced. When viewed by electron microscopy the degranulation process induced by immunological and nonimmunological activation appears identical. However, it is now known that the biochemical mechanisms involved in these two types of activation differ.

Enzymes that cause protein phosphorylation and dephosphorylation play an important role in signal transduction pathways linked to activation of cell surface receptors. Tyrosine and serine/threonine kinases and phosphatases all are important in regulating the events involved in cell growth and activation (135-138). Recently, it has been shown that tyrosine kinase is activated in RBL-2H3 rat mucosal mast cells resulting in the phosphorylation of a variety of proteins (139). Upon phosphorylation these proteins then bind to various intracellular signaling proteins. The importance of tyrosine kinase activity to coupled activation-secretion was demonstrated by the finding that inhibition of tyrosine kinase activity caused decreased histamine release in mast cells stimulated through Fc∈R1 (139-141). In contrast, if stimulation does not involve Fc∈R1 (nonimmunological stimulation), inhibition of tyrosine kinase does not diminish histamine release, while inhibition of GTP-binding proteins by pertussis toxin does (142-144). These results together suggest that signal transduction may occur by two different pathways depending upon the signalling source. Activation through Fc∈R1 appears to be associated, either directly or indirectly, with tyrosine kinase activation, while activation by neuropeptides and other nonimmunologic activators is initiated through a pertussissensitive G-protein.

When crosslinked by antigen, both the T cell receptor (TCR) and the Fc ϵ R1 on mast cells deliver an activation signal to their respective cells. This signal is mediated, in part, by the ζ chain of the T-cell receptor and by the γ chain of Fc ϵ R1. While these chains are encoded by different genes they are very similar in function. In fact, transvection studies have revealed that these two chains can be interchanged without loss of function. In addition, some T cells, including immature thymocytes, have been found to express the γ chain of the Fc ϵ R1 (145,146). Thus, it appears that Burnet (9,10)

may not have been wrong. Mast cells and T cells, indeed, may be related developmentally.

Figure 10

MAST CELL ACTIVATION Two Different Signal Transduction Pathways?

Immunologic activation: Fc∈R1

Role of tyrosine kinases

Nonimmunologic activation: substance P. compound 48/80, morphine

Role of pertussis-sensitive G-proteins

Mast Cell Secretagogues

In addition to activation by IgE, mast cells undergo receptor-mediated activation by the anaphylatoxins C3a, C4a and C5a (147). In addition, numerous other agents, with potential relevance to human disease, possess the capability of activating mast cells through nonimmunological mechanisms. Included in this group are: opioids (148), neuropeptides, drugs such as polymyxin B, amphotericin B, D-tubocurare, succinylcholine and iodinated contrast media (149), adenine nucleotides (150), hormones (151,152), and various physical stimuli (153,154). Also, inflammatory-cell-derived histamine releasing factors have been demonstrated to cause histamine release from human mast cells (Reviewed in 155). Thus, it appears that mast cells not only play a central role in allergic diseases and mastocytosis. The existence of numerous endogenous mast cell secretagogues suggests that these cells play an important role in normal physiologic processes as well.

MAST CELL SECRETORY AGONISTS

Immunologic mechanisms

IgE-mediated Anaphylatoxins Lymphokines

Non Immunologic mechanisms

Medications
Adenine Nucleotides
Hormones
Neuropeptides
Radiocontrast media
Dextran
Venoms
Physical Stimuli

adapted from Tharp, Medical Grand Rounds, July 1984 (Ref 156).

Clinical Manifestations of Mast Cell Secretion

Mast cell mediator release results in a variety of clinical manifestations. As Tharp (156) stated in Internal Medicine Grand Rounds in 1984, the signs and symptoms expressed "are dependent upon several variables that include: 1) the anatomical location and number of tissue mast cells stimulated, 2) the extent and type of mediators released by the cells, and 3) the portal of entry, concentration, and nature of the potential mast cell agonist." Rate of antigen presentation and efficiency of cross-linking of mast cell bound IgE determine whether or not the cell will be triggered to release its mediator contents. In addition, in nonanaphylactic states, the clinical symptoms will reflect the anatomic location of mast cell activation. Therefore, while air-borne allergens cause upper and lower respiratory tract symptoms, orally-ingested allergens often elicit symptoms in the gastrointestinal tract alone. Like patients with immediate hypersensitivity reactions, patients with mastocytosis often present with a constellation of symptoms reflective of mast cell activation at several anatomic sites.

CLINICAL MANIFESTATIONS OF MAST CELL SECRETION'

Anatomical Region	Clinical Manifestations
Upper Respiratory Tract	Congestion, rhinorrhea, laryngeal edema, hoarseness
Lower Respiratory Tract	Dyspnea, wheezing, cyanosis
Eye	Lacrimation, pruritus, suffusion
GI Tract	Nausea, vomiting, abdominal cramps, diarrhea
Heart	Tachycardia, arrhythmia, angina (?)
Skin	Pruritus, pain, flushing, urticaria, angioedema
Neurological	Headache, dizziness, syncope
*From Tharp, Medical Grand Rounds, 1984 (R	ef 156).

MASTOCYTOSIS

Mastocytosis is a rare disease that is characterized by increased mast cell numbers in the skin and other organs. As stated by Tharp (156), mastocytosis "represents a spectrum of clinical disorders that vary in regard to patient age, to the predominant clinical manifestations, and to the nature and extent of the mast cell proliferative process." Many patients have a benign, indolent form of the disease and, when properly medicated, live relatively normal lives. Others have a more aggressive form and, for these individuals, the prognosis is not so optimistic. The organ most frequently involved is the skin (cutaneous mastocytosis). However, many other organ systems may be involved, as well, including the liver, spleen, lymph nodes, gastrointestinal tract, bone marrow and skeletal systems. A myriad of symptoms occur, and these may be either localized to the organ system involved or systemic depending upon whether there is local or generalized mast cell mediator release.

While mastocytosis may occur at any age, in approximately 55% of cases, the onset is between birth and two years of age. Another 10% develop symptoms between ages two through fifteen, and 35% present with the disease in adulthood (157). The prevalence of disease is unclear and familial occurrence is not common. Both the manifestations of mastocytosis, as well as its prognosis, are dependent upon when the disease presents. Children typically present with cutaneous lesions, the most frequent being a solitary mastocytoma and urticaria pigmentosa. Less commonly they will present with diffuse cutaneous mastocytosis (158) and rarely, telangiectasia macularis eruptiva perstans (TMEP) which is often the presenting symptom in adults (157). The majority of children have cutaneous disease alone and the prognosis for these patients is quite good. Half of these children will experience resolution by adolescence and the remainder will experience a marked reduction in symptoms (159). In a minority of children the disease persists into adulthood, and, like patients with the adult-onset form of the disease, these individuals more commonly experience systemic involvement.

Etiology

The cause of increased mast cell accumulation in the skin and other organs has not been identified. While several hypotheses have been generated none have been confirmed. It has been postulated that the disorder may result from the development of a neoplastic clone. However, clinical observations have not supported this theory. In children with mastocytosis the disease is often self-limited with complete resolution commonly occurring by adolescence. In addition, despite frequent persistence of the adult-onset form throughout life, these patients rarely die from a neoplastic mast cell process. Moreover, the mast cells of these patients usually appear normal by transmission electron microscopy (156).

Critical to the growth and development of mast cells is an array of cytokines that are elaborated by various cell types. Recently, it has been shown that altered metabolism of one of these cytokines, mast cell growth factor (stem cell factor, c-kit ligand), may be the etiology of cutaneous mastocytosis in some patients with this form of the disease (160). In addition to stimulating the growth and differentiation of murine mast cells in vitro (11,18,20-23), mast cell growth factor causes the accumulation of these cells into the skin after dermal injection (17,161). To determine the role of this factor in mastocytosis Longley and colleagues (160), using immunohistochemical techniques and the polymerase chain reaction, looked for the expression of the mast cell growth factor gene in the skin of patients with cutaneous mastocytosis.

As demonstrated previously in Figure 1, mast cell growth factor exists in two forms, a membrane-bound form and a soluble form (18, 23, 162-165). Normally, the soluble form is produced from the membrane-bound form by proteolytic cleavage at a protease-sensitive site (18,23,162-165) that is encoded by a particular region of the gene known as exon 6. This segment of the gene may be present in the final mRNA or it may be deleted in the primary gene transcript due to alternative splicing. Therefore, in order

to produce the soluble form of this factor, both a full-length mRNA must be present as well as a functional protease.

Upon examining normal skin immunohistochemically for the presence of mast cell growth factor, Longley and colleagues (160) found that, while a small number of dermal fibroblasts, dendritic cells and resident mast cells stained positive for the presence of mast cell growth factor, keratinocytes stained to the greatest degree. The pattern of staining was diffuse and granular consistent with the presence of intracellular and membrane-bound forms of the protein. In contrast to the skin of normal subjects, the skin of patients with cutaneous mastocytosis demonstrated abundant immunoreactive mast cell growth factor, not cell-associated, but present in the extracellular spaces, between keratinocytes and free in the papillary dermis. In addition, it was found that there were no sequence abnormalities detected in the mRNA for mast cell growth factor in the one patient studied. Thus, it was concluded that the altered distribution of mast cell growth factor in the skin of patients with cutaneous mastocytosis results from abnormal production of the soluble form of this factor, possibly due to increased proteolytic processing. These findings add support to the hypothesis that mastocytosis results from an abnormal proliferation of mast cells and not from mast cell neoplasia.

Classification schemes

Classification of a disease as heterogeneous as mastocytosis is difficult. In most schemes, cutaneous mastocytosis has been separated from systemic mast cell disease (SMCD), and is described according to the distribution or the morphology of the cutaneous lesions. The characterization of systemic mastocytosis has been based upon the distribution of organ involvement, and upon the nature of the process, i.e., "benign" or "malignant" (166).

Most of the original proposed schemes categorized mastocytosis into "benign" and "malignant" forms and, have not allowed for comparisons to be made between descriptions of mastocytosis in articles from different authors. While "malignant" may be used by one author to describe a patient with systemic mastocytosis that has persisted for many years and that has caused severe gastrointestinal disease, another may use the term to describe a patient who has had the disease for only a few months but who has a rapidly progressive form of the disease. As Metcalfe stated recently in a roundtable discussion on mastocytosis, "Clearly, such definitions do not allow categorization for treatment and for assignment of prognosis." (166)

As knowledge regarding the disease process has improved, it has been realized that mastocytosis may be associated with several clinical conditions that, because of their severity, often predict the disease course. These associated diseases include various myeloproliferative disorders and mast cell leukemia (167). In 1988 a classification scheme incorporating these disease associations was proposed. An extensive study by Travis and colleagues (168) was designed to identify features

important in predicting survival in patients with systemic mastocytosis and findings from it led to the formation of a classification scheme that continues to be useful today. A slightly modified version (166) is presented in Figure 13.

Figure 13

C	CLASSIFICATION OF MASTOCYTOSIS
Indo	olent mastocytosis
Α.	Skin only Urticaria pigmentosa Diffuse cutaneous mastocytosis Mastocytoma Telangiectasia macularis eruptiva perstans
B.	Systemic (<u>+</u> urticaria pigmentosa) Bone Marrow Gastrointestinal
	tocytosis with an associated hematologic disorder irticaria pigmentosa) Dysmyelopoietic disorders Myeloproliferative disorders

III. Mast cell leukemia

C.

D.

E.

IV. Lymphadenopathic mastocytosis with eosinophilia (<u>+</u> urticaria pigmentosa) (aggressive mastocytosis)

Acute non-lymphatic leukemia

Malignant lymphoma

Chronic neutropenia

As shown in Figure 13, the terms "benign" and "malignant" are not included in the revised classification scheme because the distinction between these two forms of the disease may not always be clear-cut. The term "indolent" was adopted to describe patients with favorable prognostic features. While this term suggests that individuals in this category will have a favorable prognosis, it also allows for the possibility that some of the patients in this category may develop severe symptoms or experience a transformation of their disease into a more aggressive form (168).

Patients that, in the past, fell into the "malignant" category have been separated on clinical or pathologic criteria into three groups, all characterized by a rapid disease

course: 1) SMCD with associated hematologic disorders, 2) mast cell leukemia and 3) aggressive SMCD. Travis and colleagues (168) found that, in the SMCD patients with an associated hematologic disorder (those in category II), poor prognosis was more associated with the hematologic disorder than with the SMCD. For that reason, the term "malignant", which suggests that poor prognosis results from the malignant proliferation of mast cells was dropped.

It has been recognized for many years that most patients in the indolent category have urticaria pigmentosa or diffuse cutaneous mastocytosis at some time in their disease. In contrast, the presence of cutaneous disease in patients in categories II through IV is variable. Moreover, it has been noticed if the disease evolves into a leukemic process, urticaria pigmentosa, if present, is sometimes lost (166).

It is important to realize that, despite the prognostic category, the signs and symptoms of mastocytosis are similar and are related to the release of preformed and newly-generated mast cell mediators. Therefore, patients of all categories may experience pruritus, anaphylaxis and eventually, other problems resulting from specific organ involvement. In some instances, however, morbidity and, ultimately, mortality may be dependent upon direct mast cell infiltration as opposed to mediator release.

Since the 1988 classification scheme was proposed by Travis et al. (168), a slightly revised classification was developed in a consensus conference in 1991 and is presented in Figure 14. As shown, the major categories are the same. The only major change is that the potential organs involved in systemic disease in category I are listed (169).

Revised Mastocytosis Classification

- Indolent mastocytosis
 - A. Syncope
 - B. Cutaneous disease
 - C. Ulcer disease
 - D. Malabsorption
 - E. Bone marrow mast cell aggregates
 - F. Skeletal disease
 - G. Hepatosplenomegaly
 - H. Lymphadenopathy
- II. Hematologic disorder
 - A. Myeloproliferative
 - B. Myelodysplastic
- III. Aggressive

Lymphadenopathic mastocytosis with eosinophilia

IV. Mastocytic leukemia

Cutaneous Mastocytosis

Urticaria pigmentosa - In any form of mastocytosis the organ most frequently involved is the skin and, the most frequent cutaneous lesions are those of urticaria pigmentosa. While the lesions usually appear early in life, they may occur in adolescence and in adulthood as well. They appear as discrete reddish-brown macules and papules, and they occur in a randomized and generalized distribution. The presence of Darier's sign (urtication upon stroking due to mast cell degranulation) is a confirmatory sign for the presence of mast cell infiltrates in the skin. Associated symptoms include pruritus and dermographism. However, more severe symptoms may occur if there is extensive cutaneous involvement (170-172). The lesions commonly occur on the trunk, but any area of the skin may be affected, including the mucous membranes. The palms, soles, face, and scalp are usually spared. In infants the lesions are often bullous because tissue edema causes the skin to separate at the poorly formed dermal-epidermal junction (173,174, reviewed in 175).

The number of patients with urticaria pigmentosa that actually have systemic disease remains unclear. Upon evaluating 35 patients with cutaneous lesions alone, Czarnetzki et al. (176) found that 46% had focal mast cell accumulation in the bone marrow. In other studies a range of 10 to 70% has been described (159,177). More recently, it has been suggested that the presence or absence of bone marrow involvement in urticaria pigmentosa may be related to mast cell burden. With few cutaneous lesions there may be no bone marrow involvement, whereas with many lesions bone marrow involvement is more likely (Metcalfe, personal communication).

The prognosis for patients with urticaria pigmentosa depends on the age of onset of the skin lesions. While the majority of children demonstrate marked improvement or total resolution by adolescence or adulthood (159,178), the disease course for adults is less clear. Generally, adults with urticaria pigmentosa have chronic disease and a greater chance of eventual systemic involvement (179).

Mastocytoma - These appear at birth or shortly thereafter (180) as solitary or multiple macules, plaques, or nodules. Most mastocytomas occur on the extremities, but not on the palms or soles. As with urticaria pigmentosa the lesions may urticate with stroking. The prognosis for patients with mastocytomas is very good. In many instances, the lesions either spontaneously involute or improve markedly after several years (159,181).

Diffuse cutaneous mastocytosis - This rare form of cutaneous mastocytosis generally affects children before the age of three. The skin, which is diffusely involved, often takes on a red-yellow-brown color. An erythrodermic form is characterized by diffuse edema and thickening of the skin (173,175). Infants may first present with extensive bullae prior to the development of diffuse cutaneous mastocytosis. Thus, this condition should be included in the differential diagnosis of neonatal blistering diseases (182). While this form of cutaneous mastocytosis usually resolves before age five, these children are at risk of developing flushing, hypotension, shock, and death (175).

Telangiectasia Macularis Eruptiva Perstans (TMEP) - This form of cutaneous mastocytosis occurs primarily in adults. The lesions consist of generalized, red, telangiectatic macules on a tannish-brown background. Generally, they are smaller than the lesions associated with urticaria pigmentosa and are not pruritic (175). The prognosis for patients with TMEP is similar to that of patients with urticaria pigmentosa.

Histopathology and Diagnosis - The diagnosis of cutaneous mastocytosis should be confirmed by biopsy. In urticaria pigmentosa increased mast cell accumulation is seen in the papillary dermis and, in some instances, throughout the entire dermis. In TMEP mast cell accumulation is most prominent around the capillary venules of the superficial plexus. While nonlesional skin may have modestly increased mast cell numbers, lesional skin contains the greatest accumulations. Usually, a fifteen to twenty-fold increase is noted. However, it is important to realize that increased mast cell

numbers alone is not sufficient to make the diagnosis of cutaneous mastocytosis. Numerous other skin diseases have been associated with increased mast cell numbers as well. However, these diseases are easily differentiated from mastocytosis by other distinguishing characteristics (Reviewed in 156).

Mast cells in the skin are identified using metachromatic stains or the conjugated avidin technique (183). Analysis of the granule ultrastructure of the mast cell granules reveal a predominance of grating/lattice structures indicating that these cells are of the MC_{TC} variety (55). More recently, a study using a combination of electron microscopy and morphometric analysis showed that there may be quantitative differences between the mast cells from patients with systemic disease and those from patients with cutaneous involvement alone. Cutaneous mast cells from lesional skin of individuals with systemic disease had a larger mean cytoplasmic area, nuclear size, and granule diameter than those from adults with non-systemic disease (185).

In patients with urticaria pigmentosa and systemic disease mean levels of histamine and tryptase have been shown to be elevated, whereas they have not in patients with cutaneous disease alone (70,186). In light of the difficulty in making accurate histamine determinations and the fact that the majority of histamine is excreted as the metabolite, 1-methyl-4-imidazoleacetic acid (MelMAA), this marker is helpful in identifying patients with mastocytosis. MelMAA levels have been shown to be elevated in the majority of patients with systemic disease as well as in those with cutaneous disease alone (187).

Treatment - An important component of treatment of patients with cutaneous mast cell disease is the avoidance of factors that may trigger mediator release. Potential triggers include: temperature changes, friction, physical exertion, ethanol ingestion, and the use of nonsteroidal anti-inflammatory agents or opiates (175). In addition, it has been demonstrated that patients with urticaria pigmentosa are more predisposed to developing anaphylaxis following Hymenoptera stings despite the absence of antigenspecific IgE (188).

In children with an isolated mastocytoma, surgical removal may be necessary if spontaneous involution does not occur. Patients with urticaria pigmentosa who experience pruritus and whealing benefit from treatment with H1 antihistamines alone or in combination with H2 antihistamines (189). In addition, disodium cromoglycate and ketotifen also have been shown to be effective in some studies (190-194). More recently, a platelet-activating factor antagonist was shown to reduce pruritus and flushing in an adult with urticaria pigmentosa and systemic disease (195).

Another mode of therapy, psoralen plus ultraviolet A photochemotherapy (PUVA), has been shown to decrease both the symptoms associated with urticaria pigmentosa as well as the lesions themselves. However, relapses often occur after the cessation of therapy (196-199). Therefore, because of its possible side effects and transient benefit,

this form of therapy should be used only in refractory cases. Finally, while systemic corticosteroids have not been effective in treating cutaneous mastocytosis, the application of a potent topical corticosteroid under occlusion has had promising results (200).

Figure 15

TREATMENT OF CUTANEOUS MASTOCYTOSIS'

Mastocytoma

Excision if no resolution.

Urticaria pigmentosa and Diffuse/Erythrodermic Cutaneous Mastocytosis

H1 antihistamines
H1 and H2 antihistamines
Ketotifen
Disodium cromoglycate
Topical corticosteroids with occlusion
Ultraviolet B phototherapy
Psoralen plus ultraviolet A photochemotherapy (PUVA)

Telangiectasia macularis eruptiva perstans

Unknown

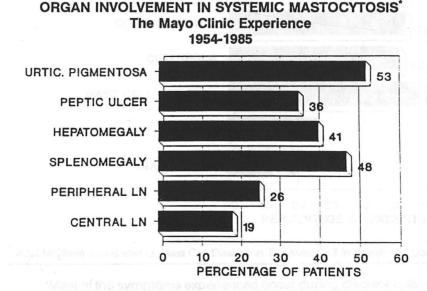
* Adapted from Soter, J Invest Dermatol 96:32S, 1991 (Ref 175).

Systemic Mastocytosis

Systemic mastocytosis is characterized by an abnormal proliferation and accumulation of mast cells in the bone marrow, spleen, liver, skin, bone, and lymph nodes. While children may develop SMCD, the incidence in adults is much greater. Affected patients are usually between the ages of 50 and 80.

The percentage of patients with cutaneous lesions varies from 50% to 100% depending upon the pattern of referral (168). Travis et al. (168), in an extensive analysis of 58 patients with SMCD, found that 53% had cutaneous disease. An even greater percentage (86%) was found by Horan and Austen (201) in a retrospective ten year review at Brigham and Women's Hospital. Involvement of other organs is variable. Travis et al. (168) found hepatomegaly in 41% of the cases examined, splenomegaly in 48%, peptic ulcer in 36% and peripheral and central lymphadenopathy in 26% and 19%, respectively. While the bone marrow was not examined in the Mayo Clinic study, Horan and Austen (201) found bone marrow involvement in 73% of the 21 patients they evaluated.

Figure 16



* Adapted from Travis and Li, Mast Cell Disease in The Mast Cell in Health and Disease, 1993 (Ref 202).

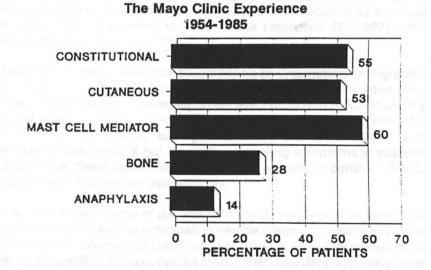
For many patients with SMCD the course is relatively benign and the prognosis is good. However, patients with a more malignant form of the disease, characterized by marked mast cell proliferation and widespread organ involvement often experience a fatal outcome.

Clinical Presentation

Patients with SMCD experience a myriad of signs and symptoms and these were categorized into five major groups in the Mayo Clinic study (168). Constitutional symptoms (fatigue, weight loss, fever and sweats) were experienced by 55% of the patients studied. Cutaneous manifestations (urticaria, pruritus and dermatographism) were experienced by 53%, while symptoms of mast cell mediator release were seen in 60%. The predominant symptoms associated with mediator release in this study included abdominal pain, respiratory symptoms, diarrhea, nausea, vomiting, flushing and syncope and, unless there was underlying asthma, the respiratory symptom experienced by most patients was dyspnea, not wheezing. Bone manifestations consisting of bone pain, arthralgias and fractures were seen in 28% and anaphylaxis occurred in 14%.



SIGNS AND SYMPTOMS OF SYSTEMIC MAST CELL DISEASE'



Adapted from Travis and Li, Mast Cell Disease in The Mast Cell in Health and Disease, 1993 (Ref 202).

Most of the symptoms experienced occur during discrete episodes, and these are often described as "attacks" by the patient. These episodes, which may be brief or prolonged, coincide with the sudden release of mast cell mediators and can be triggered by heat, exertion, emotional upset, or certain pharmacologic agents. The patient initially feels warm and is flushed. Subsequently, he or she experiences palpitations and lightheadedness which are attributed to systemic vasodilation. Dyspnea, nausea, abdominal cramping, vomiting and diarrhea occur variably among patients.

Organ Involvement

Skin - The skin manifestations seen in SMCD are similar to those seen in isolated cutaneous disease. Urticaria pigmentosa, the most common cutaneous manifestation, is seen in over 90% of patients with indolent disease, but in less than 50% of patients with an associated hematologic disorder or aggressive disease (203). Other cutaneous patterns associated with SMCD include telangiectatic, pigmented macules (TMEP); erythroderma; diffuse, infiltrative plaques and nodules, and dermographism and flushing. Each of these are well-described in several extensive reviews (156, 175).

Bone - Approximately 60-70% of patients with SMCD develop radiographic bone lesions (168) with the most common abnormalities being diffuse, sclerotic and lucent areas involving the axial skeleton (179,204,205). Less frequently, focal osteoporotic/osteosclerotic lesions are seen in the skull and long bones (179). Travis et al. (168) found the most common diffuse pattern to be demineralization (28% of patients) followed by osteosclerosis (19%). Mixed sclerosis and demineralization was seen in 10% of the patients studied. The pathophysiology of these skeletal abnormalities is thought to be at least partially dependent upon the mediators produced by the mast cell. Both heparin and PGD₂ are known to cause bone resorption (206,207) while histamine is known to promote fibrosis (208).

While generalized osteopenia was once thought to be an uncommon radiographic finding (209,210), more recent studies have shown that it occurs more often than previously recognized (168,211-215). The osteoporosis associated with SMCD is usually very severe. Vertebral crush fractures are common and are often multiple involving the thoracic and lumbar spine. In addition, back pain resulting from osteoporosis with vertebral fractures may be the major, if not the sole, presenting symptoms in patients with SMCD (212-215). Thus, these studies underscore the necessity to consider SMCD in the differential diagnosis of idiopathic osteoporosis.

Bone marrow - The bone marrow of patients with SMCD frequently has increased mast cell numbers (103,179,201) and its examination reveals diagnostic infiltrates as well as provides useful prognostic information (168,216,217). The finding of increased mast cell numbers alone is not specific for mastocytosis since other disorders, including renal osteodystrophy, multiple myeloma, other malignancies, and common osteoporosis, may demonstrate increases as well. The diagnosis is usually not very difficult, however, since the magnitude of the increase is significantly greater in mastocytosis compared to that seen in the other disorders and the lesions are distinctive. The majority of the lesions are focal and in a paratrabecular location although they may be perivascular and parafollicular. The nodular aggregates consist of spindle-shaped mast cells often associated with lymphocytes and eosinophils. Early in the course of the disease the marrow lesions are cellular, while later in the course when the mast cell number decreases, the lesions become more fibrotic. Several important poor prognostic factors have been identified based upon bone marrow analysis and include: low percentage of

fat cells with a hypercellular marrow; existence of mast cells with lobulated nuclei; and presence of a hematologic disorder (168).

Figure 18

Adult Mast Cell Disease Bone Marrow Involvement

Series	No. of Patients	Focal Lesions	Focal/Diffuse Lesions
Webb	26	84%	100%
Brunning	14	82%	100%
Horny	38	92%	100%
Ridell	18	56%	56%
Travis	58	NA	NA

From Parker, Hematologic Aspects of Mastocytosis. J Invest Dermatol 96(suppl):47S, 1991 (Ref 218).

Critical to the identification of mast cells in the bone marrow is the use of proper staining techniques. Often the metachromatic staining properties of the mast cell granules are lost during routine staining procedures due to the decalcification process. Optimal staining occurs when the biopsy specimen undergoes nondecalcified plastic embedding prior to staining with toluidine blue or other metachromatic dyes. Thus, in light of these requirements, it is important that the pathologist be consulted prior to the biopsy so that the sample is processed properly.

Various hematologic abnormalities have been demonstrated in patients with SMCD (103,168,179). These range from mild cytopenias and cytoses to premalignant and malignant syndromes. Anemia, the most common hematologic abnormality, occurs in a third to a half of patients with systemic disease, while thrombocytopenia and eosinophilia have been demonstrated in up to 25%. Both anemia and thrombocytopenia are significant indicators of poor prognosis. A significant number of patients with hypercellular marrows have other serious associated hematologic disorders including dysmyelopoietic syndromes, myeloproliferative disorders, leukemia, lymphoma and chronic neutropenia (216,219-222). These patients have a markedly reduced 5-year survival (168).

SYSTEMIC MAST CELL DISEASE Hematologic Manifestations*

Cytopenias

Anemia
Thrombocytopenia
Leukopenia/granulocytopenia

Cvtoses

Leukocytosis/granulocytosis Eosinophilia Monocytosis Thrombocytosis Lymphocytosis

*From Parker, Hematologic Aspects of Mastocytosis. J Invest Dermatol 96(suppl):52S, 1991 (Ref 223).

Liver - In the several studies in which it was examined, hepatomegaly occurred in 40%-72% of the patients studied (103,168,179,201,224,225). Despite this frequency of liver involvement, abundant pathological data is lacking. In the majority of patients with mastocytosis the liver, lymph nodes, and spleen are rarely biopsied unless there is significant organ dysfunction or a suggestion that a malignant process is occurring. For these reasons, the available data on mast cell accumulations in these organs is based primarily on data from patients with aggressive disease. A study of hepatic pathology in thirteen patients revealed hepatic fibrosis in all specimens examined (226). Fibrotic patterns included a periductal pattern and portal to portal fibrosis with fatty metamorphosis and sinusoidal dilatation also occurring in a significant number. A mononuclear cellular infiltrate was seen but inflammation was not prominent. Interestingly, excluding the alkaline phosphatase, the liver function studies were often normal despite extensive liver involvement.

Spleen - Splenomegaly is not an uncommon finding in SMCD. Travis et al. (168) found that 48% of the 58 patients they evaluated had an enlarged spleen while Horny et al. (227) demonstrated involvement in 72% of their patients. These percentages are similar to those that were obtained in three previous studies (103,179,224). Pathological

examination revealed mast cell infiltrates in a paratrabecular distribution with various degrees of trabecular fibrosis and eosinophilic infiltrates (228).

Lymph nodes - Lymph node involvement is less common than either hepatomegaly or splenomegaly. In the study by Travis et al. (168) peripheral lymphadenopathy was noted in 26% of 58 patients examined while central lymphadenopathy occurred in 19%. Pathologic evaluation revealed that mast cell infiltrates were most commonly found in the paracortex, followed by the follicles, medullary cords and the sinuses. These were accompanied by eosinophils. Lymph node involvement has been found to be more extensive in patients with either associated hematologic malignancies or aggressive non-leukemic mastocytosis (228,229).

Gastrointestinal tract - Gastrointestinal symptoms are common in patients with SMCD. Abdominal pain was seen in 35% of the patients evaluated by Travis et al. (168) and in 80% of the 21 patients retrospectively reviewed by Horan and Austen (201). Gl symptoms were also seen in 80% of the 16 patients recently evaluated by Cherner et al. (230). Two different types of abdominal pain, dyspeptic and nondyspeptic, are generally described (201,230). The dyspeptic variety is often associated with increased acid secretion (secondary to stimulation by histamine) and peptic ulcer disease (168,230), while the nondyspeptic variety is not. Cherner et al. (230) found that 38% of their patients had gastric acid hypersecretion and 44% had evidence of peptic ulcer disease. Interestingly, increased acid hypersecretion and increased incidence of peptic ulcer disease had not been demonstrated in previous studies (231-233). The low incidence of these two findings in previous studies probably resulted from their underdiagnosis, not their absence. Cherner (230) also found that a significant number of patients (30%) experienced abdominal pain of the nondyspeptic variety. This pain was localized to the lower abdomen, was not relieved by H2- receptor antagonists, and was not associated with abnormal radiographic findings or gastric acid hypersecretion.

Multiple factors may contribute to the GI abnormalities seen in systemic mastocytosis. The cause of abdominal pain may result from peptic ulcer disease (171,172,230), edema/urticaria of the GI tract or a motility disorder (171,172,231). Diarrhea, another frequent GI manifestation, appears to result from altered intestinal secretion, structural disease of the small intestine or a hypermotility/transit disorder (203). Malabsorption, caused by diffuse small intestinal mucosal dysfunction (230,232,233), may also be seen but is usually not severe.

Summary of Clinical Features

The spectrum of signs and symptoms at presentation may lead to an incorrect initial diagnosis. In fact, of the 58 patients evaluated by Travis et al (168), 80% were initially incorrectly diagnosed. Since systemic mastocytosis has a predilection for certain organs, it is important that this disease process be considered in patients presenting with unexplained bone, skin, hematologic, gastrointestinal and systemic manifestations.

INITIAL DIAGNOSIS IN SMCD PATIENTS AT MAYO CLINIC

Diagnosis	Number of Patients
Bone manifestations	
Osteoporosis	
Metastatic carcinoma	5
Multiple myeloma	4
Paget disease	2
Gaucher disease	1
Skin manifestations	•
Malignant melanoma	1
Rocky Mountain spotted fever	
"Colored hives"	9 1
Hematologic manifestations	
Lymphoma	
Acute leukemia	6
Chronic myeloid leukemia	3
Myelofibrosis	3
Myeloproliferative disorder	2
Neutropenia	2
Hypereosinophilic syndrome	1
Eosinophilic leukemoid reaction	1
ITP	1
	1
Idiopathic splenomegaly	1
Gastrointestinal manifestations	
Irritable bowel	
Primary biliary cirrhosis	1
Portal hepatitis	1
Systemic manifestations	
Diabetic shock	
Carcinoid syndrome	1
Hypertension	1
Hypotension	1
Hypersensitivity vasculitis	1
Parasitic infection	1
rarasilic injection	1

Diagnostic Evaluation

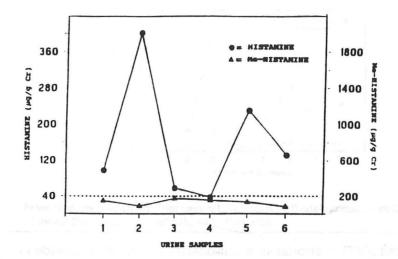
The diagnosis of systemic mastocytosis is based upon multiple criteria. Clinical presentation, physical exam, laboratory analysis, radiographic findings and histologic findings all must be considered. Since the majority of patients present with skin manifestions the diagnosis is easily confirmed by skin biopsy. Again, it must be emphasized, that the specimen must be prepared and stained properly so that the mast cells can be identified. In addition, since small increases of cutaneous mast cells can be seen in other disorders, increased numbers alone is not sufficient. At least a ten-fold increase is usually seen (234, Metcalfe, personal communication). In the absence of cutaneous lesions, however, the diagnosis is more problematic. While a blind skin biopsy may be helpful by revealing increased mast cell numbers, usually the diagnosis requires a more extensive evaluation.

Numerous biochemical mediators are released upon mast cell activation, and their identification and quantitation can help to establish the diagnosis of SMCD. Again, the finding of increased mediator levels alone is not sufficient to establish a diagnosis since individuals with episodic mast cell activation, but no evidence of abnormal mast cell proliferation (i.e. those with anaphylaxis), may have increased mediator levels as well. However, one differentiating factor between these two groups is that mediator levels in patients with mastocytosis usually are chronically elevated while, in patients with anaphylaxis, they are elevated immediately after mast cell activation only. The mast cell secretory products most easily identified clinically are histamine and its metabolites, PGD₂, tryptase, and heparin (Reviewed in 235).

While numerous methods exist for measuring histamine in both serum and urine (236-239,reviewed in 235), many lack sensitivity, specificity and reproducibility (240-242). In addition to these difficulties, there are other significant problems associated with quantifying histamine in human biological fluids. Unless blood is collected carefully, artificial elevations of histamine may be found secondary to basophil histamine release. This problem may be reduced by using a calcium chelator such as EDTA as an anticoagulant to reduced basophil activation, but it often is not eliminated (235).

Other difficulties also exist when attempting to quantitate urinary histamine. Certain bacteria can decarboxylate histidine to histamine and, if these are present in sufficient quantities in the urinary tract, urinary histamine levels may be falsely elevated (187). By measuring histamine metabolites, N-methyhistamine and N-methylimidazole acetic acid (MelMAA), this problem can be circumvented. While bacterial metabolism to histamine can occur, further metabolism does not. Therefore, while a normal individual may have elevated urinary histamine levels, the levels of histamine metabolites will be normal. As would be expected, this problem occurs more often in women since they are more predisposed to developing an overproliferation of urinary tract bacteria.

24-HOUR URINE COLLECTION FROM A NORMAL FEMALE Histamine and N-methylhistamine levels

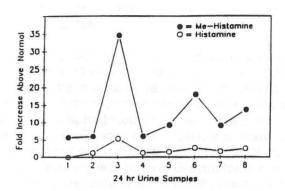


From Roberts and Oates, Biochemical Diagnosis of Systemic mast Cell Disorders. J Invest Dermatol 96(suppl)19S, 1991 (Ref 235).

Urine histamine determinations also may not be an accurate diagnostic indicator of systemic mast cell activation in patients with SMCD due to altered histamine metabolism (187,243). In some patients, while elevated levels of histamine metabolites are present in the urine, the histamine level itself may be normal. Therefore, for all of the above reasons, urinary analysis of histamine metabolites, not histamine, should be performed on every patient suspected of have systemic mast cell disease.

Granerus et al. (243) in 1983 determined whether extent of disease involvement correlated with the amount of urinary histamine metabolites in 30 patients with mastocytosis. He found that patients with skin manifestations alone had low levels of urinary MelMAA levels while those with extensive organ involvement had markedly elevated levels. Thus, urinary histamine metabolite analysis may also be useful in distinguishing between isolated cutaneous disease from more widespread systemic involvement as well as in the initial diagnosis of the disease itself.

24-HOUR URINE COLLECTION FROM A PATIENT WITH SMCD Histamine and N-methylhistamine levels*



From Roberts and Oates, Biochemical Diagnosis of Systemic mast Cell Disorders. J Invest Dermatol 96(suppl)19S, 1991 (Ref 235).

In addition to histamine metabolites, a metabolite of PGD_2 has also been found to be elevated in the urine of patients with SMCD. Using a mass spectrometric assay for the measurement of 9α -hydroxy-11, 15-dioxo-2,3,18,19, -tetranorprost-5-ene-1, 20-dioic acid Roberts (244) detected elevations of this metabolite in patients with proliferative mast cell disease. More recently, an assay for another PGD_2 metabolite has been developed and levels of this metabolite, as well, were found to be elevated and were shown to correlate with levels of N-methylhistamine in patients with SMCD (245). Unfortunately, neither of these assays are available for routine clinical use.

While heparin is a preformed mediator and is released during mast cell activation and secretion, adverse effects from this release are rarely noted. The partial thromboplastin time has been found to be elevated only during extensive mast cell activation (246,247). Therefore, for this reason, routine coagulation studies have not been recommended in the diagnostic evaluation of patients with suspected mastocytosis.

Probably the most useful indicator of mast cell activation is the presence of elevated plasma tryptase levels. As shown in Figure 6, elevations have been demonstrated in the majority of patients with mastocytosis and anaphylaxis, but not those with sepsis or myocardial infarctions (170). Interestingly, of the 17 mastocytosis patients evaluated six did not have elevated tryptase values. All of these had urticaria pigmentosa and mast cell involvement of one or more organs. However, because all were evaluated during a quiescent period (not during an "attack") mediators may not have been elevated due to a smaller mast cell burden in these six individuals.

Tryptase is measured by radioimmunoassay and serum is required. An important advantage of measuring serum tryptase is that determinations do not need to be made immediately after mast cell activation. Levels are maximal one to two hours following activation, and they usually remain elevated for several hours (248). Initially, this assay was not commercially available but now several laboratories provide this service. Tryptase determinations are available at Parkland Memorial Hospital, Zale-Lipshy University Hospital and the Aston Center.

While the finding of elevated mast cell mediators is suggestive of the diagnosis it is not diagnostic. In the absence of skin lesions a bone marrow biopsy should be performed in order to establish the diagnosis as well as to determine the disease category. In patients with cutaneous manifestations and peripheral blood abnormalities, hepatomegaly, splenomegaly, or lymphadenopathy a bone marrow biopsy should be performed as well to determine the existence of an associated hematologic disorder. Additional evaluation, including bone scan, skeletal surveys, and GI studies should be performed in those individuals presenting with bone or GI symptoms, respectively. Since decreased attention span, memory impairment and irritability have been noted in some individuals with mastocytosis (203) a neuropsychiatric evaluation may be warranted. In 1991 participants of a roundtable discussion on mastocytosis developed a consensus diagnostic workup which is presented in Figure 23.

Figure 23

CONSENSUS DIAGNOSTIC WORK-UP*

If mastocytosis is suspected on clinical grounds:

Routine

Examine skin - gross/microscopic
Bone marrow biopsy and aspiration
24-hour urine for mediators

Additional studies

Bone scan/skeletal survey Gl work-up EEG, neuropsychiatric work-up

From Metcalfe, Conclusions. J Invest Dermatol 96(suppl):64S, 1991 (Ref 169).

Treatment

The chronic treatment of mastocytosis should be aimed at blocking the effects of the released mast cell mediators as well as stabilizing the mast cell itself. Mediator release leads to the prominent manifestations of anaphylaxis, gastric hypersecretion, Gl cramping and pruritus (203).

 $\rm H_1$ -and $\rm H_2$ -receptor antagonists - $\rm H_1$ -receptor antagonists, such as hydroxyzine and the tricyclic antidepressant, doxepin, remain mainstays of treatment and have been shown to reduce pruritus, flushing and tachycardia (203). While the new long-acting, nonsedating $\rm H_1$ -antihistamines may prove to be beneficial, to date, no controlled studies have been performed. Patients with gastritis and peptic ulcer disease have been shown to be controlled with $\rm H_2$ -antihistamines. However, in patients with diarrhea $\rm H_2$ -antihistamines are usually not effective (249) and, in these individuals anticholinergics may be beneficial. Often, both $\rm H_1$ - and $\rm H_2$ -receptor antagonists are required to completely block the effects of histamine (250,251). However, despite this combination some patients continue to complain of flushing and headaches. In these individuals the levels of histamine may be very high or, more likely, the symptoms are secondary to other mast cell mediators that are unaffected by antihistamines.

Aspirin - In many patients PGD₂, as well as histamine, is elevated and, as expected, the episodes of vasodilation in these individuals are not controlled with antihistamines alone (252). While aspirin has been shown to be effective in these patients, high salicylate levels are necessary to achieve control (253). In light of the potential mast cell activating effects of aspirin, initial treatment should consist of combined antihistamine therapy. Subsequently, if no benefit is achieved, aspirin may be judiciously added and increased gradually until plasma salicylate levels rise to 20-30 mg/dl (254). If a patient does not respond to aspirin therapy he or she can not be considered a treatment failure unless effective cyclooxygenase inhibition has been ensured either by documentation of high salicylate levels or decreased production of urinary PGD₂ metabolites. Other nonsteroidal anti-inflammatory drugs have not been used extensively since blood levels of these agents can not be monitored. Unfortunately, many mastocytosis patients have GI symptomatology and often, these individuals are unable to tolerate aspirin.

Mast cell stabilizing agents - Cromolyn sodium, a stabilizer of mast cell membranes, has been found to be somewhat efficacious in mastocytosis patients, especially those with GI symptoms (189,190,253). While only 1% of orally administered cromolyn sodium is absorbed from the GI tract, it has been shown to alleviate both diarrhea and abdominal pain in patients with these symptoms (190). In addition, pruritus, whealing, flushing, and cognitive function have been shown to improve. Interestingly, despite clinical improvement, mastocytosis patients do not demonstrate decreased urine and plasma histamine levels while on treatment with this agent (189). Another treatment choice is ketotifen, an H₁-receptor antagonist with mast cell stabilizing

properties. While demonstrating effectiveness in relieving pruritus and whealing associated with mastocytosis, a double-blind, placebo-controlled crossover study of eight children revealed this drug to be no more effective than hydroxyzine in relieving the symptoms of mastocytosis (256).

Psoralens with long-wave ultraviolet irradiation (PUVA) - Mastocytosis patients with urticaria pigmentosa have been shown to have decreased whealing and pruritus after one to two months of treatment with PUVA (196-199). However, unfortunately, within months of ceasing therapy symptoms often recur. In light of the potential side effects of long-term PUVA therapy, this treatment is recommended only for those individuals with cutaneous disease unresponsive to other forms of therapy.

Steroids - In cases of severe malabsorption, which results from mast cell infiltration of the lamina propria of the small intestine, oral corticosteroids have led to marked improvement (230). Steroids have also been shown to be effective in reducing the ascites that may occur in the presence of liver fibrosis (257). Unfortunately, steroids may exacerbate the bone disease often present in patients with SMCD and, for that reason, they should be reserved for patients with severe malabsorption. More recently, tixocortol pivalate, a steroid without significant adrenal-pituitary axis suppression, was shown to alleviate both diarrhea and abdominal pain four patients (258).

Chemotherapy - Chemotherapy is not indicated in the treatment of type I mastocytosis. However, for patients with type II disease, therapy is dictated by the associated hematologic disorder. Chemotherapy has not been shown to be effective in the treatment of mast cell leukemia.

Splenectomy - Recently, the records of 26 patients with mastocytosis were reviewed to determine the role of splenectomy in the management of SMCD (259). Seventeen (65%) patients with indolent disease (type I mastocytosis) did not undergo splenectomy; symptomatic treatment only was required. Nine (35%) had type II or type IV disease, and five of these underwent splenectomy. The patients who did not undergo splenectomy died from bleeding complications caused by severe thrombocytopenia. Length of survival was in these patients was 26 months, whereas it was 34 months at the time of the report for those who had undergone splenectomy. These data suggest that, while splenectomy is of no value in patients with indolent mastocytosis, it may be beneficial in those with aggressive disease.

Additional considerations - If mastocytosis patients are to undergo surgery several precautions should be taken. Muscle relaxants that are known to degranulate mast cells should be avoided as should succinyl choline and opiates. In addition, it has been suggested that these patients be premedicated with H₁- and H₂-antihistamines prior to the procedure (253). Radiocontrast dye, too, must be administered with caution. Patients should be premedicated with antihistamines and steroids and the less, hyperosmolar agents should be used. Patients who experience recurrent episodes of

anaphylaxis should be treated with H_1 - and H_2 -antihistamines chronically and they should carry a premeasured epinephrine preparation at all times. Rarely, anaphylaxis following a Hymenoptera sting has been seen as the presenting symptom of systemic mastocytosis and interestingly, many of the patients that have been studied did not have antigen-specific IgE (188, 260). These patients too should be encouraged to carry epinephrine at all times.

Figure 24

Antihistamines: H-1 receptor blockade H-2 receptor blockade Epinephrine Steroids Cromolyn sodium Aspirin Anticholinergics PUVA Chemotherapy

From Metcalfe, The Treatment of Mastocytosis. J Invest Dermatol 96(suppl):55S, 1991 (Ref 253).

Splenectomy

Experimental Interventions

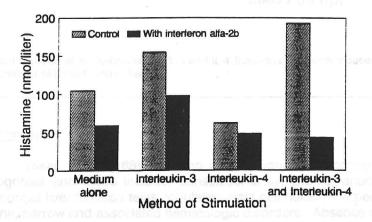
While antiproliferative drugs and radiotherapy have not been effective in the treatment of mastocytosis (168) alternative agents may be beneficial in the treatment of certain types of mastocytosis. Kluin-Nelemans et al. (261) demonstrated improvement in a male patient with aggressive mastocytosis and an associated myeloproliferative disorder (category II) after treatment with interferon alfa-2b. Prior to therapy, the patient's bone marrow contained a large percentage of atypical mast cells, fibrosis, and almost no residual hematopoiesis. After seven months of therapy the percentage of mast cells had decreased dramatically and hematopoietic cellularity significantly improved as well. Clinical improvement was noted after two months of therapy. Red cell transfusion

requirements were reduced in frequency, as were attacks of flushing, generalized itching, abdominal pain, vomiting, diarrhea, hypotension and syncope. After 12 months the patient's urticarial pigmentosa lesions had almost resolved; the liver was no longer palpable and his ascites was gone.

Over the course of therapy the urinary excretion of N-methylhistamine and N-methylimidazoleacetic acid rapidly decreased. In order to determine if the reduction in the urinary excretion of histamine metabolites resulted from decreased release of histamine from mast cells, *in vitro* experiments were performed using bone marrow mast cells obtained prior to therapy with interferon alfa-2b. As shown in Figure 26, bone marrow mast cells spontaneously released histamine when cultured in media alone. This secretion was enhanced by interleukin-3 alone and by interleukin-3 and interleukin-4 together, but not by interleukin-4 alone. When interferon alfa-2b was added to the cultures, both the spontaneous and the induced release of histamine was reduced markedly. Cell viability was not affected. These results, taken together, suggest that interferon alfa-2b has both an antisecretory and an antiproliferative effect on malignant mast cells and that it may be effective in some forms of aggressive mastocytosis.

Figure 25

EFFECT OF ALFA-2B ON IN VITRO SECRETION OF HISTAMINE BY BONE MARROW MAST CELLS*



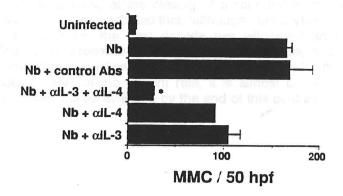
From Kluin-Nelemans et al., Response to Interferon Alfa-2b in a Patient with Systemic Mastocytosis. N Engl J Med 326:619, 1992 (Ref 261).

Experimental results in rodents suggest that cytokine manipulation may be useful in treating mastocytosis. Infection in rodents with the nematode parasite *Nippostrongylus brasiliensis* (Nb) leads to intestinal mastocytosis, eosinophilia, and

elevated serum IgE levels. Madden et al. (262) demonstrated that by injecting young Balb/c mice with monoclonal antibodies to IL-3 and IL-4 prior to Nb infection, intestinal mastocytosis does not develop. These results demonstrate that IL-3 and IL-4 are important *in vivo* in the generation of intestinal mastocytosis and that therapeutic interventions designed to inhibit these cytokines may lead to suppression of disease.

Figure 26

SUPPRESSION OF INTESTINAL MASTOCYTOSIS BY ANTI-IL-3 AND ANTI-IL-4



From Madden et al., Antibodies to IL-3 and IL-4 Suppress Helminth-Induced Intestinal Mastocytosis. J Immunol 147:1387, 1991 (Ref 262).

Prognosis

Travis et al. (168) identified several variables strongly associated with poor prognosis and these include: constitutional symptoms, anemia, thrombocytopenia, abnormal liver function tests, lobulated mast cell nuclei, low percentage fat cells in the bone marrow and associated hematologic disorders. Absence of urticaria pigmentosa, male sex, hepatomegaly, splenomegaly, normal bone radiographic findings and absence of skin and bone symptoms also are associated with poor prognosis. In addition to these variables, prognosis is dependent upon the category of disease. Patients in category I who have cutaneous involvement only have the best prognosis. While approximately half of children with isolated urticaria pigmentosa are lesion-free by adulthood, adults who present with urticaria pigmentosa often develop systemic disease. The course for patients with an associated hematologic disorder is dependent upon the

adulthood, adults who present with urticaria pigmentosa often develop systemic disease. The course for patients with an associated hematologic disorder is dependent upon the prognosis of the particular hematologic disorder itself. Survival for patients with mast cell leukemia is usually less than six months, while individuals with lymphadenopathic mastocytosis with eosinophilia survive two to four years with aggressive symptomatic management (203).

Conclusion

Mastocytosis is, as was stated by Tharp (156) ten years ago, a "spectrum of disorders" in which aberrant mast cell proliferation is heterogeneous in its predilection for particular organs. More recently, at the closing of a roundtable discussion on mastocytosis in 1991, Metcalfe (169) concluded that, "although mastocytosis remains in some ways an enigmatic disorder, the past decade has witnessed an increased awareness of its significance, the processes involved in its pathogenesis, and advances in its therapy." If our knowledge of mast cells and the cytokines involved in their growth and regulation continues to soar at the current rate, it is almost certain that more effective therapeutic modalities will be available by the end of this century.

REFERENCES

- 1. Nettleship E and Tay W: Rare forms of urticaria. Br Med J 2:323, 1869.
- 2. Ehrlich P: Bertrage zur Kenntnis der Anilinfarbungen und ihrer Verwendurg in der Mikroskopischen Technik. Arch Mikros Anat 13:263, 1877.
- 3. Sangster A: An anomalous mottled rash, accompanied by pruritus, factitious urticaria and pigmentation urticaria pigmentosa. Trans Clin Svc, London 11:161, 1878.
- 4. Unna PG, Beitrage W: Anatomie und Pathogenese dur urticaria simples und pigmentosa. Monatsschr Prakt Dermat 3H:1, 1887.
- 5. Sezary A, Levy-Coblentz G, Chauvillon P: Dermographisme et mastocytose. Bull Soc Fr Dermatol Syphiligr 43:359, 1936.
- 6. Ellis JM: Urticaria pigmentosa: a report of a case with autopsy. Arch Pathol 48:426, 1949.
- 7. Michels NA: The mast cells. Reprinted in Ann N Y Acad Sci 103:232, 1963.
- 8. Lloyd AG, Bloom GD, Balazs EA, et al: Combined biochemical and morphological ultrastructure studies on mast cell granules. Biochem J 103:75, 1967.
- 9. Burnet FM: The probable relationship of some or all mast cells to the T-cell system. Cell Immunol 30:358, 1977.
- 10. Burnet FM: Possible identification of mast cells as specialized postmitotic cells. Med Hypotheses 1:3, 1975.
- 11. Zucker-Franklin D, Grusky G, Hirayama N, Schnipper E: The presence of mast cell precursors in rat peripheral blood. Blood 58:544, 1981.
- 12. Kitamura Y, Go S, Hatanaka S: Decrease of mast cells in W/W mice and their increase by bone marrow transplantation. Blood 52:447, 1978.
- 13. Kitamura Y, Go S: Decreased production of mast cells in SI/SI^d mice. Blood 53:492, 1979.
- 14. Geissler EN, Ryan MA, Housman DE: The dominant-white spotting (W) locus of the mouse encodes the c-kit proto-oncogene. Cell 55:549, 1988.
- 15. Chabot B, Stephenson DA, Chapman VM, et al: The proto-oncogene c-kit

- Yarden Y, Kuang WJ, Yang Feng T, et al: Human proto-oncogene c-kit: a new cell surface receptor tyrosine kinase for an unidentified ligand. EMBO J 6:3341, 1987.
- 17. Zsebo KM, Williams DA, Geissler EN, et al: Stem cell factor is encoded at the SI locus of the mouse and is the ligand for the c-kit tyrosine receptor. Cell 63:213, 1990.
- 18. Anderson DM, Lyman SD, Baird A, et al: Molecular cloning of mast cell growth factor, a hematopoietin that is active in both membrane bound and soluble forms. Cell 63:235, 1990.
- 19. Flanagan JG, Leder P: The kit ligand: a cell surface molecule altered in steel mutant fibroblasts. Cell 63:185, 1990.
- 20. Huang E, Nocka E, Beir DR, et al: The hematopoietic growth factor KL is encoded by the sl locus and is the ligand of the c-kit receptor, the gene product of the W locus. Cell 63: 225, 1990.
- 21. Williams DE, Eisenman J, Baird A, et al: Identification of a ligand for the c-kit proto-oncogene. Cell 63:167, 1990.
- 22. Copeland NG, Gilbert DJ, Cho BC, et al: Mast cell growth factor maps near the steel locus on mouse chromosome 10 and is deleted in a number of steel alleles. Cell 63: 175, 1990.
- 23. Zsebo KM, Wypych J, McNiece IK, et al: Identification, purification, and biological characterization of hematopoietic stem cell factor from buffalo rat liver-conditioned medium. Cell 63:195, 1990.
- 24. Martin FH, Suggs SV, Langley KE et al: Primary structure and functional expression of rat and human stem cell factor DNAs. Cell 63:203, 1990.
- 25. Nocka K, Buck J, Levi E, Besmer P: Candidate ligand for the c-kit transmembrane kinase receptor: KL, a fibroblast derived growth factor stimulates mast cells and erythroid progenitors. EMBO J 10:3487, 1990.
- Witte UN: Steel locus defines new multipotent growth factor. Cell 63:5, 1990.
- 27. Galli SJ, Geissler EN, Wershil BK, Gordon JR, Tsai M, Hammel I: Insights into mast cell development and function derived from analyses of mice carrying mutations at Beige, W/c-kit, or SI/SCF (c-kit Ligand) loci. In *The Mast Cell in Health and Disease*. Edited by MA Kaliner and DD Metcalfe. New York, Marcel Dekker, Inc, pp 129-202, 1993.

- 28. Kirshenbaum AS, Goff JF, Dreskin SA, Irani A-M, Schwartz LB, Metcalfe DD: Interleukin 3-dependent growth of basophil-like and mast-like cells from human bone marrow. J Immunol 142:2424, 1989.
- 29. Kirshenbaum AS, Goff JP, Metcalfe DD: Human macrophages cultured on agar but not agarose resemble mast cells. Immunology 68:120, 1989.
- 30. Kirshenbaum AS, Dreskin SC, Metcalfe DD: A staphylococcal protein A rosetting assay for the demonstration of high affinity IgE receptors on rIL-3-dependent human basophil-like cells grown in mixed cultures. J Immunol 123:55, 1989.
- 31. Rimmer EF, Horton MA: Origin of human mast cell studied by dual immunofluorescence. Clin Exp Immunol 68:712, 1987.
- 32. Valent P, Ashman LK, Hinterberger W, et al: Mast cell typing: Demonstration of a distinct hematopoietic cell type and evidence of immunophenotypic relationship to mononuclear phagocytes. Blood 73:1778, 1989.
- 33. Kirshenbaum AS, Kessler SW, Goff JP, Metcalfe DD: Demonstration of the origin of human mast cells from CD34+ bone marrow progenitor cells J Immunol 146:1410, 1991.
- 34. Levi-Schaffer F, Austen KF, Gravallese PM, et al: Coculture of interleukin 3dependent mouse mast cells with fibroblasts results in a phenotypic change of the mast cells. Proc Natl Acad Sci USA 83:6485, 1986.
- 35. Ginsburg H, Ben-Shazar D, Ben-David E: Mast cell growth on fibroblast monolayers: two-cell entities. Immunology 45:371, 1982.
- 36. Fujita J, Nakayama H, Onoue H, et al: Fibroblast-dependent growth of mouse mast cells in vitro: duplication of mast cell depletion in mutant mice of W/W genotype. J Cell Physiol 134:78, 1988.
- 37. Sonoda T, Kanayama Y, Hara H, et al: Proliferation of peritoneal mast cells in the skin of W/W mice that genetically lack mast cells J Exp Med 160:138, 1984.
- 38. Nakano T, Sonoda T, Hayashi C, et al: Fate of bone marrow-derived cultured mast cells after intracutaneous, intraperitoneal and intravenous transfer into genetically mast cell deficient W/W mice. Evidence that cultured mast cells can give rise to both connective tissue type and mucosal mast cells. J Exp Med 162:1025, 1985.

- 39. Kobayashi T, Nakana T, et al: Formation of mast cell colonies in methylcellulose by mouse peritoneal cells and differentiation of these cloned cells in the skin and the gastric mucosa of W/W mice: evidence that a common precursor can give rise to both connective tissue and mucosal mast cells. J Immunol 136:1378, 1986.
- 40. Schwartz LB, Huff T: Biology of mast cells and basophils. In *Allergy: Principles and Practice*. Edited by E Middleton, Jr, CE Reed, EF Ellis, NF Adkinson, Jr, JW Yunginger, WW Busse. St Louis, Mosby, pp 135-168, 1993.
- 41. Galli SJ, Austen KF: *Mast Cell and Basophil Differentiation in Health and Disease*. New York, Raven Press, 1989.
- 42. Irani, AA, Schechter, NM, Craig SS, DeBlois G, Schwartz LB: Two types of human mast cells that have distinct neutral protease compositions. Proc Natl Acad Sci USA 83:4464, 1986.
- 43. Schechter NM, Irani A-M, Sprows JL et al: Identification of a cathepsin G-like proteinase in the MC_{TC} type of human mast cell. J Immunol 145:2652, 1990.
- 44. Irani AM, Craig SS, DeBlois G, et al: Deficiency of the tryptase-positive, chymase-negative mast cell type in gastrointestinal mucosa of patients with defective T lymphocyte function. J Immunol 138:4381, 1987.
- 45. Bauza MT, Lagunoff D: Histidine transport by isolated rat peritoneal mast cells. Biochem Pharmacol 30:1271, 1981.
- Schayer RW: Histidine decarboxylase in mast cells. Ann N Y Acad Sci 103:164, 1963.
- 47. Theoharides TC: Histamine₂-receptor antagonists in the treatment of urticaria. Drugs 37:345, 1989.
- 48. Ring J, Sedlmeier F, von der Helm D, et al: Histamine and allergic diseases. In *New Trends in Allergy II*. Edited by J Ring, G Burg. Berlin, Springer-Verlag, pp 44-77, 1986.
- 49. Joad JP, Casale TB: Airway histamine H₁-receptors. In *Current Topics in Pulmonary Pharmacology and Toxicology*. Edited by MA Hollinger. New York, Elsevier, pp 95-121, 1987.
- 50. Tung R, Kagey-Sobotka, A, Plaut M, Lichtenstein LM: H₂-antihistamines augment antigen-induced histamine release from human basophils in vitro. J Immunol 129:2113, 1982.

- 51. Rocklin RE: Modulation of cellular-immune responses in vivo and in vitro by histamine receptor-bearing lymphocytes. J Clin Inves 57:1051, 1976.
- 52. Schwartz A, Askenase PW, Gershon RK: Histamine inhibition of the in vitro induction of cytotoxic T-cell responses. Immunopharmacology 2:179, 1980.
- 53. Casale, TB: Histamine H₁ and H₂ receptors. In H₁ and H₂ Histamine Receptors. Edited by G Settipane. Providence, RI, Ocean Side Publications, pp 14-20, 1988.
- 54. Fallah HA, Maillard JL, Voison GA: Regulatory mast cells I. Suppressive action of their products on an in vitro primary immune reaction Ann Immunol 126:669, 1975.
- 55. Lima M, Rocklin RE. Histamine modulates in vitro IgG production by pokeweed mitogen-stimulated human mononuclear cells. Cell Immunol 64:324, 1981.
- 56. Kaliner MA, Check WA: Non-sedating antihistamines. Allergy Proc 9:649, 1988.
- 57. Steinbusch HWM, Mulder AH: Localization and projections of histamine immunoreactive neurons in the central nervous system of the rat. In *Frontiers in Histamine Research*. Edited by CR Ganellin, JC Schwartz. Oxford, Pergamon Press, pp 119-130, 1985.
- 58. Wantanabe T, Taguchi Y, Takeda N, et al: Purification of and antibodies against 1-histidine decarboxylase. In *Frontiers in Histamine Research*. Edited by CR Ganellin, JC Schwartz. Oxford, Pergamon Press, pp 91-102, 1985.
- 59. Haaksma EEJ, Leurs R, Timmerman H: Histamine receptors: Subclasses and specific ligands. Pharmacol Ther 47: 73, 1990.
- 60. Arrang JM, Gargbarg M, Schwartz JC: Autoinhibition of brain histamine release mediated by a novel class (H₃) of histamine receptor. Nature 302:832, 1983.
- on der Werf JF, Timmerman H: The histamine H₃ receptor: A general presynaptic histaminergic regulatory system? Trends Pharmacol Sci 10:159, 1989.
- 62. Schwartz JC, Arrang JM, Garbarg M, Pollard H: A third histamine receptor subtype: Characterization, localization and functions of the H₃ receptor. Agents Actions 30:13, 1990.
- 63. Dale HH, Laidlaw PP: The physiological action of betaimidazolylethylamine. J Physiol 41:318, 1910.

- 64. Dale HH, Laidlaw PP: Histamine shock. J Physiol 52:355, 1919.
- 65. Schwartz LB, Lewis RA, Austen KF: Tryptase from human pulmonary mast cells. Purification and characterization. J Biol Chem 256:11939, 1981.
- 66. Hopsu VK, Glenner GG: A histochemical enzyme kinetic system applied to the trypsin-like amidase and esterase activity in human mast cells. J Cell Biol 17:503, 1963.
- 67. Alter SC, Metcalfe DD, Bradford TR, et al: Regulation of human mast cell tryptase. Effects of enzyme concentration, ionic strength and the structure and negative charge density of polysaccharides. Biochem J 248:821, 1987.
- 68. Gruber BL, Marchese MJ, Suzuki K, et al: Synovial procollagenase activation by human mast cell tryptase dependence upon matrix metalloproteinase 3 activation J Clin Invest 84:1657, 1989.
- 69. Schwartz LB, Lewis RA, Seldin D, et al: Acid hydrolases and tryptase from secretory granules of dispersed human lung mast cells. J Immunol 126:1290, 1981.
- 70. Schwartz LB, Metcalfe DD, Miller JS, et al: Tryptase levels as an indicator of mast cell activation in systemic anaphylaxis and mastocytosis. N Engl J Med 316:1622, 1987.
- 71. Wenzel SE, Fowler AA III, Schwartz LB: Activation of pulmonary mast by bronchoalveolar allergen challenge. In vivo release of histamine and tryptase in atopic subjects with and without asthma. Am Rev Respir Dis 184:1002, 1988.
- 72. Castells M, Schwartz LB: Tryptase levels in nasal lavage fluid as an indicator of the immediate allergic response. J Allergy Clin Immunol 82:348, 1988.
- 73. Shalit M., Schwartz LB, Golzar N, et al: Release of histamine and tryptase in vivo after prolonged cutaneous challenge with allergen in humans. J Immunol 141:821, 1988.
- 74. Schwartz LB, Atkins PC, Bradford TR, et al: Release of tryptase together with histamine during the immediate cutaneous response to allergen. J Allergy Clin Immunol 80:850, 1987.
- 75. Butrus SI, Ochsner KI, Abelson MB, et al: The level of tryptase in human tears: an indicator of activation of conjunctival mast cells. Ophthalmology 97:1678, 1990.

- 76. Schechter NM, Fraki JE, Geesin JE, et al: Human skin chymotryptic protease. Isolation and relation to cathepsin G and rat mast cell proteinase, J Biol Chem 258:2973, 1983.
- 77. Caughey GH, Zerweck EH, Vanderslice P: Structure, chromosomal assignment, and deduced amino acid sequence of a human gene for mast cell chymase. J Biol Chem 266:12956, 1991.
- 78. Wintroub BU, Schechter NB, Lazarus GS, et al: Angiotensin I conversion by human and rat chymotryptic proteinases. J Invest Dermatol 83:335, 1984.
- 79. Reilly CF, Tewksbury DA, Schechter NM et al: Rapid conversion of angiotensin I to angiotensisn II by neutrophil and mast cell proteinases, J Biol Chem 257:8619, 1982.
- 80. Briggaman RA, Schechter NM, Fraki J et al: Degradation of the epidermal-dermal junction by a proteolytic enzyme from human skin and human polymorphonuclear leukocytes. J Exp Med 160:1027, 1984.
- 81. Goldstein SM, Kaempfer CE, Proud D et al: Detection and partial characterization of a human mast cell carboxypeptidase. J Immunol 139:2724, 1987.
- 82. Goldstein SM, Kaempfer CE, Kealey JT et al: Human mast cell carboxypeptidase. Purification and characterization, J Clin Invest 83:1630, 1989.
- 83. Reynolds DS, Stevens RL, Gurley DS et al: Isolatiion and molecular cloning of mast cell carboxypeptidase A. A novel member of the carboxypeptidase gene family. J Biol Chem 264:20094, 1989.
- 84. Reynolds DS, Gurley DS, Austen KF: Cloning and characterization of the novel gene for mast cell carboxypeptidase A. J Clin Invest 89:273, 1992.
- 85. Zon LI, Gurish MF, Stevens RL et al: GATA-binding transcription factors in mast cells regulate the promoter of the mast cell carboxypeptidase A gene. J Biol Chem 266:22948, 1991.
- 86. Reynolds DS, Gurley DS, Stevens RL et al: Cloning of cDNAs that encode human mast cell carboxypeptidase A, and comparison of the protein with mouse mast cell carboxypeptidase A and rat pancreatic carboxypeptidases. Proc Natl Acad Sci USA 86:9480, 1989.
- 87. Irani AA, Goldstein SM, Wintroub BU et al: Human mast cell carboxypeptidase. Selective localization to MC_{TC} cells. J Immunol 147:247, 1991.

- 88. Schwartz LB, Riedel C, Schratz JJ et al: Localization of carboxypeptidase A to the macromolecular heparin-proteoglycan-protein complex in secretory granules of rat serosal mast cells. J Immunol 128:1128, 1982.
- 89. Goldstein SM, Kaempfer CE, Wintroub BU: Human mast cell carboxypeptidase, Monogr Allergy 27:132, 1990.
- Goldstein SM, Kaempfer CE, Wintroub BU: Proteoglycan binding of human skin mast cell carboxypeptidase and other secretory granule proteases. J Invest Dermatol 92:435, 1989.
- 91. Thompson HL, Schulman ES, Metcalfe DD: Identification of chondroitin sulfate E in human lung mast cells. J Immunol 140:2708, 1988.
- 92. Stevens RL, Fox CC, Lichtenstein LM et al: Identification of chondroitin sulfate E proteoglycans and heparin proteoglycans in the secretory granules of human lung mast cells. Proc Natl Acad Sci USA 85:2284, 1988.
- 93. Metcalfe DD, Bland CE, Wasserman SI; Biochemical and functional characterization of proteoglycans isolated from basophils of patients with chronic myelogenous leukemia. J Immunol 132:1943, 1984.
- 94. Henderson WR, Kaliner M: Mast cell granules peroxidase; location, secretion, and SRS-A inactivation. J Immunol 122:1322, 1979.
- 95. Atkins PC, norman ME, Weiner H et al: Release of neutrophil chemotactic activity during immediate hypersensitivity reactions in humans. Ann Intern Med 86:415, 1977.
- 96. Nagy L, Lee TH, Kay AB: Neutrophil chemotactic activity in antigen-induced late asthmatic reactions. N Engl J Med 306:497, 1982.
- 97. Lee TH, Nagy L, Nagakura T et al: Identification and partial pruification of an exercise-induced neutrophil chemotactic factor in bronchial asthma. J Clin Invest 69:889, 1982.
- 98. Wasserman SI, Soter NA, Center DM et al: Recognition and characterization of a neutrophil chemotactic factor which appears in serum during experimental cold challenge. J Clin Invest 60:189, 1977.
- Soter NA, Wasserman SI, Pathkar MA et al: Solar urticaria: release of mast cell mediators into the circulation after experimental challenge. J Invest Dermatol 72:282, 1979.

- Soter NA, Wasserman SI, Austen KF et al: Release of mast mediators and alteration in lung function in patients with cholinergic urticaria. N Engl J Med 302:604, 1980.
- 101. Kay AB, Austen KF: IgE-mediated release of an eosinophil leukocyte chemotactic factor from human lung. J Immunol 107:899, 1971.
- 102. Goetzl EJ, Austen KF: Purification and synthesis of eosinophilotactic tetrapeptides of human lung tissue. Identification as eosinophil chemotactic factor of anaphylaxis. Proc Natl Acad Sci USA 72:4123, 1975.
- 103. Webb TA, Lin CY, Yan LT: Systemic mast cell disease: A clinical and hematopathologic study of 26 cases. Cancer 49:927, 1982.
- 104. Valone FH, Boggs JM, Goetzl EJ. Lipid mediators of hypersensitivity and inflammation. In *Allergy: Principles and Practice*. Edited by E Middleton, Jr, CE Reed, EF Ellis, NF Adkinson, Jr, JW Yunginger, WW Busse. St Louis, Mosby, pp 302-319, 1993.
- Lands WEM: The biosynthesis and metabolism of prostaglandins, Annu Rev Physiol 41:633, 1979.
- 106. Moncada S, Vane JR: Pharmacology and endogenous roles of prostaglandin endoperoxides, thromboxane A₂ and prostacyclin. Pharmacol Rev 30:293, 1978.
- 107. Andersen NH, Hartzell CJ, De B: Chemistry and structure of cyclooxygenasederived eicosanoids: a historical perpective. In Advances in Prostaglandin, Thromboxane and Leukotriene Research, Vol 14. Edited by JE Pike, DR Morton. New York, Raven Press, 1985.
- 108. Roberts LJ, Sweetman BJ, Lewis RA, Austen KF and Oates JA: Increased production of prostaglandin D₂ in patients with systemic mastocytosis. N Engl J Med 303:1400, 1980.
- 109. Goetzl EJ, Pickett WC: Novel structural determinants of the human neutrophil chemotactic activity of leukotriene B. J Exp Med 153:482, 1981.
- 110. Soter NA, Lewis RA, Corey EJ et al: Local effects of synthetic leukotrienes in human skin. J Invest Dermatol 80:115, 1983.
- 111. Lichtenstein, Schleimer RP, MacGlashan DW, Jr et al: Mediator release from human mast cells. In *Asthma: Physiology, Immunopharmacology, and Treatment*. New York, Academic Press, 1984.

- 112. Mills DC, MacFarlane DE: Stimulation of human platelet adenylate cyclase by prostaglandin D2. Thromb Res 5:401, 1974.
- 113. Beasley RCW, Featherstone, RL Church MK et al: Effect of a thromboxane receptor antagonist on PGD₂- and allergen-induced bronchoconstriction. J Appl Physiol 66:1685, 1989.
- 114. Dahlen S, Hansson G, Hedqvist P et al: Allergen challenge of lung tissues from asthmatics elicits bronchial contraction that correlates with the release of leukotrienes C₄, D₄, and E₄. Proc Natl Acad Sci USA 80:1712, 1983.
- Lynch JM, Worthen GS, Henson PM: Platelet-activating factor. In *Development of Anti-asthmatic Drugs*. Edited by DR Buckle, H Smith. London, Butterworth, 1984.
- 116. Pinckard RN, McManus LM, Hanahan DJ: Chemistry and biology of acetyl glyceryl ether phosphorylcholine (platelet-activating factor). In *Advances in Inflammation, Vol 4*. New York, Raven Press, 1982.
- Schleimer RP, MacGlashan DW, Peters SP et al: Characterization of inflammatory mediator release from purified human lung mast cells. Am Rev Respir Dis 133:614, 1986.
- Paul WE: Pleiotropy and redundancy: T cell-derived lymphokines in the immune response. Cell 57:521, 1989.
- Akira S, Hirano T, Taga T, Kishimoto T: Biology of multifunctional cytokines: IL-6 and related molecules (IL 1 and TNF). FASEB J. 4:2860, 1990.
- Costa JJ, Burd PR, Metcalfe DD: Mast Cell Cytokines. In The Mast Cell in Health and Disease. Edited by MA Kaliner, DD Metcalfe. New York, Marcel Dekker Inc, pp 443-466, 1993.
- 121. Brown MA, Pierce JH, Watson CJ et al: B cell stimulatory factor-1/interleukin-4 mRNA is expressed by normal and transformed mast cells. Cell 50:809, 1987.
- 122. Young JD-E, Liu C-C, Butler et al: Identification, purification, and characterization of a mast cell associated cytolytic factor related to tumor necrosis factor. Proc Natl Acad Sci USA 87:9175, 1987.
- 123. Gordon JR, Burd PR, Galli SJ: Mast cells as a source of multifunctional cytokines. Immunol Today 11:458, 1990.
- 124. Steffen M, Abboud M, Potter GK et al: Presence of tumour necrosis factor or a related factor in human basophil/mast cells. Immunology 66:445, 1989.

- 125. Klein LM, Lavker RM, Matis WL et al: Degranulation of human mast cells induces an endothelial antigen central to leukocyte adhesion. Proc Natl Acad Sci USA 86:8972, 1989.
- 126. Heusser C, Church MK: IL-4 in human mast cells. In XIVth International Congress of Allergology and Clinical Immunology, 1991.
- 127. Charlesworth, EN, Bochner BS, Lichtenstein LM et al: Interleukin-1 release during the cutaneous late-phase response to antigen. J Allergy Clin Immunol 85:266, 1990.
- 128. Massey W, Friedman B, Kato M et al: Allergen-induced late-phase reaction sites contain IL-3 and GM-CSF activity. J Allergy Clin Immunol 87:207, 1991.
- 129. Lee CE, Dixon PS and Charlesworth EN: Interleukin-6 is release in the cutaneous late phase response to antigen challenge. J. Allergy Clin Immunol 87:208, 1991.
- 130. Hamid Q, Ying S, Varney V et al: Localization of cells expressing mRNA for the cytokine gene cluster, IL-3, IL-4, IL-5, and GM-CSF in allergen-induced cutaneous late-phase reactions in atopic patients. J Allergy Clin Immunol 87:208, 1991.
- 131. Malveaux FJ, Conroy MC, Adkinson NF et al: IgE receptors on human basophils: relationship to serum IgE concentration. J Clin Invest 62:176, 1978.
- 132. Coleman JW, Godfrey RC: The number and affinity of IgE receptors on dispersed human lung mast cells. Immunology 44:859, 1981.
- 133. MacGlashan DW, Lichtenstein LM: Studies of antigen binding on human basophils. I. Antigen binding and functional consequences. J Immunol 180:2330, 1983.
- 134. Church MK, Caulfield JP: Mast cell and basophil functions. In *Allergy*. Edited by ST Holgate, MK Church. London, Gower Medical Publishing, pp 5.1-5.12, 1993.
- 135. Koch CA, Anderson D, Moran MF et al: Sh2 and SH3 domains: elements that control interactions of cytoplasmic signaling proteins. Science 252:668, 1991.
- 136. Finkel TH, Kubo RT, Cambier JC: T-cell development and transmembrane signaling: changing biological responses through an unchanging receptor. Immunol Today 12:79, 1991.
- 137. Cantley LC, Auger KR, Carpenter C et al: Oncogenes and signal transduction. Cell 64:281, 1991.

- 138. Hollenburg MD: Structure-activity relationships for transmembrane signaling: the receptor's turn. FASEB J 5:178, 1991.
- 139. Connelly PA, Farrell DA, Merenda JM et al: Tyrosine phosphorylation is an early signaling event common to Fc receptor crosslinking in human neutrophils and rat basophilic leukemia cells (RBL- 2H3). Biochem Biophys Res Commun 177:192, 1991.
- 140. Liotta M, Holowka D, Baird B: Evidence for the importance of tyrosine phosphorylation in IgE receptor mediated cellular degranulation . FASEB J 5:1676, 1991.
- 141. Stephan V, Benhamou M Gutkind SJ et al: IgE mediated tyrosine phosphorylation in RBL-2H3 cells: relationship to GTP-binding protein activation and phosphoinositol hydrolysis. FASEB J 5:A1676, 1991.
- 142. Aridor M, Traub LM, Sagi-Eisenberg R: Exocytosis in mast cells by basic secretagogues: evidence for direct activation of GTP binding proteins. J Cell Biol 111:909, 1990.
- 143. Aridor M, Sagi-Eisenberg R: Neomycin is a potent secretagogue of mast cells that directly activates a GTP-binding protein involved in exocytosis. J Cell Biol 111:2885, 1990.
- 144. Jozaki K, Kuriu A, Waki N et al: Proliferative potential of murine peritoneal mast cells after degranulation induced by compound 48/80, substance P, tetradecanoylphorbol acetate, or calcium ionophore A23187. J Immunol 145:4252, 1990.
- 145. Orloff DG, Ra C, Frank SJ et al: Family of disulfide linked dimers containing the ζ and n chains of the T cell receptor and the γ of Fc receptors. Nature 347:189, 1990.
- 146. Howard, Rodewald HR, Kinet JP, Reinherz EL: CD3ζ substitutes for Fc∈R1-γ in assembly and functional expression of the high affinity IgE receptor: evidence for inter-receptor complementation. Proc Natl Acad Sci USA 87:7015, 1990.
- 147. Gorski JP, Hugli TE and Muller HJ: The third anaphylatoxin of the human complement system. Proc Natl Acad Sci USA 76:5299, 1979.
- 148. Casale TB, Bowman S, Kaliner M: Induction of human cutaneous mast cell degranulation by opiates and endogenous opioid peptides: evidence for opiate and non-opiate receptor participation. J Allergy Clin Immunol 73:775, 1984.

- Metcalfe DD, Kaliner MA, Donolon MA: The mast cell. CRC Crit Rev Immunol 3:23, 1981.
- 150. Tharp MD, Welch MP, Sullivan TJ: Cutaneous mast cell stimulation by adenosine-triphosphate (ATP). J Invest Dermatol 78:355, 1982.
- 151. Asboe-Hansen G: Effect of the adrenocorticotropic hormone of the pituitary on mesenchymal tissues. Scan J Clin Lab Invest 2:271, 1950.
- 152. Tharp MD, Thirlby R, Sullivan TJ: Gastrin induces histamine release from human cutaneous mast cells. J Allergy Clin Immunol, 1984.
- 153. Casale TB, Keahey TM, Kaliner MA: Exercise induced anaphylactic syndromes. JAMA 225:2049, 1986.
- Huston DP, Bressler RB, Kaliner M et al: Prevention of mast cell degranulation by ketotifen in patients with physical urticarias, Ann Intern Med 104:507, 1986.
- 155. White MV: Mast cell secretagogues. In *The Mast Cell in Health and Disease*. Edited by MA Kaliner, DD Metcalfe. New York, Marcel Dekker, Inc, pp 109-128, 1993.
- 156. Tharp MD: The spectrum of mastocytosis. Southwestern Medical Center Internal Medicine Grand Rounds, 1984.
- 157. Sondergaard J, Asboe-Hansen G: Mastocytosis in childhood. In *Pediatric Dermatology*. Edited by R Happle, E Grosshans. Berlin, Springer-Verlag, pp 148-154, 1987.
- 158. Caplan RM: Urticaria pigmentosa and systemic mastocytosis. JAMA 194:175,
- 159. Caplan RM: The natural course of urticaria pigmentosa. Arch Dermatol 87:146, 1963
- 160. Longley JB, Morganroth GS, Tyrrell L et al: Altered metabolism of mast cell growth factor (c-kit Ligand) in cutaneous mastocytosis. N Engl J Med 328:1302, 1993.
- 161. Tsai M, Shih L-S, Newlands GFJ et al: The rat c-kit ligand, stem cell factor, induces the development of connective tissue type and mucosal mast cells in vivo: analysis by anatomical distribution, histochemistry and protease phenotype. J Exp Med 174:125, 1991.

- 162. Anderson DM, Williams DE, Tushinski R et al: Alternate splicing of mRNAs encoding human mast cell growth factor and localization of the gene to chromosome 12q22-q24. Cell Growth Differ 2:373, 1991.
- 163. Lu HS, Clogston CL, Wypych J et al: Amino acid sequence and posttranslational modification of stem cell factor isoloated from buffalo rat liver cell conditioned medium. J Biol Chem 266:8102, 1991.
- 164. Flanagan JG, Chan DC, Leder P: Transmembrane form of the kit ligand growth factor is determined by alternative splicing and is missing in the SL^d mutant. Cell 64:1025, 1991.
- 165. Huang EJ, Nocka KH, Buck J, Besmer P: Differential expression and processing of two cell associated forms of the kit ligand: KL-1 and KL-2. Mol Biol Cell 3:349, 1992.
- 166. Metcalfe DD: Classification and diagnosis of mastocytosis: current status. J Invest Dermatol 96:2S, 1991.
- 167. Parwaresch MR, Horny H-P, Lennert K: Tissue mast cells in health and disease. Pathol Res Pract 179:439, 1985.
- 168. Travis WD, Li C-Y, Bergstralh EJ, Yam LT, Swee RG: Systemic mast cell disease. Analysis of 58 cases and literature review. Medicine 67:345, 1988.
- 169. Metcalfe DD: Conclusions of a roundtable discussion on mastocytosis. J Invest Dermatol 96:64S, 1991.
- 170. Bloom G, Duner H, Pernow B, Winberg J, Zettepstrom R: Spontaneous histamine shock in urticaria pigmentosa. Acta Ped 47:152, 1958.
- 171. Demis DJ: The mastocytosis syndrome: Clinical and biological studies. Ann Int Med 59:194, 1963.
- 172. Demis, Walton MD, Higdon RS: Histaminuria in urticaria pigmentosa. A clinical study and review of recent literature with definition of the mastocytosis syndrome. Arch Dermatol 83:127, 1961.
- 173. Orkin M, Good RA Clawson CC Fisher I, Windhorst DB: Bullous mastocytosis. Arch Dermatol 101:547, 1970.
- 174. Robinson HM Jr, Kile RL, Hitch JM, Robinson RCV: Bullous urticaria pigmentosa. Arch Dermatol 85:346, 1962.

- 175. Soter NA: The skin in mastocytosis. J Invest Dermatol 95:32S, 1991.
- 176. Czarnetzki BM, Kolde G, Schoemann A, Urbanitz S, Urbanitz D: Bone marrow findings in adult patients with urticaria pigmentosa. J Am Acad Dermatol 18:45, 1988.
- 177. Ridel B, Olafsson JH, Roupe G: The bone marrow in urticaria pigmentosa and systemic mastocytosis. Arch Dermatol 122:422, 1986.
- 178. Klaus SN and Winkelmann RK: Course of urticaria pigmentosa in children. Arch Derm 86:68, 1962.
- 179. Sagher F, Even-Paz Z: *Mastocytosis and the Mast Cell.* Chicago, Yearbook Medical Publishers, 1967.
- 180. Chagrin L, Sachs P: Urticaria pigmentosa appearing as a solitary nodular lesion. Arch Dermatol Syphilol 69:345, 1954.
- Demis DJ: Mast Cell Disease. In Clinical Dermatology. Edited by DJ Demis, RL Dobson, J McGuire. Philadelphia, Harper and Row, 1982.
- 182. Golitz LE, Weston WL, Lane AT: Bullous mastocytosis: diffuse cutaneous mastocytosis with extensive blisters mimicking scalded skin syndrome or erythema multiforme. Pediatr Dermatol 1:288, 1984.
- 183. Kasper CS, Tharp MD: Quantification of cutaneous mast cells using morphometric point counting and a conjugated avidin stain. J Am Acad Dermatol 16:326, 1987.
- 184. Craig SS, Schechter NM, Schwartz LB: Ultrastructural analysis of human T and TC mast cells identified by immunoelectron microscopy. Lab Invest 58:682, 1988.
- 185. Tharp MD, Glass MJ, Seelig Jr LL: Ultrastructural morphometric analysis of lesional skin: mast cells from patients with systemic and nonsystemic mastocytosis. J Am Acad Dermatol 18:298, 1988.
- 186. Friedman BS, Steinberg SC, Meggs WJ et al.:Analysis of plasma histamine levels in patients with mast cell disorders. Am J Med 87:649, 1989.
- 187. Keyzer JJ, De Monchy JGR, Van Doormaal JJ et al: Improved diagnosis of mastocytosis by measurement of urinary histamine metabolites. N Engl J Med 309:1603, 1983.

- 188. Muller UR, Horat W, Wuthrich B et al: Anaphylaxis after hymenoptera stings in three patients with urticaria pigmentosa. J Allergy Clin Immunol 72:685, 1983.
- 189. Frieri M, Alling DW, Metcalfe DD: Comparison of the therapeutic efficacy of cromolyn sodium with that of combined chlorpheniramine and cimetidine in systemic mastocytosis: results of a double-blind clinical trial. Am J Med 78:9, 1985.
- 190. Soter NA, Austen KF, Wasserman SI: Oral disodium cromoglycate in the treatment of systemic mastocytosis. N Engl J Med 301:465, 1979.
- Czarnetzki BM, Behrendt H: Urticaria pigmentosa: clinical picture and response to oral disodium cromoglycate. Br J Dermatol 105:563, 1981.
- 192. Evans S, Vickers CFH: Bullous urticaria pigmentosa (cutaneous mastocytosis) and sodium cromoglycate therapy. Acta Derm Venereol (Stockh) 61:572, 1982.
- 193. Welch EA, Alper JC, Bogaars H, Farrell DS: Treatment of bullous mastocytosis with disodium cromoglycate. J Am Acad Dermatol 9:349, 1983.
- 194. Czarnetzki BM: A double blind cross-over study of the effect of ketotifen in urticaria pigmentosa. Dermatologica 166:44, 1983.
- 195. Guinot P, Summerhayes C, Berdah L et al: Treatment of adult systemic mastocytosis with a PAF-acether antagonist BN52063 (letter). Lancet II:114, 1988.
- 196. Czarnetzki PM, Rosenbach T, Kolde G, Frosch PJ: Phototherapy of urticaria pigmentosa: clinical response and changes of cutaneous reactivity, histamine and chemotactic leukotrienes. Arch Dermatol Res 277:105, 1985.
- 197. Granerus G, Roupe G, Swanbeck G: Decreased urinary histamine metabolite after successful PUVA treatment of urticaria pigmentosa. J Invest Dermatol 76:1, 1981.
- 198. Christophers E, Honigsmann H, Wolff K: PUVA treatment of urticaria pigmentosa. Br J Dermatol 98:701, 1978.
- 199. Vella Briffa D, Eady RAJ, James MP et al: Photochemotherapy (PUVA) in the treatment of urticaria pigmentosa. Br J Dermatol 109:67, 1983.
- 200. Barton J, Lavker RM, Schecter NM, Lazarus GS: Treatment of urticaria pigmentosa with corticosteroids. Arch Dermatol 121:1516, 1985.

- 201. Horan RF, Austen KF: Systemic mastocytosis: Retrospective review of a decade's clinical experience at the Brigham and Women's Hospital. J Invest Dermatol 96:5S, 1991.
- 202. Travis WD, Chin-Yang L: Mast cell disease. In The Mast Cell in Health and Disease. Edited by MA Kaliner, DD Metcalfe. New York, Marcel Dekker, Inc, pp 723-741, 1993.
- 203. Metcalfe DD: Mastocytosis syndromes. In *Allergy: Principles and Practice*. Edited by E Middleton, Jr, CE Reed, EF Ellis, NF Adkinson, Jr, JW Yunginger, WW Busse. St Louis, Mosby, pp 1537-1551, 1993.
- 204. Havard CWH, Scott RB: Urticaria pigmentosa with visceral and skeletal lesions. Q J Med 28:459, 1959.
- 205. Poppel MH, Gruber WF, Silber R et al: The roentgen manifestations of urticaria pigmentosa (mastocytosis). Am J Roentgenol 82:239, 1959.
- 206. Goldhaber P: Heparin enhancement of factors stimulating bone resorption in tissue culture. Science 147:407, 1965.
- 207. Raisz LG, Martin TJ; Prostaglandins in bone mineral metabolism. In *Bone and Mineral Research*. Edited by WA Peck. Amsterdam, Elsevier, pp 286-310, 1983.
- 208. Hawkins RA, Claman HN, Clark RAF et al: Increased dermal mast cell populations in progressive systemic sclerosis: a link in chronic fibrosis. Ann Intern Med 102:182, 1985.
- 209. Hirson C: Urticaria pigmentosa with disseminated mastocytosis controlled with chlorambucil. Proc R Soc Med 58:697, 1965.
- 210. Ives DR, Thompson DM: Urticaria pigmentosa with spinal osteoporosis. Proc R Soc Med 66:175, 1973.
- 211. Chines A, Pacifici R, Avioli LV, Teitelbaum SL, Korenblat PE: Systemic mastocytosis presenting as osteoporosis: as clinical and histomorphometric study. J Clin Endocrinol Metab 72:140, 1991.
- 212. deGennes C, deVernejoul MC: Bone involvement in mastocytosis: a report of 9 cases. Calcil Tissue Int 44:S-82, 1989.
- 213. Lidor C, Gepstein R, Prisch B et al: Skeletal mastocytosis presenting as osteoporosis. Calcif Tissue Int 44:S-61, 1989.
- 214. McKenna MJ, Frame B. The mast cell and bone. Clin Orthop 200:226, 1985.

- 215. Rafii M, Firooznia H, Golimbu C, Balthazar E: Pathologic fracture in systemic mastocytosis: radiographic spectrum and review of the literature. Clin Orthop 180:260, 1983.
- 216. Lawrence JB, Friedman BS, Travis WD et al: Hematologic manifestations of systemic mast cell disease: a prospective study of laboratory and morphologic features and their relation to prognosis, Am J Med 91:612, 1991.
- 217. Horny HP, Parwaresch MR, Lennert K: Bone marrow findings in systemic mastocytosis. Hum Pathol 16:808, 1985.
- 218. Parker RI: Hematologic aspects of mastocytosis: I: Bone marrow pathology in adult and pediatric systemic mast cell disease. J Invest Dermatol 96:47S, 1991.
- 219. Bennett JM, Catovsky D, Daniel MT et al: Proposed revised criteria for the classification of acute myeloid leukemia: a report of the French-American-British cooperative group. Ann Intern Med 103:620, 1985.
- 220. Laszlo J; Myeloproliferative disorders (MPD): myelofibrosis, myelosclerosis, extrameduallary hematopoiesis, undifferentiated MPD, and hemorrhagic thrombocythemia. Semin Hematol 12:409, 1975.
- 221. Non-Hodgkin's lymphoma pathologic classification project: National Cancer Institute sponsored study of classifications of non-Hodgkin's lymphomas:summary and description of a working formulation for clinical usage. Cancer 49:2112, 1982.
- 222. Travis WD, Li C-Y, Yam LT, Bergstrahl MS, Swee RG: Significance of systemic mast cell disease with associated hematologic disorders. Cancer 62:965, 1988.
- Parker RI: Hematologic aspects of mastocytosis: II: Management of hematologic disorders in association with systemic mast cell disease. J Invest Dermatol 96:52S, 1991.
- 224. Lennert K: Zur pathologischen anatomie von urticaria pigmentosa und mastzellenreticulose. Klin Wschv 40:61, 1962.
- 225. Horny H-P, Kaiserling E, Campbell M et al: Liver findings in generalized mastocytosis. Cancer 63:532, 1989.
- 226. Yam LT, Chan CH, Li C-Y: Hepatic involvement in systemic mast cell disease. Am J Med 80:819, 1986.
- 227. Horny H-P, Ruck MT, Kaiserling E: Spleen findings in generalized mastocytosis. Cancer 70:459, 1992.

- 228. Travis WD, Li C-Y: Pathology of the lymph node and spleen in systemic mast cell disease. Modern Pathol 1:4, 1988.
- 229. Horny H-P, Kaiserling E, Parwaresch MR, Lennert K: Lymph node findings in generalized mastocytosis. Histopathology 21, 439, 1992.
- 230. Cherner JA, Jensen R T, Dubois A et al: Gastrointestinal dysfunction in systemic mastocytosis, a prospective study. Gastroenterology 95:657, 1988.
- 231. Ammann RW, Vetter D, Dehyle P et al: Gastrointestinal involvement in systemic mastocytosis. Gut 17:107, 1976.
- 232. Bank S, Marks IN: Malaborption in systemic mast cell disease. Gastroenterology 45:535, 1963.
- 233. Bredfeldt JE, O'Laughlin JC, Durham JB et al: Malabsorption and gastric hyperacidity in systemic mastocytosis. Am J Gastroenterol 74:133, 1980.
- 234. Garriga MM, Friedman MM, Metcalfe DD: A survey of the number and distribution of mast cells in the skin of patients with mast cell disorders. J Allergy Clin Immunol 82:425, 1988.
- 235. Roberts LJ II, Oates JA: Biochemical diagnosis of systemic mast cell disorders. J Invest Dermatol 96:19, 1991.
- 236. Beavan MA, Jacobsen S, Horakova Z: Modification of the enzymatic isotopic assay of histamine and its applications to measurement of histamine in tissues, serum, and urine. Clin Chim Acta 37:91, 1972.
- 237. Shaff RE, Beaven MA: Increased sensitivity of the enzymatic assay of histamine: measurement of histamine in plasma and serum. Anal Biochem 94:425, 1979.
- Dyer J, Warren K, Merlin S, Metcalfe DD, Kaliner M: Measurement of plasma histamine: description of an improved method and normal values. J Allergy Clin Immunol 70:82, 1982.
- 239. McBride PT, Bradley D, Kaliner MA: Evaluation of a radioimmunoassay for histamine measurement in biological fluids. J Allergy Clin Immunol 82:638, 1988.
- 240. Gleich GJ Hull WM Jr: Measurement of histamine: a quality control study. J Allergy Clin Immunol 66:295, 1980.

- 241. Warren K, Dyer J, Merlin S, Kaliner M: Measurement of urinary histamine: comparison of fluorometric and radioisotopic-enzymatic assay procedures. J Allergy Clin Immunol 71:206, 1983.
- 242. Roberts LJ II, Aulsebrook KA, Oates JA: Comparative evaluation of the radioenzymatic method for the determination of urinary histamine with a mass spectrometric assay. J Chromatogr 338:41, 1985.
- 243. Granerus G, Olafsson JH, Roupe G: Studies on histamine metabolism in mastocytosis. J Invest Dermatol 80:410, 1983.
- 244. Roberts LJ II: Quantification of the PGD_2 urinary metabolite 9α -hydroxy-11,15-dioxo-2,3,18,19-tetranorprost-5-ene-1,20-dioic acid by stable isotope dilution mass spectrometric assay. Methods Enzymol 86:559, 1982.
- 245. Liston TE, Roberts LJ II: Metabolic fate of radiolabelled prostaglandin D₂ in a normal human male volunteer. J Biol Chem 260:13172, 1985.
- 246. Guillet GY, Dore N, Maleville J: Heparin liberation in urticaria pigmentosa. Arch Dermatol 118:532, 1982.
- Campbell EW, Hector D, Gossain V: heparin activity in systemic mastocytosis.
 Ann Intern Med 90:940, 1979.
- 248. Schwartz LB, Yunginger JW, Miller J et al: Time course of appearance and disappearance of human mast cell tryptase in the circulation after anaphylaxis. J Clin Invest 38:1551, 1989.
- 249. Hirschowitz BI, Groarke JF: Effect of cimetidine on gastric hypersecretion and diarrhea in systemic mastocytosis. Ann Int Med 90:769, 1979.
- 250. Gerrard JW and Ko C: Urticaria pigmentosa: Treatment with cimetidine and chlorpheniramine. J of Peds 94:843, 1979.
- 251. Simon RA: Treatment of mastocytosis. N Engl J Med 302:231, 1980.
- 252. Roberts LJ, Sweetman BJ, Lewis RA, Austen KF, Oates JA: Increased production of prostaglandin D₂ in patients with systemic mastocytosis. N Engl J Med 303:1400, 1980.
- 253. Metcalfe DD: The treatment of mastocytosis: An overview. J Invest Dermatol 96:55S, 1991.

- 254. Roberts LJ, Fields JP, Oates JA: Mastocytosis without urticaria pigmentosa: A frequently unrecognized cause of recurrent syncope. Trans Assoc Amer Phys 95:36, 1982.
- 255. Horan RF, Sheffer AL, Austen KF: Cromolyn sodium in the management of systemic mastocytosis. J Allergy Clin Immunol 85:852, 1990.
- 256. Kettelhut BV, Metcalfe DD: A double blind placebo controlled trial of ketotifen versus hydroxyzine in the treatment of pediatric mastocytosis. J Allergy Clin Immunol 83:866, 1989.
- 257. Reisberg IR, Oyakawa S: mastocytosis with malabsorption, myelofibrosis, and massive ascites. Am j Gastroent 82:52, 1987.
- 258. Friedman BS, Metcalfe DD: Effects of tixocortol pivalate on gastrointestinal disease in systemic mastocytosis: a preliminary study. Clin Exp Allergy 21:183, 1991.
- 259. Friedman B, Darling G, Norton J, Hamby L, Metcalfe DD: Splenectomy in the management of systemic mast cell disease. Surgery 107:94, 1990.
- 260. Kors JW, van Doormaal JJ, Monchy JGR: Anaphylactoid shock following hymenoptera sting as a presenting symptom of systemic mastocytosis. J Int Med 233:255, 1993.
- 261. Kluin-Nelemans HC, Jansen JH, Breukelman H et al: Response to interferon alfa-2b in a patient with systemic mastocytosis. N Engl J Med 326:619, 1992.
- 262. Madden KB, Urban J, Ziltener HJ et al: Antibodies to IL-3 and IL-4 suppress helminth-induced intestinal mastocytosis.