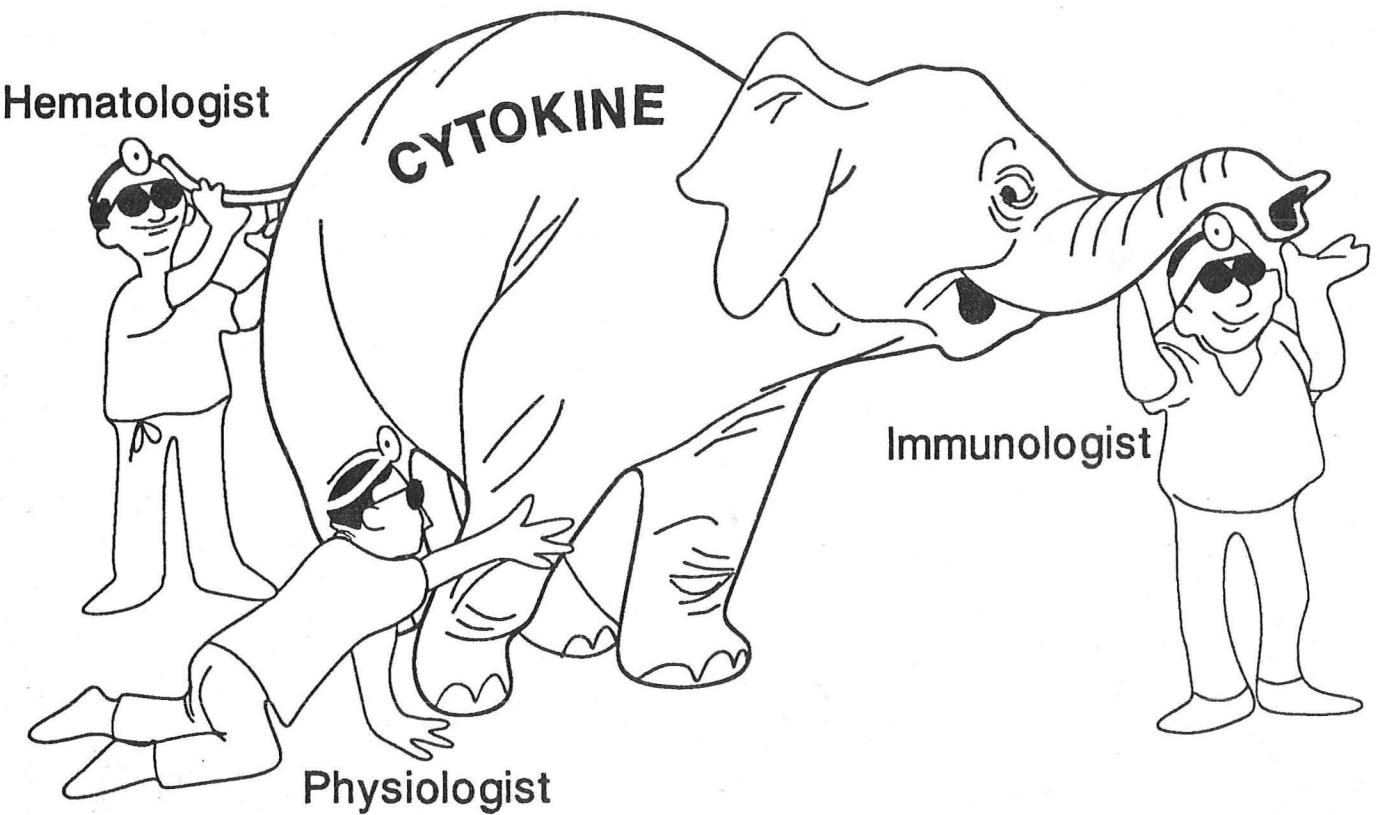


Immuno.

CYTOKINES: MESSENGER MOLECULES OF THE
IMMUNE SYSTEM



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Medical Grand Rounds

Thursday, March 24, 1988

A BRIEF HISTORY OF THE CYTOKINES

I. The Age of Acronyms

- 1966 *David, Bloom & Bennett* - Macrophage migration inhibition factor (MIF) described
 - small molecular substance that inhibited the migration of macrophages
- 1969 *Dumonde* - "Lymphokines"
 - soluble antigen nonspecific products of lymphocytes that mediated the manifestations of delayed-type hypersensitivity reactions
- 1974 *Cohen* - "Cytokine"
 - Soluble products from cells other than lymphocytes also effected immune responses
- 1979 *International Lymphokine Workshop* - "Interleukins"
 - Substances that communicated signals between different populations of leukocytes

II. The Age of Molecular Biology

- 1982 *Gray & Goeddel* - Gene for gamma interferon cloned
- 1983-> present Gene for many cytokines cloned -
 Interleukins 1 α , 1 β , 2, 3, 4, 5, 6
 GM-CSF, G-CSF, M-CSF
 TNF α , LT

REFERENCES

1. Bloom, B.R. and Bennett, B. 1966. Mechanism of a reaction *in vitro* associated with delayed-type hypersensitivity. *Science* 153:80-82.
2. David, J. 1966. Delayed hypersensitivity *in vitro*: Its mediation by cell-free substances formed by lymphoid cell-antigen interactions. *Proc. Natl. Acad. Sci. USA* 56:72-77.
3. Dumonde, D.C., Wolstencroft, R.A., Panayi, G.S., Matthew, M., Morley, J., and Howson, W.T. 1969. "Lymphokines": Non-antibody mediators of cellular immunity generated by lymphocyte activation. *Nature* 224:38-42.
4. Cohen, S., Bigazzi, P.E., and Yoshida, T. 1974. Similarities of T cell function in cell-mediated immunity and antibody production. *Cell. Immunol.* 12:150-159.

5. Waksman, B.H., and Namba, Y. 1976. On soluble mediators of immunologic regulation. *Cell. Immunol.* 21:161-176.
6. Aarden, L.A., et al. 1979. Revised nomenclature for antigen-nonspecific T cell proliferation and helper factors. *J. Immunol.* 123:2928-2929.

ACTIVITIES ATTRIBUTED TO CYTOKINES

AEF	HCGF	MIF
BAF	HPGF	MP
BCDF	HSF-II	Meg-CSF
BCDF γ	Hemopoietin I	Multi-CSF
BCGF-I	IFN- β 2	NIF-T
BCGF-II	IgA-EF	OAF
BPA	IL-HP1	PIF
BSF-1	Interferon α	PSF
BSF-2	Interferon β	Pluripoietin
CSF-1	Interferon γ	SIRS
CSF-2	KHF	SMAF
CSF α	LAF	SRF
CSF β	LEM	TCGF
Cachectin	LDCF	TCGF-2
Catabolin	LIF	T _H F
ECSF	Lymphotoxin	TNF
EDF	MAF	TRF
EP	MCF	TRF _M
ETAF	MCGF	T _S F
Eo-CSF	MCGF-2	Urinary CSF
GRF	MFF	26Kd protein

CYTOKINES WHOSE GENES HAVE BEEN CLONED AND EXPRESSED

A. Primarily Monocyte/Macrophage Derived

1. Interleukin 1 α
2. Interleukin 1 β
3. Tumor necrosis factor α

B. Primarily T Lymphocyte-Derived

1. Interleukin 2
2. Interleukin 4
3. Interleukin 5
4. Interleukin 6
5. Lymphotoxin
6. Interferon γ

C. Hemopoietic Growth Factors

1. Interleukin 3
2. Granulocyte Macrophage Colony Stimulating Factor
3. Macrophage Colony Stimulating Factor
4. Granulocyte Colony Stimulating Factor

D. Factors primarily produced by cells outside the immune system or whose major action is not immunologic

1. Interferon α
2. Interferon β
3. Transforming Growth Factor β
4. Platelet Derived Growth Factor

Interleukin 1 α (IL1 α)
Interleukin 1 β (IL1 β)

Other names:

Lymphocyte Activating Factor (LAF)
 B Cell Activating Factor (BAF)
 Leukocyte Endogenous Mediator (LEM)
 Endogenous Pyrogen (EP)
 Hemopoietin I
 Catabolin
 Mononuclear Cell Factor (MCF)
 Osteoclast Activating Factor (OAF)
 Epidermal Cell Derived Thymocyte Activating Factor (ETAF)

Natural Products:

IL1 α	protein, Mr 17,000	pI=5.5
IL1 β	protein, Mr 17,000	pI=7.0

Deduced Size of Mature Protein:

IL1 α	159 amino acids	Mr 17,500
IL1 β	153 amino acids	Mr 17,500

Gene:

Chromosome: single copy genes, chromosome 2

Organization: Seven exons, six introns

IL1 α - 10.2 Kb

IL1 β - 7.0 Kb

Cellular Sources:

Cell	Stimulus
Monocytes, macrophages	bacterial endotoxin, muramyl dipeptide immune complexes, C5a
Keratinocytes	cytokines (M-CSF, TNF α , IFN γ) muramyl dipeptide, bacterial endotoxin phorbol esters
Langerhans cells	bacterial endotoxin
Large granular lymphocytes	bacterial endotoxin
Endothelial cells	bacterial endotoxin, thrombin
Mesangial cells	-
Astrocytes	bacterial endotoxin

Glioma cell lines
Microglial cells - bacterial endotoxin

Receptors:

Specificity: IL1 α or IL1 β , human material is active in mice

Avidity: Murine T cells - high avidity: $K_d=5\times 10^{-12} M$; low avidity: $K_d=4\times 10^{-10} M$
 Human fibroblasts - $K_d=8\times 10^{-12} M$
 Human EBV transformed B cells- $K_d=2-6\times 10^{-10} M$

Number: Murine T cells - resting: high avidity, 12/cell
 activated: low avidity, 180/cell
 high avidity, 23/cell
 low avidity, 580/cell
 Murine T cell tumors: high avidity, 46-340/cell
 low avidity, 2550-17,700/cell
 Human fibroblasts: 3000/cell
 Human EBV transformed B cells: 110-220/cell
 Other cells: 20-5000/cell

Size: Mr 80,000

Organization: Single chain

Distribution: T cells, B cell lines, fibroblasts, endothelial cells, melanoma cells, hepatoma cells, epithelial cells.

Known Activities:

Cell	Activity
T Lymphocytes	Co-mitogenic with lectins Increase lymphokine production (IL2, IFN γ) Augment activity of cytotoxic T cells Increase receptors for IL-2
B Lymphocytes	Maturation of pre-B cells Cofactor in clonal expansion Promote differentiation of antibody forming cells
NK Cells	Enhance killing and synergistically enhance killing with IFN- γ

Macrophages	Increase production of prostaglandin Increase tumoricidal activity
Hemopoietic Cells	Facilitate response of immature precursors to colony stimulating factors
Smooth Muscle Cells	Stimulate growth Stimulate prostaglandin production
Endothelial Cells	Increase proliferation Increase prostacyclin, prostaglandin, platelet activating factor, GM-CSF, G-CSF production Increase adhesiveness for polymorphonuclear leukocytes, monocytes, lymphocytes Increase procoagulant and plasminogen activator activity Induce new surface antigens
Fibroblasts	Increase PGE ₂ production Increase collagenase production Increase proliferation Increase production of GM-CSF, G-CSF, IL6
Chondrocytes	Increase production of collagenase, neutral proteases, plasminogen activator Increase production of PGE ₂
Non-neuronal cells of nerves	Stimulate production of Nerve Growth Factor
Hepatocytes	Increase synthesis of complement factors B and C3, metallothionein, α_1 - antichymotrypsin, α_1 acid glycoprotein, inter- α_1 -trypsin inhibitor, serum amyloid associated protein Decrease synthesis of albumin, transferrin, fibrinogen Decrease activity of cytochrome P-450
Mesangial Cells	Increase proliferation
Osteoblasts	Increase proliferation
Osteoclasts	Increase bone resorption Decrease alkaline phosphatase
Epidermal Cells	Increase synthesis of collagen type IV
Adipocytes	Decrease lipoprotein lipase activity

Tumor Cells	Cytostasis
Glial Cells	Increase proliferation
Pancreatic Cells	Cytotoxicity
Thyroid Cells	Cytotoxicity
Pituitary Cells	Stimulate secretion of ACTH, LH, GH, TSH

Activity *in vivo*:

Animals

1. Fever
2. Induce acute phase reactants from liver
3. Slow wave sleep
4. Anorexia
5. Induce colony stimulating activity
6. Accelerate recovery from Listeriosis
7. Protect granulocytopenic mice from lethal gram-negative infection
8. Protect from lethal irradiation
9. Protect from oxygen damage (with TNF α)
10. Downregulate hepatic steroid receptors, reduce gluconeogenesis and decrease plasma glucose
11. Stimulate secretion of corticotropin-releasing factor from the hypothalamus
12. Intra-articular injection causes loss of cartilage matrix and synovitis.

Man

1. Found in synovial fluid of patients with various forms of arthritis
2. Found in gingival fluid of patients with periodontal disease

Tumor Necrosis Factor α (TNF α)

Other Names:

Cachectin

Natural Product:

Protein, Mr 17,000

Deduced Size of Mature Protein:

157 amino acids, Mr 17,300

Gene:

Chromosome: Single copy gene, chromosome 6 (p23-q12)
 Organization: No information

Cellular Sources:

Macrophages activated by bacterial endotoxin, M-CSF, IFN- γ
 Activated T cells, mast cells

Receptors:

Specificity: TNF α or LT, human material is active in rodents

Avidity: $K_d = 2 \times 10^{-10} M$

Number: 200-7500/cell, increased by IFN- γ

Size: Mr 140,000

Organization: 2 polypeptides: Mr 80,000 and Mr 60,000

Distribution: Fibroblasts
 Activated T lymphocytes
 Activated B lymphocytes
 Many tumor cells

Known Activities

Cell	Activity
T Cells	Enhance proliferation, IL2 receptor expression
B Cells	Enhance proliferation and differentiation of activated B cells into antibody forming cells

NK Cells	Enhance cytolytic potential, synergize with IL2
Hepatocytes	Increase production of complement factors B, C3, α 1 antichymotrypsin Decrease production of albumin, transferrin
Hemopoietic Stem Cells	Inhibition of proliferation and differentiation
Endothelial Cells	Enhance production of IL1, GM-CSF, G-CSF, platelet activating factor, procoagulant activity, neutrophil adherence, new antigen expression
Fibroblasts	Stimulate production of PGE ₂ , collagenase, GM-CSF, G-CSF, membrane IL1, IL6
Adipocytes	Inhibit lipoprotein lipase
Osteoclasts	Increase bone resorption, decrease alkaline phosphatase
Neutrophils	Stimulate phagocytosis, respiratory burst, degranulation and production of platelet activating factor
Tumor Cells	Cytotoxicity
Macrophages	Induce IL1, PGE ₂ and platelet activating factor production Chemotactic

Activity *in vivo*:

Animals

1. Fever
2. Induce colony stimulating activity
3. Shock
4. Wasting
5. Necrosis of certain transplantable tumor
6. Protect from oxygen damage (with IL1)
7. Prevent nephritis in (NZBxNZW)F₁ mice
8. Antibody to TNF α prevents:
 - a. Skin and gut lesions in acute graft versus host disease
 - b. Neurological manifestations of malaria (murine cerebral malaria)
 - c. Death from lethal doses of endotoxin

Man

1. Identified in synovial fluid of patients with rheumatoid arthritis

Interleukin 2 (IL2)

Other Names:

T cell growth factor (TCGF)

Natural Product:

Glycoprotein (O-linked sugar only): Mr 15-17,000

Deduced Size of Mature Protein:

133 amino acids, Mr 15,400

Gene:

Chromosome: Single copy gene, chromosome 4 (q26-q28)

Organization: 4 exons, 3 introns (5 Kb)

Cellular Sources:

Activated T cells

Some activated B cells

Receptors:

Specificity: IL2 only, human material is active in the mouse

Avidity: Low avidity: $K_d = 1-3 \times 10^{-8} M$

Intermediate avidity: $K_d = 1 \times 10^{-9} M$

High avidity: $K_d = 2-20 \times 10^{-12} M$

Number: Low avidity: $3-6 \times 10^4$ /activated T cell, 0/resting T cell

Intermediate avidity: 0/activated T cell, 700/resting T cell,
2000/NK cell

High avidity: $3-4 \times 10^3$ /activated T cell, 0/resting T cell

Size: Low avidity: Mr 55,000

Intermediate avidity: Mr 70,000

High avidity: Mr 125,000

Organization: Low avidity: single chain (p55)

Intermediate avidity: single chain (p70)

High avidity: heterodimer (p55 + p70)

Gene: p55 - chromosome 10 (p14-p15)
 8 exons, 7 introns (>25Kb)
 p70 - no information

Distribution:

Low avidity: Activated T cells, activated B cells,
 IFN- γ activated macrophages

Intermediate avidity: Resting T cells, NK cells

High avidity: Activated T cells, activated B cells

Known Actions:

Cell	Action
T Cells	Stimulate the growth of activated T cells Stimulate functional differentiation of activated T cells to cytotoxic effector cells
B Cells	Stimulate the growth of activated B cells Stimulate functional differentiation of activated B cells to antibody secreting cells
NK Cells	Enhance lytic potential
Macrophages	Functional activation

Activity *in vivo*

Animals

1. Protect mice from lethal gram negative infection
2. Polyclonal activation of T and B cells
3. Promote the generation of specific cytotoxic T cells

Man

1. Found in synovial fluid of patients with rheumatoid arthritis
2. Results of intravenous administration
 - a) Two to sixteen-fold increase in circulating T lymphocytes (CD4+ and CD8+)
 - b) Increase circulating T cells with IL2 receptors
 - c) Increase plasma levels of γ interferon

Interleukin 4 (IL4)

Other Names:

B cell stimulatory factor 1 (BSF-1)
 B cell growth factor I (BCGF-I)
 B cell differentiation factor γ (BCDF γ)
 T cell growth factor 2 (TCGF-2)
 Mast cell growth factor-2 (MCGF-2)

Natural Product:

Mouse: Glycoprotein, Mr 20,000
 Human: No information

Deduced Size of Mature Protein:

129 amino acids, Mr 15,000

Gene: No information

Cellular Sources:

Activated T cells
 Mast cells (mouse)

Receptors:

<u>Specificity:</u>	IL4 only, species specific
<u>Avidity:</u>	$K_d = 0.5-1.0 \times 10^{-10} M$
<u>Number:</u>	100-2500/cell
<u>Size:</u>	Mr 139,000
<u>Organization:</u>	Single chain
<u>Distribution:</u>	T lymphocytes, B lymphocytes, Mast cells, Macrophages, Myeloid precursors, Erythroid precursors, Fibroblasts, Melanocytes, Keratinocytes, Brain cells, Muscle cells, Hepatocytes, Pancreatic cells, Bladder cells

Known Actions:

Cell	Activity
B Cells	Enhance proliferation of activated cells Increase expression of class II MHC molecules on murine B cells Increase expression of low affinity receptors for IgE (CD23)

	Promote production of IgG1, IgE from stimulated murine B cells
T Cells	Promote growth of activated T cells Promote generation of antigen specific cytotoxic T cells
Mast Cells	Support proliferation of murine mast cells
Macrophages	Enhance tumoricidal activity Increase expression of class II MHC molecules on murine macrophages Enhance antigen presenting capacity of murine macrophages Induce giant multinucleated cells from murine macrophages
Hemopoietic precursors	Costimulate growth of murine erythroid, macrophage, granulocyte and mast cell colonies

Activity *in vivo*:

Animals

1. Antibody to IL4 abrogates the induction of IgE responses in the mouse

Man

No information

Interleukin 5 (IL-5)

Other Names:

B cell growth factor II (BCGF-II)
 T cell replacing factor (TRF)
 Killer helper factor (KHF)
 Eosinophil differentiation factor (EDF)
 Eosinophil-colony stimulating factor (Eo-CSF)
 IgA-enhancing factor (IgA-EF)

Natural Product:

Glycoprotein oligomer: Mr 50-60,000

Deduced Size of Mature Protein:

115 amino acids, Mr 13,000

Gene:

Chromosome: No information
 Organization: 4 exons, 3 introns (2.1 Kb)

Cellular Sources:

Activated T cells

Receptors:

No information

Known Actions:

Cell	Activity
B Cells	Induce the differentiation of activated B cells into antibody secreting cells Facilitate the production of IgA from activated B cells
T cells	Facilitate the generation of murine cytotoxic T cells
Homopoietic Precursors	Induce immature precursors to differentiate into eosinophils

Activity *in vivo*:

No information

Interleukin 6 (IL-6)

Other Names:

Interferon β_2 (IFN- β_2)
 B Cell Stimulatory Factor 2 (BSF-2)
 Hepatocyte Stimulatory Factor II (HSF-II)
 Hybridoma Plasmacytoma Growth Factor (HPGF, IL-HP1)
 Myeloma Cell Growth Factor (MCGF)
 26 Kd Protein

Natural Product:

Glycoprotein: Mr 26,000

Deduced Size of Mature Protein:

184 amino acids, Mr 21,000

Gene:

Chromosome: Single copy gene, chromosome 7

Organization: 5 exons, 4 introns

Cellular Sources:

Activated T cells	
Fibroblasts:	constitutive, increased by poly(I)poly(C), IL1, TNF α , IFN β_1 , PDGF
Monocytes:	constitutive, increased by bacterial endotoxin
EBV-transformed B lymphoblastoid cells:	constitutive
Myeloma cells:	constitutive
Atrial myxoma cells:	constitutive
Carcinoma cell lines:	constitutive, increased by TNF α , lymphotoxin

Receptors:

Specificity: IL6 only, human IL6 is active on murine cells

Avidity: $K_d = 3.4 \times 10^{-10} M$

Number: 140-890/cell (fresh myeloma cells); 2700/cell (CESS EBV-transformed B lymphoblastoid cells)

Size: No information

Organization: No information

Distribution: Resting T cells, activated B cells, plasma cells, myeloma cells, myeloid precursors, astrocytes, hepatocytes, fibroblasts

Known Actions:

Cell	Activity
B Cells	Stimulate terminal maturation of activated B cells or EBV transformed B cells into antibody secreting cells
	Promote growth of murine hybridomas and plasmacytomas
	Promote growth of myeloma cells
T Cells	Enhance production of IL2 from murine T cells
	Enhance differentiation of murine cytotoxic T cells
	Stimulate murine thymocyte proliferation
Hemopoietic Stem Cells	Stimulate growth of granulocyte-macrophage progenitors
	Stimulate growth of multipotential hemopoietic progenitors
Fibroblasts	Inhibit growth Enhance expression of class I MHC molecules
Hepatocytes	Induce production of haptoglobin, fibrinogen, α_1 antitrypsin

Activity *in vivo*:Animals

1. Induce fever

Man

1. High levels in blood and synovial fluid of patients with inflammatory arthritis

Lymphotoxin (LT)

Other Names:

Tumor necrosis factor β (TNF β)

Natural Product:

Glycoprotein: Mr 25,000

Glycoprotein trimer: Mr 60-70,000

Deduced Size of Mature Protein:

171 amino acids, Mr 18,600

Gene:

Chromosome: single copy gene, chromosome 6 (p23-q12)

Organization: No information

Cellular Sources:

Activated T cells

B lymphoblastoid cell lines

Receptors:

Specificity: LT or TNF α , human material is active in rodents

Avidity: $K_d = 2 \times 10^{-10} M$

Number: 200-2000/cell, increased by IFN- γ

Size: Mr 140,000

Organization: 2 polypeptides - Mr 80,000 and Mr 60,000

Distribution: Same as receptors for TNF α

Known Activities:

Presumably the same as TNF α

Activity *in vivo*:

Presumably the same as TNF α

Interferon Gamma (IFN γ)

Other Names:

Macrophage activating factor (MAF)

Natural Product:

Glycoprotein: Mr 20,000, Mr 25,000

Deduced Size of Mature Protein:

146 amino acids, Mr 17,100

Gene:

Chromosome:	single copy gene, chromosome 12
Organization:	4 exons, 3 introns

Cellular Sources:

Activated T cells
Natural killer cells

Receptors:

<u>Specificity:</u>	IFN γ only, species specific
<u>Avidity:</u>	$K_d = 5-10 \times 10^{-10} M$
<u>Number:</u>	2,000 - 10,000/cell
<u>Size:</u>	Mr 70,000 - 80,000
<u>Organization:</u>	Single protein
<u>Gene:</u>	Encoded by gene on chromosome 6 (long arm)
<u>Distribution:</u>	Monocyte/macrophages, fibroblasts, endothelial cells, T lymphocytes, B lymphoblasts

Known Actions

Cell	Activity
Macrophages	Activation, enhanced production of reactive oxygen species Increase expression of MHC molecules Increase production of IL1, TNF α
Fibroblasts	Inhibit collagen synthesis, induce expression of class II MHC molecules

	Inhibit heparin releasable lipoprotein lipase activity Increase resistance to intracellular pathogens
Adipocytes	Inhibit heparin releasable lipoprotein lipase activity
T Cells	Enhance differentiation of cytotoxic effectors Increase production of LT
Endothelial Cells	Induce expression of class II MHC antigens Increase resistance to intracellular pathogens
NK Cells	Augment killing
B Cells	Enhance generation of immunoglobulin secreting cells from activated murine B cells (IgM, IgG2a) Counteract the effects of IL4
Hemopoietic Stem Cells	Inhibit growth and differentiation
Tumor Cells	Increase expression of receptors for TNF α
Hepatocytes	Increase resistance to replication by exoerythrocytic stages of Plasmodium
Many Cells	Increase expression of class II MHC antigens

Activity *in vivo*:

Animals

1. Activate macrophages to produce reactive oxygen species and kill microorganisms
2. Protect from Listeria monocytogenes infection
3. Promote killing of intracellular pathogens - *Toxoplasma gondii*, *Leishmania donovani*, *Listeria monocytogenes*, *Mycobacterium intracellulare*
4. Accelerate fatal glomerulonephritis in (NZBxNZW)F₁ mice
5. Antibodies to IFN γ
 - a) cause remission in (NZBxNZW)F₁ mice
 - b) inhibit allograft rejection
 - c) prevent many of the toxic effects of bacterial endotoxin

Man

1. Local injection into lepromatous leprosy lesions —> T cell and

-
-
-
- 2. monocyte infiltration, increased HLA-DR antigen expression
- 2. Induce hypertryglyceridemia and inhibit lipoprotein
lipase activity
- 3. Ameliorate rheumatoid arthritis, psoriatic arthritis
- 4. Exacerbate multiple sclerosis

Colony Stimulating Factors

Interleukin 3 (IL3)

Other Names:

Multipotential Colony Stimulating Factor (Multi-CSF)
 Hematopoietic Cell Growth Factor (HCGF)
 P-Cell Stimulating Factor (PSF)
 Mast Cell Growth Factor (MCGF)
 Erythroid Colony-Stimulating Factor (ECSF)
 Megakaryocyte Colony Stimulating Factor (Meg-CSF)
 Eosinophil Colony Stimulating Factor (Eo-CSF)
 Burst-Promoting Activity (BPA)

Natural Product:

Glycoprotein, Mr 20-26,000

Deduced Size of Mature Protein:

133 amino acids, Mr 15,000

Gene:

Chromosome: Single copy gene, chromosome 5 (long arm)
 Organization: 5 exons, 4 introns

Cellular Source:

Activated T Cells

Receptors:

<u>Specificity:</u>	IL-3 only, species specific
<u>Avidity:</u>	* $K_d = 2-5 \times 10^{-11} M$
<u>Number:</u>	*1000-5000/cell
<u>Size:</u>	*Mr 135,000
<u>Organization:</u>	*Heterodimer (Mr 75,000 + Mr 60,000)
<u>Distribution:</u>	Hemopoietic precursors, neutrophils, eosinophils, macrophages, mast cells

* Murine IL3 receptor

Known Actions:

<u>Cell</u>	<u>Activity</u>
Hemopoietic Precursors	Stimulate proliferation and differentiation of precursors of all cell lineages
Mast Cells	Stimulate growth of mast cell lines
Myeloid Cells	Stimulate growth of myeloid cell lines

Activity *in vivo*:

No information

Granulocyte-Macrophage Colony Stimulating Factor (GM-CSF)Other Names:

Colony Stimulating Factor α (CSF α)
 Pluripoietin
 Colony Stimulating Factor 2 (CSF2)
 Neutrophil migration inhibitory factor from T cells (NIF-T)

Natural Product:

Glycoprotein: Mr 22,000

Deduced Size of Mature Protein:

127 amino acids, Mr 14,300

Gene:

Chromosome: Single copy gene, chromosome 5 (q21-q32)
 Organization: 4 exons, 3 introns (4Kb)

Cellular Sources:

Activated T cells
 HTLV-I infected lymphoblastoid cell lines
 Endothelial cells, fibroblasts stimulated by TNF α , IL1
 Myeloid leukemia cells
 Bladder carcinoma cells

Receptors:

Specificity: GM-CSF only, species specific

Avidity: $K_d = 1-3 \times 10^{-11} M$

Number: *70 high avidity receptors, 350 low avidity receptors per cell
Size: *Mr 50,000
Organization: Single protein
Distribution: Myeloid leukemia cells, mature neutrophils, monocytes, eosinophils

Known Actions:

Cell	Activity
Hemopoietic Stem Cells	Stimulate monocyte and neutrophil colonies Stimulate pure neutrophil colonies Stimulate eosinophil colonies Facilitate growth of erythroid progenitors (BFU-E)
Mature Neutrophils	Inhibit migration Prime for responses to other stimuli Enhance phagocytosis, antibody dependent cellular cytotoxicity Increase expression of the LFA-1 family of adhesion molecules
Eosinophils	Enhance leukotriene production Increase antibody-dependent cellular cytotoxicity
Monocytes	Enhance tumoricidal activity Enhance functional activity

Activity *in vivo*

Animals

Mice

1. Evidence of tissue macrophage expansion and activation in transgenic mice expressing a GM-CSF transgene.

Monkeys

1. Leukocytosis (neutrophils, eosinophils)
2. Priming of neutrophils
3. Enhance recovery of neutrophils, platelets after bone marrow transplantation

Man

1. Dose dependent increase in neutrophils, eosinophils in AIDS patients

2. Increase in neutrophils, monocytes, eosinophils and lymphocytes in patients with myelodysplastic syndromes

Macrophage Colony Stimulating Factor (M-CSF)

Other Names:

Colony stimulating factor-1 (CSF-1)
Urinary colony stimulating factor

Natural Product:

Glycoprotein dimer: Mr 70-90,000

Deduced Size of Mature Protein:

224 amino acids, Mr 26,000

Gene:

Chromosome: Single copy gene, chromosome 5 (long arm)
Organization: No information

Cellular Sources:

Fibroblasts:	Constitutive
Endothelial cells:	Constitutive
Monocytes:	Constitutive, increased by bacterial endotoxin, interferon- γ , phorbol esters

Receptors:

Specificity: M-CSF only, human M-CSF is active on mouse cells

Avidity: $*K_d = 4 \times 10^{-10} M$

Number: $*3,000-15,000/\text{cell}$

Size: Mr 165,000

Organization: Single protein with tyrosine kinase activity
Related (possibly identical) to the product of the
c-fms protooncogene encoded on the long arm of
chromosome 5

Distribution: Cells of monocyte/macrophage lineage
Choriocarcinoma cells

* Murine M-CSF

Known Actions:

<u>Cell</u>	<u>Activities</u>
Monocytes/Macrophages	Increase antibody dependent cellular cytotoxicity Increase PGE ₂ production Increase cytokine production (IL1, TNF) Increase tumoricidal activity
Hemopoietic Precursors	Stimulate growth of monocyte colonies

Activity *in vivo*:Animals

1. Leukocytosis
2. Prevent myelosuppression induced by interferons

Man

1. No information

Granulocyte-Colony Stimulating Factor (G-CSF)Other Names:

Colony stimulating factor β (CSF β)
Pluripotin

Natural Product:

Glycoprotein (O-linked sugar only): Mr 19,600

Deduced Size of Mature Protein:

177 amino acids, Mr 19,000

Gene:

Chromosome: Single copy gene, chromosome 17
Organization: 5 exons, 4 introns

Cellular Sources:

Fibroblasts, endothelial cells - after stimulation with IL1, TNF α
Monocytes - after stimulation by bacterial endotoxin, interferon- γ phorbol esters

Receptors:

Specificity: G-CSF only, human G-CSF is active on mouse cells

Avidity: * $K_d = 6-8 \times 10^{-11} M$

Number: *50-500/cell

Size: *Mr 150,000

Organization: *Single protein

Distribution: *Monomyelocytic cell lines
Mature neutrophils

* Murine G-CSF

Known Actions:

Cells	Activity
Hemopoietic Precursors	Induce differentiation of myelocytic cell lines Induce growth of pure neutrophil colonies
Mature neutrophils	Enhance antibody dependent cellular cytotoxicity Prime for enhanced response to other signals

Activity *in vivo*:

Animals

1. Increase circulating neutrophils, T cells and phagocytic activity of neutrophils in monkeys, mice

Man

No information

**ADDITIONAL CYTOKINES PRODUCED BY CELLS OF THE IMMUNE SYSTEM OR
ACTING ON CELLS OF THE IMMUNE SYSTEM**

1. Interferon α
2. Interferon β
3. Transforming Growth Factor β
4. Platelet Derived Growth Factor

ACTIVITIES IN SEARCH OF GENE PRODUCTS

1. Migration Inhibition Factor (MIF)
2. Soluble Immune Response Suppressor (SIRS)
3. Allogeneic Effect Factor (AEF)
4. Antigen Specific Helper Factor (T_H^F)
5. Antigen Specific Suppressor Factor (T_S^F)
6. Skin Reactive Factor (SRF)
7. Genetically-related Factor of Macrophages (GRF)
8. Specific Macrophage Arming Factor (SMAF)
9. Interleukin 4a
10. IgT
11. Proteolysis Inducing Factor (PIF)

SUMMARY OF THE CYTOKINES

1. Small protein or glycoprotein mediators produced by a wide variety of cells
 - a) Monocyte/macrophages - IL1, TNF α , IL6, GF-CSF, M-CSF, G-CSF
 - b) T lymphocytes - TNF α , IL2, IL4, IL5, IL6, LT, IFN γ , IL3, GM-CSF
 - c) B lymphocytes - IL2, IL6, LT
 - d) NK cells - IL1, IFN γ
 - e) Fibroblasts - IL6, GM-CSF, M-CSF, G-CSF
 - f) Endothelial cells - IL1, GM-CSF, M-CSF, G-CSF
 - g) Mast cells - IL4, TNF α
2. Active at low concentrations on a variety of cells expressing small numbers of high avidity specific receptors
 - a) Macrophage activation - IFN- γ , M-CSF, GM-CSF, IL-4, IL-2, IL-1, TNF α , LT
 - b) T lymphocyte growth and differentiation - IL-1, TNF α , IL-2, IL-4, IL-5, IL-6, LT
 - c) B lymphocyte growth and differentiation - IL-1, TNF α , IL-2, IL-4, IL-5, IL-6, LT, IFN γ
 - d) Natural killer cell function - IL-1, TNF α , IL-2, LT
 - e) Connective tissue - IL-1, TNF α , LT, IFN γ
 - f) Hepatocytes - IL-1, TNF α , LT, IL-6
 - g) Hemopoietic precursors - stimulation - IL-3, GM-CSF, G-CSF, M-CSF, IL-1, IL-4, IL-5, IL-6; inhibition - TNF α , IFN γ
3. Production of cytokines and responsiveness to cytokines are tightly regulated by environmental stimuli and other cytokines
 - a) Cytokine regulation of cytokine production

<u>Cytokine</u>	<u>Target Cell</u>	<u>Product</u>
IL1	Endothelial cell	GM-CSF, G-CSF
	Fibroblast	GM-CSF, G-CSF, IL6
	T cell	IL2, IFN γ
TNF α	Monocyte	IL1
	Endothelial cell	IL1, GM-CSF, G-CSF

	Fibroblast	GM-CSF, G-CSF, IL6
IL6	T cells	IL2
IFN γ	Monocyte T cell	IL1, TNF α LT
M-CSF	Monocyte	IL1, TNF α

b) Cytokine regulation of cytokine receptors

<u>Cytokine</u>	<u>Target Cell</u>	<u>Receptor</u>
IL1	T cells Hemopoietic stem cells	IL2 CSFs
TNF α	T cells	IL2
IL4	T cells	IL2
IFN γ	Monocytes Tumor cells	IL2 TNF α

4. Many manifestations of inflammation can be accounted for by the action of multiple cytokines

a) Acute phase response

1. IL1 - increase: complement factor B, C3, serum amyloid associated protein, α_1 -antichymotrypsin, α_1 acid glycoprotein, inter- α -trypsin inhibitor decrease: albumin, transferrin, fibrinogen
2. TNF α - increase: complement factor B, C3, α_1 antichymotrypsin decrease: albumin, transferrin
3. IL6 - increase: haptoglobin, fibrinogen, α_1 antitrypsin

b. Fever:

1. IL1
2. TNF α
3. IL6

c. Hypertriglyceridemia

1. TNF α
2. IL2
3. IFN γ

d. Neutrophilia

1. IL1
2. TNF α
3. GM-CSF
4. G-CSF

Interleukin 1

GENERAL

1. Dinarello, C.A. 1988. Biology of interleukin 1. FASEB J. 2:108-115.
2. Lumpkin, M.D. 1987. The regulation of ACTH secretion by IL-1. Science 238:452-454.
3. Mizel, S.B. 1987. Interleukin 1 and T-cell activation. Immunol. Today 8:330-332.
4. Dinarello, C.A. 1984. Interleukin-1. Rev. Infec. Dis. 6:51-95.
5. Dinarello, C.A. 1985. An update on human interleukin 1: From molecular biology to clinical relevance. J. Clin. Immunol. 5:287.
6. Dinarello, C.A., and Wolff, S.M. 1982. Molecular basis of fever in humans. Am. J. Med. 72:799-819.
7. Bernheim, H.A., Block, L.H., and Atkins, E. 1979. Fever: Pathogenesis, pathophysiology, and purpose. Ann. Int. Med. 91:261-270.
8. Kampschmidt, R.F. 1984. The numerous postulated biological manifestations of interleukin-1. J. Leukocyte Biol. 36:341-355.
9. Oppenheim, J.J., and Gery, I. 1982. Interleukin 1 is more than an interleukin. Immunol. Today 3:113, 1982.

GENE STRUCTURE

10. Lomedico, P.T., Gubler, U., Hellmann, C.P., Dukovich, M., Giri, J.G., Pan, Y-CE, Collier, K., Semionow, R., Chua, A.O., and Mizel, S.B. 1984. Cloning and expression of murine interleukin-1 cDNA in *Escherichia coli*. Nature 312:458-462.
11. Auron, P.E., Webb, A.C., Rosenwasser, L.J., Mucci, S.F., Rich, A., Wolff, S.M., and Dinarello, C.A. 1984. Nucleotide sequence of human monocyte interleukin 1 precursor cDNA. Proc. Natl. Acad. Sci. USA 81:7907-7911.
12. March, C.J., Mosley, B., Larsen, A., Cerretti, D.P., Braedt, G., Price, V., Gillis, S., Henney, C.S., Kronheim, S.R., Grabstein, K., Conlon, P.J., Hopp, T.P., and Cosman, D. 1985. Cloning, sequence and expression of two distinct human interleukin-1 complementary DNAs. Nature 315:641-647.
13. Gubler, U., Chua, A.O., Stern, A.S., Hellmann, C.P., Vitek, M.P., Dechiara, T.M., Benjamin, W.R., Collier, K.J., Dukovich, M., Familietti, P.C., Fiedler-Nagy, C., Jenson, J., Kaffka, K., Kilian, P.L., Stremlo, D., Wittreich, B.H., Woehle, D., Mizel, S.B., and Lomedico, P.T. 1986. Recombinant human interleukin 1 α : Purification and biological characterization. J. Immunol. 136:2492-2497.

14. Gray, P.W., Glaister, D., Chen, E., Goeddel, D.V., and Pennica, D. 1986. Two interleukin 1 genes in the mouse: cloning and expression of the cDNA for murine interleukin 1 β . *J. Immunol.* 137:3644-3648.
15. Furutani, Y., Notake, M., Fukui, T., Ohue, M., Nomura, H., Yamada, M., and Nakamura, S. 1986. Complete nucleotide sequence of the gene for human interleukin 1 alpha. *Nucleic Acids Res.* 14:3167-3179.
16. Clark, B.D., Collins, K.L., Gandy, M.S., Webb, A.C., and Auron, P.E. 1986. Genomic sequence for prointerleukin 1-beta: possible evolution from a reverse transcribed prointerleukin-1 alpha gene. *Nucleic Acids Res.* 14:7897-7914.

RECEPTOR

17. Dower, S.K., Call, S.M., Gillis, S., and Urdal, D.L. 1986. Similarity between the interleukin 1 receptors on a murine T-lymphoma cell line and on a murine fibroblast cell line. *Proc. Natl. Acad. Sci. USA* 83:1060-1064.
18. Dower, S.K., Kronheim, S.R., March, C.J., Conlon, P.J., Hopp, T.P., Gillis, S., and Urdal, D.L. 1985. Detection and characterization of high affinity plasma membrane receptors for human interleukin 1. *J. Exp. Med.* 162:501-515.
19. Dower, S.K., Kronheim, S.R., Hopp, T.P., Cantrell, M., Deeley, M., Gillis, S., Henney, C.S., and Urdal, D.L. 1986. The cell surface receptors for interleukin-1 α and interleukin-1 β are identical. 1986. *Nature* 324:266-268.
20. Matsushima, K., Akahoshi, T., Yamada, M., Furutani, Y., and Oppenheim, J.J. 1986. Properties of a specific interleukin 1 (IL 1) receptor on human Epstein Barr Virus-transformed B lymphocytes: identity of the receptor for IL 1- α and IL 1- β . *J. Immunol.* 136:4496-4502.
21. Lowenthal, J.W., and MacDonald, H.R. 1986. Binding and internalization of interleukin 1 by T cells: direct evidence for high- and low-affinity classes of interleukin 1 receptor. *J. Exp. Med.* 164:1060-1074.
22. Chin, J., Cameron, P.M., Rupp, E., and Schmidt, J.A. 1987. Identification of a high-affinity receptor for native human interleukin 1 β and interleukin 1 α on normal human lung fibroblasts. *J. Exp. Med.* 165:70-86.
23. Farrar, W.L., Kilian, P.L., Ruff, M.R., Hill, J.M., and Pert, C.B. 1987. Visualization and characterization of interleukin 1 receptors in brain. *J. Immunol.* 139:459-463.

PHYSIOLOGY

24. Gery, I., Gershon, R.K., and Waksmann, B.H. 1972. Potentiation of the T lymphocyte response to mitogens I. The responding cell. *J. Exp. Med.* 136:128-142.
25. Gery, I., and Waksmann, B. 1972. Potentiation of the T lymphocyte response to mitogens II. The cellular source of potentiating mediator(s). *J. Exp. Med.* 136:143-155.

26. Smith, K.A., Lachman, L.B., Oppenheim, J.J., and Favata, M.F. 1980. The functional relationship of the interleukins. *J. Exp. Med.* 151:1551-1556.
27. Kasahara, T., Mukaida, N., Hatake, K., Motoyoshi, K., Kawai, T., and Shiori-Nakano, K. 1985. Interleukin 1 (IL1) dependent lymphokine production by human leukemic T cell line HSB.2 subclones. *J. Immunol.* 134:1682-1689.
28. Farrar, J.J., Benjamin, W.R., and Hilfiker, M.L. 1982. The biochemistry, biology, and role of interleukin 1 in the induction of cytotoxic T cell and antibody-forming B cell responses. *Immunol. Rev.* 63:129-166.
29. Kaye, J., Gillis, S., Mizel, S.B., Shevach, E.M., Malek, T.R., Dinarello, C.A., Lachman, L.B., and Janeway, C.A., Jr. 1984. Growth of a cloned helper T cell line induced by monoclonal antibody specific for the antigen receptor; Interleukin 1 is required for the expression of receptors for Interleukin 2. *J. Immunol.* 133:1339-1345.
30. Mannel, D.N., Mizel, S.B., Diamantstein, T., and Falk, W. 1985. Induction of interleukin 2. *J. Immunol.* 134:3108-3110.
31. Davis, L., and Lipsky, P.E. 1986. Signals involved in T cell activation. II. Distinct roles of intact accessory cells, phorbol esters and interleukin 1 in activation and cell cycle progression of resting T lymphocytes. *J. Immunol.* 136:3588.
32. Hoffman, M.K., Koenig, S., Mittler, R.S., Oettgen, H.F., Ralph, P., Galanos, C., and Hammerling, U. 1979. Macrophage factor controlling differentiation of B cells. *J. Immunol.* 122:497-502.
33. Giri, J.G., Kincade, P.W., and Mizel, S.B. 1984. Interleukin 1-mediated induction of k-light chain synthesis and surface immunoglobulin expression of pre-B cells. *J. Immunol.* 132:223-228.
34. Leibson, H.J., Marrack, P., and Kappler, J. 1982. B cell helper factors II. Synergy among three helper factors in the response of T cell- and macrophage-depleted B cells. *J. Immunol.* 129:1398-1402.
35. Howard, M., Mizel, S.B., Lachman, L., Ansel, J., Johnson, B., Paul, W.E. 1983. Role of interleukin 1 in anti-immunoglobulin induced B cell proliferation. *J. Exp. Med.* 157:1529.
36. Lipsky, P.E. 1985. The role of interleukin 1 in human B cell activation. *Contemporary Topics in Molecular Immunology* 10:195-217.
37. Dempsey, R.A., Dinarello, C.A., Mier, Rosenwasser, L.J., Allegretta, M., Brown, T.E., and Parkinson, D.R. 1982. The differential effects of human leukocyte pyrogen/lymphocyte-activating factor, T cell growth factor, and interferon on human natural killer activity. *J. Immunol.* 129:2504-2510.
38. Dinarello, C.A., Marnow, S.O., and Rosenwasser, L.J. 1983. Role of arachidonate metabolism in the immunoregulatory function of human leukocytic pyrogen/LAF/IL 1. *J. Immunol.* 130:890-895.

39. Byars, N., Schreiber, A.B., and Allison, A.C. 1984. Interleukin 1-containing conditioned media stimulate the proliferation of capillary endothelial cells. *Fed. Proc.* 43:462-468.
40. Rossi, V., Breviario, F., Ghezzi, P., Dejana, E., and Mantovani, A. 1985. Interleukin-1 induces prostacyclin in vascular cells. *Science* 229:174-176, 1985.
41. Bevilacqua, M.P., Pober, J.S., Wheeler, M.E., Mendrick, D., Cotran, R.S., Gimbrone, M.A., Jr. 1985. Interleukin 1- (IL1) acts on vascular endothelial cells to increase their adhesivity for blood leukocytes. *Fed. Proc.* 44:1494-1499, 1985.
42. Nawroth, P.P., Handley, D.A., Esmon, C.T., and Stern, D.M. 1986. Interleukin 1 induces endothelial cell procoagulant while suppressing cell-surface anticoagulant activity. *Proc. Natl. Acad. Sci. USA* 83:3460-3464, 1986.
43. Bevilacqua, M.P., Pober, J.S., Majeau, G.R., Cotran, R.S., and Gimbrone, J.A., Jr. 1984. Interleukin-1 induces biosynthesis and cell surface expression of procoagulant activity on human vascular endothelial cells. *J. Exp. Med.* 160:618-623.
44. Pober, J.S., Bevilacqua, M.P., Mendrick, D.L., Lapierre, L.A., Fiers, W., and Gimbrone, M.A., Jr. 1986. Two distinct monokines, interleukin 1 and tumor necrosis factor, each independently induce biosynthesis and transient expression of the same antigen on the surface of cultured human vascular endothelial cells. *J. Immunol.* 136:1680-1687.
45. Postlethwaite, A.E., Lachman, L.B., Mainardi, C., and Kang, A.H. 1983. Interleukin 1 stimulation of collagenase production by cultured fibroblasts. *J. Exp. Med.* 157:801-806.
46. Dayer, J-M, Breard, J., Chessl, L., and Krane, S.M. 1979. Participation of monocyte-macrophages and lymphocytes in the production of a factor that stimulates collagenase and prostaglandin release by rheumatoid synovial cells. *J. Clin. Inest.* 64:1386-1392.
47. Schmidt, J.A., Mizel, S.B., Cohen, D., and Green, I. 1982. Interleukin 1, a potential regulator of fibroblast proliferation. *J. Immunol.* 128:2177-2182, 1982.
48. Schmidt, J.A., Oliver, C.N., Lepe-Zuniga, T.L., Green, I., and Gery, I. 1984. Silica stimulated monocytes release fibroblast proliferation factors identical to IL 1. A potential role for IL 1 in the pathogenesis of silicosis. *J. Clin. Invest.* 73:1462-1472.
49. VanDamme, J., DeLey, M., Opdenakker, G., Billiau, A., DeSomer, P., and Van Beeumen, J. 1985. Homogeneous interferon-inducing 22K factor is related to endogenous pyrogen and interleukin-1. *Nature* 314:266-268.
50. Saklatvala, J., Pilsworth, L.M.C., and Sarsfield, S.J. 1984. Pig catabolin is a form of interleukin 1; Cartilage and bone resorb, fibroblasts make prostaglandin and collagenase, and thymocyte proliferation is augmented in response to one protein. *Biochem. J.* 244:461-466.

51. Gowen, M., Wood, D.D., and Ihrie, E.J. 1984. Stimulation by human interleukin 1 of cartilage breakdown and production of collagenase and proteoglycanase by human chondrocytes but not by human osteoblasts *in vitro*. *Biophys. Acta.* 797:186-193.
52. Richardson, H.J., Elford, P.R., and Sharrard, R.M. 1985. Modulation of connective tissue metabolism by partially purified human IL 1. *Cell. Immunol.* 90:41-51.
53. Wood, D.D., Bayne, E.K., Goldring, M.B., Gowen, M., Hamerman, D., Humes, J.L., Ihrie E.J., Lipsky, P.E., and Staruch, M.J.. 1985. The four biochemically distinct species of human interleukin 1 all exhibit similar biologic activities. *J. Immunol.* 134:895-903.
54. Lovett, D.H., Ryan, J.L., and Sterzel, R.B. 1983. Stimulation of rat mesangial cell proliferation by macrophage interleukin 1. *J. Immunol.* 131:2830-2836.
55. Gowen, M., Wood, D.D., and Ihrie, E.J. 1983. An IL 1 like factor stimulates bone resorption *in vitro*. *Nature (London)* 306:378-380.
56. Rifas, L., Shen, F., and Mitchell, K. 1984. Macrophage derived growth for osteoblast-like cells and chondrocytes. *Proc. Natl. Acad. Sci. USA* 81:4558-4562.
57. Matsushima, K., Bano, M., Kidwell, W.R., and Oppenheim, J.J. 1985. Interleukin 1 increases collagen type IV production by murine mammary epithelial cells. *J. Immunol* 134:904-909.
58. Beutler, B.A., and Cerami, A. 1985. Recombinant interleukin 1 suppresses lipoprotein lipase activity in 3T3-L1 cells. *J. Immunol.* 135:3969-3971.
59. Rupp, E.A., Cameron, P.M., Ranawat, C.S., Schmidt, J.A., and Bayne, E.K. 1986. Specific bioactivities of monocyte-derived interleukin 1 α and interleukin 1 β are similar to each other on cultured murine thymocytes and on cultured human connective tissue cells. *J. Clin. Invest.* 78:836-839.
60. Perlmutter, D.H., Goldberger, G., Dinarello, C.A., Mizel, S.B., and Colten, H.R. 1986. Regulation of class III major histocompatibility complex gene products by interleukin-1. *Science* 232:850-852.
61. Mochizuki, D.Y., Eisenman, J.R., Conlon, P.J., Larsen, A.D., and Tushinski, R.J. 1987. Interleukin 1 regulates hematopoietic activity, a role previously ascribed to hemopoietin 1. *Proc. Natl. Acad. Sci. USA* 84:5267-5271.
62. Zucali, J.R., Effenbein, G.J., Barth, K.C., and Dinarello, C.A. 1987. Effects of human interleukin 1 and human tumor necrosis factor on human T lymphocyte colony formation. *J. Clin. Invest.* 80:772-777.
63. Onozaki, K., Matsushima, K., Aggarwal, B.B., and Oppenheim, J.J. 1985. Human interleukin 1 is a cytoidal factor for several tumor cell lines. *J. Immunol.* 135:3962-3968.
64. Lovett, D., Kozan, B., Hadam, M., Resch, K. and Gemsa, D. 1986. Macrophage cytotoxicity: Interleukin 1 as a mediator of tumor cytostasis. *J. Immunol.* 136:340-347.

65. Bendtzen, K., Mandrup-Poulsen, T., Nerup, J., Nielsen, J.H., Dinarello, C.A., and Svenson, M. 1986. Cytotoxicity of human pI 7 interleukin 1 for pancreatic islets of Langerhans. *Science* 232:1545-1547.
66. Libby, P., Warner, S.J.C., and Friedman, G.B. 1988. Interleukin 1: a mitogen for human vascular smooth muscle cells that induces the release of growth-inhibitory prostanoids. *J. Clin. Invest.* 81:487-498.
67. Zucali, J.R., Dinarello, C.A., Oblon, D.J., Gross, M.A., Anderson, L., and Weiner, R.S. 1986. Interleukin 1 stimulates fibroblasts to produce granulocyte-macrophage colony-stimulating activity and prostaglandin E₂. *J. Clin. Invest.* 77:1857-1863.
68. Bagby, G.C., Jr., Dinarello, C.A., Wallace, P., Wagner, C., Hefeneider, S., and McCall, E. 1986. Interleukin 1 stimulates granulocyte macrophage colony-stimulating activity release by vascular endothelial cells. *J. Clin. Invest.* 78:1316-1323.
69. Kaushansky, K., Lin, N., and Adamson, J.W. 1988. Interleukin 1 stimulates fibroblasts to synthesize granulocyte-macrophage and granulocyte colony-stimulating factors. Mechanism for the hematopoietic response to inflammation. *J. Clin. Invest.* 81:92-97.
70. Sieff, C.A., Tsai, S., and Faller, D.V. 1987. Interleukin 1 induces cultured human endothelial cell production of granulocyte-macrophage colony-stimulating factor. *J. Clin. Invest.* 79:48-51.
71. Lindholm, D., Heumann, R., Meyer, M., and Thoenen, H. 1987. Interleukin-1 regulates synthesis of nerve growth factor in non-neuronal cells of rat sciatic nerve. *Nature* 330:658-659.
72. Bernton, E.W., Beach, J.E., Holaday, J.W., Smallridge, R.C., and Fein, H.G. 1987. Release of multiple hormones by a direct action of interleukin-1 on pituitary cells. *Science* 238:519-521.
73. VanDamme, J., Cayphas, S., Opdenakker, G., Billiau, A., and Van Snick, J. 1987. Interleukin 1 and poly(rI)*poly(rC) induce production of a hybridoma growth factor by human fibroblasts. *Eur. J. Immunol.* 17:1-7.
74. Perlmutter, D.H., Dinarello, C.A., Punyal, P.I., and Colten, H.R. 1986. Cachectin/tumor necrosis factor regulates hepatic acute-phase gene expression. *J. Clin. Invest.* 78:1349-1354.

IN VIVO ACTIVITIES

75. Czuprynski, C.J., Brown, J.F., Young, K.M., Cooley, A.J. and Kurtz, R.S. 1988. Effects of murine recombinant interleukin 1 α on the host response to bacterial infection. *J. Immunol.* 140:962-968.
76. VanDerMeer, J.W.M., Barza, J., Wolff, S.M., and Dinarello, C.A. 1988. A low dose of recombinant interleukin 1 protects granulocytopenic mice from lethal gram-negative infection. *Proc. Natl. Acad. Sci. USA* 85:1620-1623.

77. Neta, R., Douches, S., and Oppenheim, J.J. 1986. Interleukin 1 is a radioprotector. *J. Immunol.* 136:2483-2485.
78. White, C.W., Ghezzi, P., Dinarello, C.A., Caldwell, S.A., McMurtry, I.F., and Repine, J.E. 1987. Recombinant tumor necrosis factor/cachectin and interleukin 1 pretreatment decreases lung oxidized glutathione accumulation, lung injury and mortality in rats exposed to hyperoxia. *J. Clin. Invest.* 79:1868-1873.
79. Hill, M.R., Stith, R.D., and McCallum, R.E. 1986. Interleukin 1: A regulatory role in glucocorticoid-regulated hepatic metabolism. *J. Immunol.* 137:858-862.
80. Sapolsky, R., Rivier, C., Yamamoto, G., Plotsky, P., and Vale, W. 1987. Interleukin-1 stimulates the secretion of hypothalamic corticotropin-releasing factor. *Science* 238:522-524.
81. Berkenbosch, F., Van Oers, J., Del Rey, A., Tilders, F., and Besedovsky, H. 1987. Corticotropin-releasing factor-producing neurons in the rat activated by interleukin-1. *Science* 238:524-526.
82. Dingle, J.T., Page Thomas, D.P., King, B., and Bard, D.R. 1987. *In vivo* studies of articular tissue damage mediated by catabolin/interleukin 1. *Ann. Rheum. Dis.* 46:527-533.
83. Pettipher, E.R., Higgs, G.A., and Henderson, B. 1986. Interleukin 1 induces leukocyte infiltration and cartilage proteoglycan degradation in the synovial joint. *Proc. Natl. Acad. Sci. USA* 83:8749-8753.
84. Fontana, A., Hengartner, H., Weber, E., Fehr, K., Grob, P.J., and Cohen, G. 1982. Interleukin 1 activity in the synovial fluid of patients with rheumatoid arthritis. *Arthritis Rheum.* 2:49-53.
85. Wood, D.D., Ihrie, E.J., Dinarello, C.A., and Cohen, P.L. 1983. Isolation of an interleukin-1-like factor from human joint effusions. *Arthritis Rheum.* 26:975-983.
86. Oppenheim, J.J., Charon, J.A., and Luger, T.A. 1982. Evidence for an *in vivo* inflammatory role of interleukin 1 (IL1). *Transplant Proc.* 14:553-555.
87. Vogel, S.N., Douches, S.D., Kaufman, E.N., and Neta, R. 1987. Induction of colony stimulating factor *in vivo* by recombinant interleukin 1 α and recombinant tumor necrosis factor α . *J. Immunol.* 138:2143-2148.

TNF α GENERAL

1. Beutler, B., and Cerami, A. 1987. Cachectin: More than a tumor necrosis factor. *N. Engl. J. Med.* 316:379-385.
2. Le, J., and Vilcek, J. 1987. Tumor necrosis factor and interleukin 1: Cytokines with multiple overlapping biological activities. *Lab. Invest.* 56:234-248.
3. Cerami, A., and Beutler, B. 1988. The role of cachectin/TNF in endotoxic shock and cachexia. *Immunol. Today* 8:28-31.

GENE STRUCTURE

4. Shirai, T., Yamaguchi, H., Ito, H., Todd, C.W., and Wallace, R.B. 1985. Cloning and expression in *Escherichia coli* of the gene for human tumour necrosis factor. *Nature* 313:803-806.
5. Pennica, D., Nedwin, G.E., Hayflick, J.S., Seburg, P.H., Derynck, R., Palladino, M.A., Kohr, W.J., Aggarwal, B.B., and Goeddel, D.V. 1984. Human tumor necrosis factor: precursor structure, expression, and homology to lymphotoxin. *Nature* 312:724-729.
6. Spies, T., Morton, C.C., Nedospasov, S.A., Fiers, W., Pious, D., and Strominger, J.L. 1986. Genes for the tumor necrosis factors α and β are linked to the human major histocompatibility complex. *Proc. Natl. Acad. Sci. USA* 83:8699-8702.

RECEPTOR

7. Rubin, B.Y., Anderson, S.L., Sullivan, S.A., Williamson, B.D., Carswell, E.A., and Old, L.J. 1985. High affinity binding of 125 I-labeled human tumor necrosis factor (LuKII) to specific cell surface receptors. *J. Exp. Med.* 162:1099-1104.
8. Aggarwal, B.B., Eessalu, T.E., and Hass, P.E. 1985. Characterization of receptors for human tumour necrosis factor and their regulation by gamma-interferon. *Nature* 318:665-667.
9. Tsujimoto, M., Kip, Y.K., and Vilcek, J. 1985. Tumor necrosis factor: specific binding and internalization in sensitive and resistant cells. *Proc. Natl. Acad. Sci. USA* 82:7626-7630.
10. Kehrl, J.H., Miller, A., and Fauci, A.S. 1987. Effect of tumor necrosis factor α on mitogen activated human B cells. *J. Exp. Med.* 166:786-791.

PHYSIOLOGY

11. Perlmutter, D.H., Dinarello, C.A., Punyal, P.I., and Colten, H.R. 1986. Cachectin/tumor necrosis factor regulates hepatic acute-phase gene expression. *J. Clin. Invest.* 78:1349-1354.

12. Cuturi, M.C., Murphy, M., Costa-Giom, M.P., Weinmann, R., Perussia, B., and Trinchieri, G. 1987. Independent regulation of tumor necrosis factor and lymphotoxin production by human peripheral blood lymphocytes. *J. Exp. Med.* 165:1581-1594.
13. Ming, W.J., Bersani, L., and Mantovani, A. 1987. Tumor necrosis factor is chemotactic for monocytes and polymorphonuclear leukocytes. *J. Immunol.* 138:1469-1474.
14. Le, J., Weinstein, D., Gubler, U., and Vilcek, J. 1987. Induction of membrane-associated interleukin 1 by tumor necrosis factor in human fibroblasts. *J. Immunol.* 138:2137-2142.
15. Leeuwenberg, J.F.M., VanDamme, J., Jeunhomme, G.M.A.A., and Buurman, W.A. 1987. Interferon β 1, an intermediate in the tumor necrosis factor α -induced increased MHC class I expression and an autocrine regulator of the constitutive MHC class I expression. *J. Exp. Med.* 166:1180-1185.
16. Kohase, M., Henriksen-DeStefano, D., May, L., Vilcek, J., and Sehgal, P. 1986. Induction of β 2-interferon by tumor necrosis factor: a homeostatic mechanism in the control of cell proliferation. *Cell* 45:659-666.
17. Camussi, G., Bussolino, F., Salvidio, G., and Baglioni, C. 1987. Tumor necrosis factor/cachectin stimulates peritoneal macrophages, polymorphonuclear neutrophils, and vascular endothelial cells to synthesize and release platelet-activating factor. *J. Exp. Med.* 166:1390-1404.
18. Munker, R., Gasson, J., Ogawa, M., and Koeffler, H.P. 1986. Recombinant human TNF induces production of granulocyte-monocyte colony-stimulating factor. *Nature* 323:79-82.
19. Aggarwal, B.B., Kohr, W.J., Hass, P.E., Moffat, B., Spencer, S.A., Henzel, W.J., Bringman, T.S., Nedwin, G.E., Goeddel, D.V., and Harkins, R.N. 1985. Human tumor necrosis factor. Production, purification, and characterization. *J. Biol. Chem.* 260:2345-2354.
20. Beutler, B.A., Milsark, I.W., and Cerami, A. 1985. Cachectin/tumor necrosis factor: production, distribution, and metabolic fate *in vivo*. *J. Immunol.* 135:3972-3977.
21. Wong, G.H.W., and Goeddel, D.V. 1986. Tumour necrosis factors α and β inhibit virus replication and synergize with interferons. *Nature* 323:819-822.
22. Dayer, J-M., Beutler, B., and Cerami, A. 1985. Cachectin/tumor necrosis factor stimulates collagenase and prostaglandin E₂ production by human synovial cells and dermal fibroblasts. *J. Exp. Med.* 162:2163-2168.
23. Bertolini, D.R., Nedwin, G., Bringman, T., Smith, D., and Mundy, G.R. 1986. Stimulation of bone resorption and inhibition of bone formation *in vitro* by human tumor necrosis factors. *Nature* 319:516-519.

24. Stern, D.M., and Nowroth, P.P. 1986. Modulation of endothelial cell hemostatic properties by tumor necrosis factor. *J. Exp. Med.* 163:740-748.
25. Gamble, J.R., Harlan, J.M., Klebanoff, S.J., and Vadas, M.A. 1985. Stimulation of the adherence of neutrophils to umbilical vein endothelium by human recombinant tumor necrosis factor. *Proc. Natl. Acad. Sci. USA* 82:8667-8671.
26. Klebanoff, S.J., Vadas, M.A., Harlan, J.M., Sparks, L.H., Gamble, J.R., Agosti, J.M., and Waltersdorph, A.M. 1986. Stimulation of neutrophils by tumor necrosis factor. *J. Immunol.* 136:4220-4225.
27. Beutler, B., and Cerami, A. 1986. Cachectin and tumor necrosis factor as two sides of the same biological coin. *Nature* 320:584-588.
28. Pober, J.S., Bevilacqua, M.P., Mendrick, D.L., Lapierre, L.A., Fiers, W., and Gimbrone, M.A., Jr. 1986. Two distinct monokines, interkeukin 1 and tumor necrosis factor, each independently induce biosynthesis and transient expression of the same antigen on the surface of cultured human vascular endothelial cells. *J. Immunol.* 136:1680-1687.
29. Ostensen, M.E., Thiele, D.L., and Lipsky, P.E. 1987. Tumor necrosis factor α enhances cytolytic activity of human natural killer cells. *J. Immunol.* 138:4185-4191.
30. Jelinek, D.F., and Lipsky, P.E. 1987. Enhancement of human B cell proliferation and differentiation by tumor necrosis factor- α and interleukin 1. *J. Immunol.* 139:2970-2976.
31. Yokota, S., Geppert, T.D., and Lipsky, P.E. 1988. Enhancement of antigen- and mitogen-induced human T lymphocyte proliferation by tumor necrosis factor- α . *J. Immunol.* 140:531-536.
32. Hackett, R.J., Davis, L.S., and Lipsky, P.E. 1988. Comparative effects of tumor necrosis factor α and IL1 β on mitogen-induced T cell activation. *J. Immunol.*, in press.

IN VIVO ACTIVITIES

33. Mahmood, T., Busch, H.M., Racis, S.P., and Krey, P.R. 1987. Lymphokines in inflammatory arthritis. *Arthritis Rheum.* 30:S83, 1987.
34. White, C.W., Ghezzi, P., Dinarello, C.A., Caldwell, S.A., McMurtry, I.F., and Repine, J.E. 1987. Recombinant tumor necrosis factor/cachectin and interleukin 1 pretreatment decreases lung oxidized glutathione accumulation, lung injury and mortality in rats exposed to hyperoxia. *J. Clin. Invest.* 79:1868-1873.
35. Vogel, S.N., Douches, S.D., Kaufman, E.N., and Neta, R. 1987. Induction of colony stimulating factor *in vivo* by recombinant interleukin 1a and recombinant tumor necrosis factor a. *J. Immunol.* 138:2143-2148.
36. Dinarello, C.A., Cannon, J.G., Wolff, S.M., Bernheim, H.A., Beutler, B., Cerami, A., Figari, I.S., Palladino, M.A., Jr., and O'Connor, J.V. 1986. Tumor necrosis factor

- (cachectin) is an endogenous pyrogen and induces production of interleukin 1. J. Exp. Med. 163:1433-1450.
37. Kettelhut, I.C., Fiers, W., and Goldberg, A.L. 1987. The toxic effects of tumor necrosis factor *in vivo* and their prevention by cyclooxygenase inhibitors. Proc. Natl. Acad. Sci. USA 84:4273-4277.
 38. Grau, G.E., Fajardo, L.F., Piguet, P-F, Allet, B., Lambert, P-H, and Vassali, P. 1987. Tumor necrosis factor (Cachectin) as an essential mediator in murine cerebral malaria. Science 237:1210-1212.
 39. Piguet, P-F., Grau, G.E., Allet, B., and Vassalli, P. 1987. Tumor necrosis factor/cachectin is an effector of skin and gut lesions of the acute phase of graft-vs-host-disease. J. Exp. Med. 166:1280-1289.
 40. Tracey, K.J., Fong, Y., Hesse, D.G., Manogue, K.R., Lee, A.T., Kuo, G.C., Lowry, S.F., and Cerami, A. 1987. Anti-cachectin/TNF monoclonal antibodies prevent septic shock during lethal bacteraemia. 1987. Nature 330:662-664.
 41. Jacob, C.O., and McDevitt, H.O. 1988. Tumour necrosis factor- α in murine autoimmune 'lupus' nephritis. Nature 331:356-358.

Interleukin 2 (IL2)

GENERAL

1. Smith, K.A. 1980. T-cell growth factor. *Immunol. Rev.* 51:337-357.
2. Robb, R.J. 1984. Interleukin 2: The molecule and its function. *Immunol. Today* 5:203-209.
3. Farrar, J.J., Benjamin, W.R., Hilfiker, M.L., Howard, M., Farrar, W.L., and Fuller-Farrar, J. 1982. The biochemistry, biology, and role of interleukin 2 in the induction of cytotoxic T cell and antibody forming B cell responses. *Immunol. Rev.* 63:129-166.

GENE STRUCTURE

4. Devos, R., Plaetinck, G., Cheroutre, H., Simons, G., Degrave, W., Tavernier, J., Remaut, E., and Fiero, W. 1983. Molecular cloning of human interleukin 2 cDNA and its expression in *E. coli*. *Nucleic Acids Res.* 11:4307-4323.
5. Taniguchi, T., Matsui, H., Fujita, T., Takaoka, C., Kashima, N., Yoshimoto, R., and Hamuro, J. 1983. Structure and expression of a cloned cDNA for human interleukin-2. *Nature* 302:305-310.
6. Fujita, T., Takaoka, C., Matsui, H., and Taniguchi, T. 1983. Structure of the human interleukin 2 gene. *Proc. Natl. Acad. Sci. USA* 80:7437-7441.
7. Holbrook, N.J., Smith, K.A., Fornace, A.J., Jr., Comeau, C.H., Wiskocil, R.L., and Crabtree, G.R. 1984. T-cell growth factor: Complete nucleotide sequence and organization of the gene in normal and malignant cells. *Proc. Natl. Acad. Sci. USA* 81:1634-1638.
8. Siegel, L.J., Harper, M.E., Wong-Staal, F., Gallo, R.C., Nash, W.G., and O'Brien, S.J. 1984. Gene for T-cell growth factor: Location on human chromosome 4q and feline chromosome B1. *Science* 223:175-178.
9. Fujita, T., Shibuya, H., Ohashi, T., Yamanishi, K., and Taniguchi, T. 1986. Regulation of human interleukin-2 gene: Functional DNA sequences in the 5' flanking region for the gene expression in activated T lymphocytes. *Cell* 46:401-405.

RECEPTOR

10. Tsudo, M., Uchiyama, T., and Uchino, H. 1984. Expression of Tac antigen on activated normal human B-cells. *J. Exp. Med.* 160:612-617.
11. Herrmann, F., Cannistra, S.A., Levine, H., and Griffin, J.D. 1985. Expression of interleukin 2 receptors and binding of interleukin 2 by gamma interferon-induced human leukemic and normal monocytic cells. *J. Exp. Med.* 162:1111-1116.

12. Birchenall-Sparks, M.C., Farrar, W.L., Rennick, D., Kilian, P.L., and Ruscetti, F.W. 1986. Regulation of expression of the interleukin-2 receptor on hematopoietic cells by interleukin-3. *Science* 233:455-458.
13. Treiger, B.F., Leonard, W.J., Svetlik, P., Rubin, L., Nelson, D.L., and Greene, W.C. 1986. A secreted form of the human interleukin-2 receptor encoded by an "anchor minus" cDNA. *J. Immunol.* 136:4099-4105.
14. Bich-Thuy, L.T., Dukovich, M., Peffer, N.J., Fauci, A.S., Kehrl, J.H., and Greene, W.C. 1987. Direct activation of human resting T cells by IL 2: The role of an IL 2 receptor distinct from the Tac protein. *J. Immunol.* 139:1550-1556.
15. Kehrl, J.H., Dukovich, M., Whalen, G., Katz, P., Fauci, A.S., and Greene, W.C. 1988. Novel interleukin 2 (IL-2) receptor appears to mediate IL-2-induced activation of natural killer cells. *J. Clin. Invest.* 81:200-205.
16. Tsudo, M., Kozak, R.W., Goldman, C.K., and Waldmann, T.A. 1986. Demonstration of a non-Tac peptide that binds interleukin-2: A potential participant in a multichain interleukin-2 receptor complex. *Proc. Natl. Acad. Sci. USA* 83:9694-9698.
17. Robb, R.J., Munck, A., and Smith, K.A. 1981. T-cell growth factor receptors: Quantification, specificity, and biological relevance. *J. Exp. Med.* 154:1455-1474.
18. Leonard, W.J., Depper, J.M., Uchiyama, T., Smith, K.A., Waldmann, T.A., and Greene, W.C. 1982. A monoclonal antibody that appears to recognize the receptor for human T cell growth factor: Partial characterization of the receptor. *Nature* 300:267-269.
19. Robb, R.J., and Greene, W.C. 1983. Direct demonstration of the identity of T cell growth factor binding protein and the Tac antigen. *J. Exp. Med.* 158:1332-1337.
20. Leonard, W.J., Depper, J.M., Robb, R.J., Waldmann, T.A., and Green, W.C. 1983. Characterization of the human receptor for T cell growth factor. *Proc. Natl. Acad. Sci. USA* 80:6957-6961.
21. Leonard, W.J., Depper, J.M., Kronke, M., Robb, R.J., Waldmann, T.A., and Greene, W.C. 1985. The human receptor for T-cell growth factor: Evidence for variable post-translational processing phosphorylation, sulfation, and the ability of precursor forms of the receptor to bind TCGF. *J. Biol. Chem.* 260:1872-1880.
22. Urdal, D.L., March, C.J., Gillis, S., Larsen, A., and Dower, S.K. 1984. Purification and chemical characterization of the receptor for interleukin-2 from activated human T lymphocytes and from a human T-cell lymphoma cell-line. *Proc. Natl. Acad. Sci. USA* 81:6481-6485.
23. Leonard, W.J., Depper, J.M., Crabtree, G.R., Rudikoff, S., Pumphrey, J., Robb, R.J., Kronke, M., Svetlik, P.B., Peffer, N.J., Waldmann, T.A., and Greene, W.C. 1984. Molecular cloning and expression of cDNAs for the human interleukin-2 receptor. *Nature* 311:626-631.

24. Nikaido, T., Shimizu, N., Ishida, N., Sabe, H., Teshigawara, K., Maeda, M., Uchiyama, T., Yodoi, J., and Honjo, T. 1984. Molecular cloning of cDNA encoding human interleukin-2 receptor. *Nature* 311:631-635.
25. Cosman, D., Cerretti, D.P., Larsen, A., Park, L., March, C., Dower, S., Gillis, S., and Urdal, D. 1984. Cloning, sequence and expression of human interleukin-2 receptor. *Nature* 312:768-771.
26. Leonard, W.J., Depper, J.M., Kronke, M., Peffer, N.J., Svetlik, P.B., Kanehisa, M., Sullivan, M., and Greene, W.C. 1985. Structure of the human interleukin-2 receptor gene. *Science* 230:633-639.
27. Ishida, N., Kanamori, H., Noma, T., Kikaido, T., Sabe, H., Suzuki, A., Shimizu, A., and Honjo, T. 1985. Molecular cloning and structure of the human interleukin-2 gene. *Nucleic Acids Res.* 13:7579-7589.
28. Leonard, W.J., Donlon, T.A., Lebo, R.V., and Greene, W.C. 1985. Localization of the gene encoding the human interleukin-2 receptor on chromosome 10. *Science* 228:1547-1549.
29. Hemler, M.D., Brenner, M.B., McLean, J.M., and Strominger, J.L. 1984. Antigen stimulation regulates the level of expression of IL-2 receptors on human T-cells. *Proc. Natl. Acad. Sci. USA* 81:2172-2175.
30. Robb, R.J., Greene, W.C., and Rusk, C.M. 1984. Low and high affinity cellular receptors for interleukin 2. Implications for the level of Tac antigen. *J. Exp. Med.* 160:1126-1146.
31. Lowenthal, J.W., Zubler, R.H., Nabholz, M., and MacDonald, H.W. 1985. Similarities between interleukin-2 receptor number and affinity on activated B and T lymphocytes. *Nature* 315:669-672.
32. Weissman, A.M., Harford, J.B., Svetlik, P.B., Leonard, W.J., Depper, J.M., Waldmann, T.A. Greene, W.C., and Klausner, R.D. 1986. Only high affinity receptors for interleukin-2 mediate internalization of ligand. *Proc. Natl. Acad. Sci. USA* 83:1463-1466.
33. Robb, R.J. 1986. Conversion of low-affinity interleukin 2 receptors to a high-affinity state following fusion of cell membranes. *Proc. Natl. Acad. Sci. USA* 83:3992-3996.
34. Sharon, M., Klausner, R.D., Cullen, B.R., Chizzonite, R., and Leonard, W.J. 1986. Novel interleukin-2 receptor subunit detected by cross-linking under high affinity conditions. *Science* 234:859-863.
35. Teshigaward, K., Wang, H-M., Kato, K., and Smith, K.A. 1987. Interleukin 2 high-affinity receptor expression requires two distinct binding proteins. *J. Exp. Med.* 165:223-238.
36. Robb, R.J., Rusk, C.M., Yodoi, J., and Greene, W.C. 1987. Interleukin 2 binding molecule distinct from the Tac protein: Analysis of its role in formation of high-affinity receptors. *Proc. Natl. Acad. Sci. USA* 84:2002-2006.

37. Dukovich, M., Wano, Y., Bich Thuy, LT, Katz, P., Cullen, B.R., Kehrl, J.H., and Greene, W.C. 1987. A second human interleukin-2 binding protein that may be a component of high-affinity interleukin-2 receptors. *Nature* 327:518-522.
38. Siegel, J.P., Sharon, M., Smith, P.L., and Leonard, W.J. 1987. The IL-2 receptor β chain (p70): Role in mediating signals for LAK, NK, and proliferative activities. *Science* 238:75-78.
39. Rubin, L.A., Hoekzema, G.S., Nelson, D.L., Greene, W.C., and Jay, G. 1987. Reconstitution of a functional interleukin 2 receptor in a nonlymphoid cell. *J. Immunol.* 139:2355-2360.
40. Tanaka, T., Saiki, O., Doi, S., Negoro, S., and Kishimoto, S. 1988. Interleukin 2 functions through novel interleukin 2 binding molecules in T cells. *J. Immunol.* 140:470-473.
41. Tanaka, T., Saiki, O., Doi, S., Fuji, M., Sugamura, K., Hara, H., Negoro, S., and Kishimoto, S. 1988. Novel receptor-mediated internalization of interleukin 2 in B cells. *J. Immunol.* 140:866-870.
42. Saiki, O., Tanaka, T., Doi, S., and Kishimoto, S. 1988. Expression and the functional role of a p70/75 interleukin 2-binding molecule in human B cell. *J. Immunol.* 140:853-858.

PHYSIOLOGY

43. Brandhuber, B.J., Boone, T., Kenney, W.C., and McKay, D.B. 1987. Three-dimensional structure of interleukin-2. *Science* 238:1707-1709.
44. Welte, K., Wang, C.Y., Mertelsman, R., Venuta, S., Feldman, S.P., and Moore, M.A.S. 1982. Purification of human interleukin 2 to apparent homogeneity and its molecular heterogeneity. *J. Exp. Med.* 156:454-464.
45. Robb, R.J., Kutny, R.M., and Chowdhry, V. 1983. Purification and partial sequence analysis of human T cell growth factor. *Proc. Natl. Acad. Sci. USA* 80:5990-5994.
46. Gillis, S., Ferm, M.M., Ou, W., and Smith, K.A. 1978. T-cell growth factor: Parameters of production and a quantitative microassay for activity. *J. Immunol.* 120:2027-2032.
47. Ortaldo, J.R., Mason, A.T., Gerard, J.P., Henderson, L.E., Farrar, W., Hopkins, R.F., III, Herberman, R.B., and Rabin, H. 1984. Effects of natural and recombinant IL2 on regulation of IFN gamma production and natural killer activity: Lack of involvement of the Tac antigen for these immunoregulatory effects. *J. Immunol.* 133:779-783.
48. Benveniste, E.N., and Merrill, J.E. 1986. Stimulation of oligodendroglial proliferation and maturation by interleukin-2. *Nature* 321:610-613.

49. Grimm, E.A., Robb, R.J., Roth, J.A., Neckers, L.M., Lachman, L.B., Wilson, O.J., and Rosenberg, S.A. 1983. Lymphokine activated killer cell phenomenon: III. Evidence that IL-2 is sufficient for direct activation of peripheral blood lymphocytes into lymphokine activated killer cells. *J. Exp. Med.* 158:1356-1361.
50. Depper, J.M., Leonard, W.J., Drogula, C., Kronke, M., Waldmann, T.A., and Greene, W.C. 1985. Interleukin-2 (IL-2) augments transcription of the interleukin-2 receptor gene. *Proc. Natl. Acad. Sci. USA* 82:4230-4234.
51. Farrar, W.J., and Anderson, W. 1985. Interleukin-2 stimulates association of protein kinase C with plasma membrane. *Nature* 315:233-236.
52. Ben Aribia, M-H, Leroy, E., Lantz, O., Metivier, D., Autran, B., Charpentier, B., Hercend, T., and Senik, A. 1987. rIL 2-induced proliferation of human circulating NK cells and T lymphocytes: synergistic effects of IL 1 and IL 2. *J. Immunol.* 139:443-451.
53. Benedict, S.H., Mills, G.B., and Gelfand, E.W. 1987. Interleukin 2 activates a receptor-associated protein kinase. *J. Immunol.* 139:1694-1697.
54. Taira, S., Matsui, M., Hayakawa, K., Yokoyama, T., and Nariuchi, H. 1987. Interleukin 2 secretion by B cell lines and splenic B cells stimulated with calcium ionophore and phorbol ester. *J. Immunol.* 139:2957-2964.
55. Mary, D., Aussel, C., Ferrua, B., and Fehlmann, M. 1987. Regulation of interleukin 2 synthesis by cAMP in human T cells. *J. Immunol.* 139:1179-1184.
56. Jelinek, D.F., J.B. Splawski, and Lipsky, P.E. 1986. The roles of interleukin 2 and interferon- γ in human B cell activation, growth, and differentiation. *Eur. J. Immunol.* 16:925-932.

IN VIVO ACTIVITIES

57. Buchan, G., Barrett, K., Fujita, T., Taniguchi, T., Maini, R., and Feldmann, M. 1988. Detection of activated T cell products in the rheumatoid joint using cDNA probes to interleukin 2 (IL-2), IL-2 receptors and IFN- γ . *Clin. Exp. Immunol.* 71:295-301.
58. Wilkins, J.A., Warrington, R.J., Sigurdson, S.L., and Rutherford, W.J. 1983. The demonstration of an interleukin 2-like activity in the synovial fluid of rheumatoid arthritis patients. *J. Rheum.* 10:109-113.
59. Nouri, A.M.E., Panayi, G.S., Goodman, S.M. 1984. Cytokines and the chronic inflammation of rheumatic disease. II. The presence of interleukin 2 in synovial fluids. *Clin. Exp. Immunol.* 58:402-409.
60. Rosenberg, S.A., Spiess, P.J., and Schwarz, S. 1983. *In vivo* administration of interleukin-2 enhances specific alloimmune responses. *Transplantation* 35:631-634.
61. Granelli-Piperno, A., Andrus, L., and Reich, E. 1984. Antibodies to interleukin 2. Effects on immune responses *in vitro* and *in vivo*. *J. Exp. Med.* 160:738-750.

62. Robb, R.J., Rusk, C.M., Yodoi, J., and Greene, W.C. 1987. An interleukin-2 binding molecule distinct from the Tac protein: Analysis of its role in formation of high affinity receptors. Proc. Natl. Acad. Sci. USA 84:2002-2006.
63. Chong, K-T. 1987. Prophylactic administration of interleukin-2 protects mice from lethal challenge with gram-negative bacteria. Infect. Immun. 55:668-673.
64. Weyand, C., Goronzy, J., Fathman, C.G., and O'Hanley, P. 1987. Administration *in vivo* of recombinant interleukin 2 protects mice against septic death. J. Clin. Invest. 79:1756-1763.
65. Lotze, M.T., Matory, Y.L., Ettinghausen, S.E., Rayner, A.A., Sharow, S.O., Seipp, C.A.Y., Custer, M.C., and Rosenberg, S.A. 1985. *In vivo* administration of purified human interleukin 2 II. Half life, immunologic effects, and expansion of peripheral lymphoid cells *in vivo* with recombinant IL 2. J. Immunol. 135:2865-2875.
66. Kolitz, J.E., Welte, K., Wong, G.Y., Holloway, K., Merluzzi, V.J., Engert, A., Bradley, E.C., Konrad, M., Polivka, A., Gabrilove, J.L., Sykora, K.W., Miller, G.A., Fiedler, W., Krown, S., Oettgen, H.F., and Mertelsmann, R. 1987. Expansion of activated T-lymphocytes in patients treated with recombinant interleukin 2. J. Biol. Response Modifiers 6:412-429.
67. Rosenberg, S.A., Lotze, M.T., Muul, L.M., Leitman, S., Chang, A.E., Ettinghausen, S.E., Matory, Y.L., Skibber, J.M., Shilone, E., Vetto, J.T., Seipp, C.A., Simpson, C., Reichert, C.M. 1985. Observations on the systemic administration of autologous lymphokine-activated killer cells and recombinant interleukin-2 to patients with metastatic cancer. N. Engl. J. Med. 313:1485-1490.
68. Colvin, R.B., Fuller, T.C., MacKeen, L., Kung, P.C., Ip, s.H., and Cosimi, A.B. 1987. Plasma interleukin 2 receptor levels in renal allograft recipients. Clin. Immunol. Immunopathol. 43:273-276.

Interleukin 4 (IL-4)

GENERAL

1. Kishimoto, T. 1987. B-cell stimulatory factors (BSFs): Molecular structure, biological function, and regulation of expression. *J. Clin. Immunol.* 7:343-355.
2. Paul, W.E. 1987 Interleukin 4/B cell stimulatory factor 1: one lymphokine, many functions. *FASEB J.* 1:456-461.

GENE STRUCTURE

3. Yokota, T., Otsuka, T., Mosmann, T., Banchereau, J., DeFrance, T., Blanchard, D., DeVries, J.E., Lee, F., and Arai, K-I. 1986. Isolation and characterization of a human interleukin cDNA clone, homologous to mouse B-cell stimulatory factor 1, that expresses B-cell- and T-cell-stimulating activities. *Proc. Natl. Acad. Sci. USA* 83:5894-5898.

RECEPTOR

4. Park, L.S., Friend, D., Sassenfeld, H.M., and Urdal, D.L. 1987. Characterization of the human B cell stimulatory factor 1 receptor. *J. Exp. Med.* 166:476-488.
5. Lowenthal, J.W., Castle, B.E., Christiansen, J., Schreurs, J., Rennick, D., Arai, N., Hoy, P., Takebe, Y., and Howard, M. 1988. Expression of the high affinity receptors for murine interleukin 4 (BSF-1) on hemopoietic and nonhemopoietic cells. *J. Immunol.* 140:456-464.

PHYSIOLOGY

6. Brown, M.A., Pierce, J.H., Watson, C.J., Falco, J., Ihle, J.N., and Paul, W.E. 1987. B cell stimulatory factor-1/interleukin-4 mRNA is expressed by normal and transformed mast cells. *Cell* 50:809-818.
7. Rennick, D., Yang, G., Muller-Sieburg, C., Smith, C., Arai, N., Takabe, Y., and Gemmell, L. 1987. Interleukin 4 (B-cell stimulatory factor 1) can enhance or antagonize the factor-dependent growth of hemopoietic progenitor cells. *Proc. Natl. Acad. Sci. USA* 84:6889-6893.
8. Snapper, C.M., and Paul, W.E. 1987. B cell stimulatory factor-1 (Interleukin 4) prepares resting murine B cells to secrete IgG1 upon subsequent stimulation with bacterial lipopolysaccharide. *J. Immunol.* 139:10-17.
9. Snapper, C.M., Finkelman, F.D., and Paul, W.E. 1988. Differential regulation of IgG1 and IgE synthesis by interleukin 4. *J. Exp. Med.* 167:183-196.
10. Defrance, T., Aubry, J.P., Rousset, F., Vanberviliet, B., Bonnefoy, J.Y., Arai, N., Takebe, Y., Yokota, T., Lee, F., Arai, K., DeVries, J., and Banchereau, J. 1987. Human recombinant interleukin 4 induces Fc receptors (CD23) on normal human B lymphocytes. *J. Exp. Med.* 165:1459-1467.

11. Defrance, T., Vanbervliet, B., Aubry, J-P., Takebe, Y., Arai, N., Miyajima, A., Yokota, T., Lee, F., Arai, K-I., deVries, J.E., and Banchereau, J. 1987. B cell growth-promoting activity of recombinant human interleukin 4. *J. Immunol.* 139:1135-1141.
12. Hu-Li, J., Shevach, E.M., Mizucuchi, J., Ohara, J., Mosmann, T., and Paul, W.E. 1987. B cell stimulatory factor 1 (interleukin 4) is a potent costimulant for normal resting T lymphocytes. *J. Exp. Med.* 165:157-172.
13. Spits, H., Yssel, H., Takebe, Y., Arai, N., Yokota, T., Lee, F., Arai, K-I., Benchereau, J., and deVries, J.E. 1987. Recombinant interleukin 4 promotes the growth of human T cells. *J. Immunol.* 139:1142-1147.
14. Widmer, M.B., Acres, R.B., Sassenfeld, H.M., and Grabstein, K.H. 1987. Regulation of cytolytic cell populations from human peripheral blood by B cell stimulatory factor 1 (interleukin 4). *J. Exp. Med.* 166:1447-1455.
15. Zlotnik, A., Fischer, M., Roehm, N., and Zipori, D. 1987. Evidence for effects of interleukin 4 (B cell stimulatory factor 1) on macrophages: Enhancement of antigen presenting ability of bone marrow-derived macrophages. *J. Immunol.* 138:4275-4279.
16. Crawford, R.M., Finbloom, D.S., Ohara, J., Paul, W.E., and Meltzer, M.S. 1987. B cell stimulatory factor-1 (interleukin 4) activates macrophages for increased tumoricidal activity and expression of Ia antigens. *J. Immunol.* 139:135-141.
17. McInnes, A., and Rennick, D.M. 1988. Interleukin 4 induces cultured monocytes/macrophages to form giant multinucleated cells. *J. Exp. Med.* 167:598-611.

IN VIVO EFFECTS

18. Finkelman, F.D., Katona, I.M., Urban, J.F., Jr., Snapper, C.M., Ohara, J. and Paul, W.E. 1986. Suppression of *in vivo* polyclonal IgE responses by monoclonal antibody to the lymphokine B-cell stimulatory factor 1. *Proc. Natl. Acad. Sci. USA* 83:9675-9678.

Interleukin 5 (IL5)

GENERAL

1. Harriman, G.R., and Strober, W. 1987. Interleukin 5, a mucosal lymphokine? *J. Immunol.* 139:3553-3555.
2. Kishimoto, T. 1987. B-cell stimulatory factors (BSFs): Molecular structure, biological function, and regulation of expression. *J. Clin. Immunol.* 7:343-355.

GENE STRUCTURE

3. Campbell, H.D., Tucker, W.Q.J., Hort, Y., Martinson, M.E., Mayo, G., Clutterbuck, E.J., Sanderson, C.J., Young, I.G. 1987. Molecular cloning, nucleotide sequence, and expression of the gene encoding human eosinophil differentiation factor (interleukin 5). *Proc. Natl. Acad. Sci. USA* 84:6629-6633.
4. Yokota, T., Coffman, R.L., Hagiward, H., Rennick, D.M., Takebe, Y., Yokota, K., Gemmell, L., Shrader, B., Yang, G., Meyerson, P., Luh, J., Hoy, P., Pene, J., Briere, F., Spits, H., Banchereau, J., DeVries, J., Lee, F.D., Arai, N., and Arai, K-I. 1987. Isolation and characterization of lymphokine cDNA clones encoding mouse and human IgA-enhancing factor and eosinophil colony-stimulating factor activities: Relationship to interleukin 5. *Proc. Natl. Acad. Sci. USA* 84:7388-7392.

PHYSIOLOGY

5. Rasmussen, R., Takatsu, K., Harada, N., Takahashi, T., and Bottomly, K. 1988. T cell-dependent hapten-specific and polyclonal B cell responses require release of interleukin 5. *J. Immunol.* 140:705-712.
6. Takatsu, K., Kikuchi, Y., Takahashi, T., Honjo, T., Matsumoto, M., Harada, N., Yamaguchi, N., and Tominaga, A. 1987. Interleukin 5, a T-cell-derived B-cell differentiation factor also induces cytotoxic T lymphocytes. *Proc. Natl. Acad. Sci. USA* 84:4234-4238.
7. Waren, D.J., and Moore, M.A.S. 1988. Synergism among interleukin 1, interleukin 3, and interleukin 5 in the production of eosinophils from primitive hemopoietic stem cells. *J. Immunol.* 140:94-99.
8. Loughnan, M.S., Takatsu, K., Harada, N., and Nossal, G.J.V. 1987. T-cell-replacing factor (interleukin 5) induces expression of interleukin 2 receptors on murine splenic B cells. *Proc. Natl. Acad. Sci. USA* 84:5399-5403.

Interleukin 6 (IL6)

GENERAL

1. Kishimoto, T. 1987. B-cell stimulatory factors (BSFs): Molecular structure, biological function, and regulation of expression. *J. Clin. Immunol.* 7:343-355.
2. Billiau, A. 1987. Interferon β_2 as a promoter of growth and differentiation of B cells. *Immunol. Today* 8:84-87.
3. Sehgal, P.B., May, L.T., Tamm, I., and Vilcek, J. 1987. Human β_2 interferon and B-cell differentiation factor BSF-2 are identical. *Science* 235:731-732.

GENE STRUCTURE

4. Zilberstein, A., Ruggieri, R., Korn, J.H., and Revel, M. 1986. Structure and expression of cDNA and genes for human interferon-beta-2, a distinct species inducible by growth-stimulatory cytokines. *EMBO J.* 5:2529-2537.
5. Sehgal, P.B., Zilberstein, A., Ruggieri, R.-M., May, L.T., Ferguson-Smith, A., Slate, D.L., Revel, M., and Ruddle, F.H. 1986. Human chromosome 7 carries the B_2 interferon gene. *Proc. Natl. Acad. Sci. USA* 83:5219-5222.
6. Hirano, T., Yasukawa, K., Harada, H., Taga, T., Watanabe, Y., Matsuda, T., Kashiwamura, S-I., Nakajima, K., Koyama, K., Iwamatsu, A., Tsunasawa, S., Sakiyama, F., Matsui, H., Takahara, Y., Taniguchi, T., and Kishimoto, T. 1986. Complementary DNA for a novel human interleukin (BSF-2) that induces B lymphocytes to produce immunoglobulin. *Nature* 324:73-76.
7. Gauldie, J., Richards, C., Harnish, D., Lansdorp, P., and Baumann, H. 1987. Interferon β_2 /B-cell stimulatory factor type 2 shares identity with monocyte-derived hepatocyte-stimulating factor and regulates the major acute phase protein response in liver cells. *Proc. Natl. Acad. Sci. USA* 84:7251-7255.
8. Brakenhoff, J.P.J., deGroot, E.R., Evers, R.F., Pannekoek, H., and Aarden, L.A. 1987. Molecular cloning and expression of hybridoma growth factor in *Escherichia coli*. *J. Immunol.* 139:4116-4121.

RECEPTOR

9. Taga, T., Kawanishi, Y., Hardy, R.R., Hirano, T., and Kishimoto, T. 1987. Receptors for B cell stimulatory factor 2. Quantitation, specificity, distribution, and regulation of their expression. *J. Exp. Med.* 166:967-981.

PHYSIOLOGY

10. Zilberstein, A., Ruggieri, R., and Revel, M. 1985. Human interferon- β_2 : is it an interferon-inducer? In *The Interferon System*. Eds. Rossi, G.B., and Dianzani, E. Raven Press, New York, Vol. 24, pp. 73-83.

11. Hirano, T., Taga, T., Nakano, N., Yasukawa, K., Kashiwamura, S., Shimizu, K., Nakajima, K., Pyun, K.H., and Kishimoto, T. 1985. Purification to homogeneity and characterization of human B-cell differentiation factor (BCDF or BSFp-2). Proc. Natl. Acad. Sci. USA 82:5490-5494.
12. Content, J., DeWit, L., Poupart, P., Opdenakker, G., vanDamme, J., and Billiau, A. 1985. Induction of a 26-kDa-protein mRNA in human cells treated with an interleukin-1-related, leukocyte-derived factor. Eur. J. Biochem. 152:253-257.
13. Kohase, M., Henriksen-DeStefano, D., May, L.T., Vilcek, J., and Sehgal, P.B. 1986. Induction of β_2 -interferon by tumor necrosis factor: A homeostatic mechanism in the control of cell proliferation. Cell 45:659-666.
14. Walther Z., May, L.T., and Sehgal, P.B. 1988. Transcriptional regulation of the interferon- β_2 /B cell differentiation factor BSF-2/hepatocyte-stimulating factor gene in human fibroblasts by other cytokines. J. Immunol. 140:974-977.
15. Ikebuchi, K., Wong, G.G., Clark, S.C., Ihle, J.N., Hirai, Y., and Ogawa, M. 1987. Interleukin 6 enhancement of interleukin 3-dependent proliferation of multipotential hemopoietic progenitors. Proc. Natl. Acad. Sci. USA 84:9035-9039.
16. Garman, R.D., Jacobs, K.A., Clark, S.C., and Raulet, D.H. 1987. B-cell-stimulatory factor (β_2 interferon) functions as a second signal for interleukin 2 production by mature murine T cells. Proc. Natl. Acad. Sci. USA 84:7629-7633.
17. Takai, Y., Wong, G.G., Clark, S.C., Burakoff, S.J., and Herrmann, S.H. 1988. B cell stimulatory factor-2 is involved in the differentiation of cytotoxic T lymphocytes. J. Immunol. 140:508-512.
18. Baumann, H., Richards, C., and Gauldie, J. 1987. Interaction among hepatocyte-stimulating factors, interleukin 1, and glucocorticoids for regulation of acute phase plasma proteins in human hepatoma (HepG2) cells. J. Immunol. 139:4122-4128.
19. May, L.T., Helfgott, D.C., and Sehgal, P.B. 1986. Anti-B-interferon antibodies inhibit the increased expression of HLA-B7 mRNA in tumor necrosis factor-treated human fibroblasts: Structural studies of the β_2 interferon involved. Proc. Natl. Acad. Sci. USA 83:8957-8961.
20. Tosata, G., Seamon, K.B., Goldman, N.D., Sehgal, P.B., May, L.T., Washington, G.C., Jones, K.D., and Pike, S.E. 1988. Monocyte-derived human B-cell growth factor identified as interferon- β_2 (BSF-2, IL-6). Science 239:502-504.
21. Kawano, M., Hirano, T., Matsuda, T., Taga, T., Horii, Y., Iwato, K., Asaoku, H., Tang, B., Tanabe, O., Tanaka, H., Kuramoto, A., and Kishimoto, T. 1988. Autocrine generation and requirement of BSF-2/IL6 for human multiple myelomas. Nature 332:83-85.
22. Muraguchi, A., Hirano, T., Tang, B., Matsuda, T., Horii, Y., Nakajima, K., and Kishimoto, T. 1988. The essential role of B cell stimulatory factor 2 (BSF-2/IL-6) for the terminal differentiation of B cells. J. Exp. Med. 167:332-344.

IN VIVO EFFECTS

23. Houssiau, F.A., de Deuxchaisnes, C.N., and Van Snick, J. 1988. Interleukin-6 in synovial fluid and serum of patients with rheumatoid arthritis and other inflammatory arthritides. *Arthritis Rheum*, in press.
24. Hirano, T., Matsuda, T., Turner, M., Miyasaka, N., Buchan, G., Tang, B., Sato, K., Shimizu, M., Maini, R., Feldmann, M., and Kishimoto, T. 1988. Excessive production of B cell stimulatory factor-2 (BSF-2) in rheumatoid arthritis. (Manuscript submitted for publication)

Lymphotoxin

GENE STRUCTURE

1. Gray, P.W., Aggarwal, B.B., Benton, C.V., Bringman, T.S., Henzel, W.J., Jarrett, J.A., Leung, D.W., Moffat, B., Ng, P., Svedersky, L.P., Palladino, M.A., and Nedwin, G.E. 1984. Cloning and expression of cDNA for human lymphotoxin, a lymphokine with tumour necrosis activity. *Nature* 312:721-724.

RECEPTOR

2. Aggarwal, B.B., Eessalu, T.E., and Hass, P.E. 1985. Characterization of receptors for human tumour necrosis factor and their regulation by γ -interferon. *Nature* 318:665-667.

PHYSIOLOGY

3. Aggarwal, B.B., Moffat, B., and Harkins, R.N. 1984. Human lymphotoxin. Production by a lymphoblastoid cell line, purification, and initial characterization. *J. Biol. Chem.* 259:686-691.
4. Aggarwal, B.B., Henzel, W.J., Moffat, B., Kohr, W.J., and Harkins, R.N. 1985. Primary structure of human lymphotoxin derived from 1788 lymphoblastoid cell line. *J. Biol. Chem.* 260:2334-2344.
5. Kehrl, J.H., Alvarez-Mon, M., Delsing, G.A., and Fauci, A.S. 1987. Lymphotoxin is an important T cell-derived growth factor for human B cells. *Science* 238:1144-1146.

Gamma Interferon

GENERAL

1. Lengyel, P. 1982. Biochemistry of interferons and their actions. *Annu. Rev. Biochem.* 51:251-282.
2. Bonnem, E.M., and Oldham, R.K. 1986. Gamma-interferon: Physiology and speculation on its role in medicine. *J. Biol. Response Modifiers* 6:275-301.
3. Billiau, A. 1988. Gamma-interferon: the match that lights the fire? *Immunol. Today.* 9:37-40.

GENE STRUCTURE

4. Gray, P.W., and Goeddel, D.V. 1982. Structure of the human immune interferon gene. *Nature* 298:859-863.
5. Friedman, R.L., Manly, S.P., McMahon, M., Kerr, I.M., and Stark, G.R. 1984. Transcriptional and posttranscriptional regulation of interferon-induced gene expression in human cells. *Cell* 38:745-755.
6. Gray, P., and Goeddel, D.V. 1982. Structure of the human immune interferon gene. *Nature* 298:859-863.
7. Rashidbaigi, A., Langer, J.A., Jung, V., Jones, C., Morse, H.G., Tischfield, J.A., Trill, J.J., Kung, H.F., and Pestka, S. 1986. The gene for the human immune interferon receptor is located on chromosome 6. *Proc. Natl. Acad. Sci. USA* 83:384-388.

RECEPTOR

8. Aguet, M. 1980. High-affinity binding of ^{125}I -labeled mouse interferon to a specific cell surface receptor. *Nature* 284:459-461.
9. Anderson, P., Yip, Y.K., and Vilcek, J. 1982. Specific binding of ^{125}I -human interferon- γ to high affinity receptors on human fibroblasts. *J. Biol. Chem.* 257:11301-11304.
10. Anderson, P., Yip, Y.K., and Vilcek, J. 1983. Human interferon- γ is internalized and degraded by cultured fibroblasts. *J. Biol. Chem.* 258:6497-6502.
11. Branca, A.A., and Baglioni, C. 1981. Evidence that type I and type II interferons have different receptors. *Nature* 294:768-770.
12. Branca, A.A., Faltynek, C.R., D'Alessandro, S.B., and Baglioni, C. 1982. Interaction of interferon with cellular receptors: Internalization and degradation of cell-bound interferon. *J. Biol. Chem.* 257:13291-13296.

13. Celada, A., Allen, A., Esparza, I., Gray, P.W., and Schreiber, R.D. 1986. Demonstration and partial characterization of the interferon-gamma receptor on human monocytes and human monocyte-like cell lines. *J. Clin. Invest.* 76:2196-2205.
14. Faltynek, C.R., Branca, A.A., McCandless, S., and Baglioni, C. 1983. Characterization of interferon receptor on human lymphoblastoid cells. *Proc. Natl. Acad. Sci. USA* 80:3269-3273.
15. Littman, S.J., Faltynek, C.K., and Baglioni, C. 1985. Binding of human recombinant ^{125}I -interferon- γ to receptors on human cells. *J. Biol. Chem.* 260:1191-1195.
16. Rashidbaigi, A., Kung, H., and Pestka, S. 1985. Characterization of receptors for immune interferon in U937 cells with ^{32}P -labeled human recombinant immune interferon. *J. Biol. Chem.* 260:8514-8519.
17. Sarkar, F.H., and Gupta, S.L. 1984. Receptors for human γ interferon: Binding and cross-linking of ^{125}I -labeled human recombinant γ interferon to receptors on WISH cells. *Proc. Natl. Acad. Sci. USA* 81:5160-5164.

PHYSIOLOGY

18. Rinderknecht, E., O'Connor, B.H., and Rodriguez, H. 1984. Natural human interferon- γ : Complete amino acid sequence and determination of sites of glycosylation. *J. Biol. Chem.* 259:6790-6797.
19. Akagawa, K.S., and Tokunaga, T. 1985. Lack of binding of bacterial lipopolysaccharide to mouse lung macrophages and restoration of binding by gamma interferon. *J. Exp. Med.* 162:1444-1459.
20. Bhardwaj, N., Nash, T.W., and Horwitz, M.A. 1986. Interferon- γ activated human monocytes inhibit the intracellular multiplication of *Legionella pneumophila*. *J. Immunol.* 137:2662-2669.
21. Boraschi, D., Censini, S., Bartalini, M., and Tagliabue, A. 1985. Regulation of arachidonic acid metabolism in macrophages by immune and nonimmune interferons. *J. Immunol.* 135:502-505.
22. Brandwein, S.R. 1986. Regulation of interleukin 1 production by mouse peritoneal macrophages: Effects of arachidonic acid metabolites, cyclic nucleotides, and interferons. *J. Biol. Chem.* 261:8624-8632.
23. Cassatella, M.A., Della-Bianca, V., Berton, G., and Rossi, F. 1985. Activation by gamma interferon of human macrophage capability to produce toxic oxygen molecules is accompanied by decreased K_m of the superoxide-generating NADPH oxidase. *Biochem. Biophys. Res. Commun.* 132:908-914.
24. Esparza, I., Fox, R.I., and Schreiber, R.D. 1986. Interferon-gamma-dependent modulation of C3b receptors (CR1) on human peripheral blood monocytes. *J. Immunol.* 136:1360-1365.

25. Ezekowitz, R.A.B., Bampton, M., and Gordon, S. 1983. Macrophage activation selectively enhances expression of Fc receptors for IgG2a. *J. Exp. Med.* 157:807-812.
26. Guyre, P.M., Morganelli, P.M., and Miller, R. 1983. Recombinant interferon gamma increases immunoglobulin G Fc receptors on cultured human mononuclear phagocytes. *J. Clin. Invest.* 72:393-397.
27. Hamilton, T.A., Ribsbee, J.E., Scott, W.A., and Adams, D.O. 1985. Gamma-interferon enhances the secretion of arachidonic acid metabolites from murine peritoneal macrophages stimulated with phorbol diesters. *J. Immunol.* 134:2631-2636.
28. Hamilton, T.A., Somers, S.D., Becton, D.L., Celada, A., Schreiber, R.D., and Adams, D.O. 1986. Analysis of deficiencies in IFN- γ -mediated priming for tumor cytotoxicity in peritoneal macrophages from A/J mice. *J. Immunol.* 137:3367-3371.
29. Mentzer, S.J., Faller, D.V., and Burakoff, S.J. 1986. Interferon-gamma induction of LFA-1-mediated homotypic adhesion of human monocytes. *J. Immunol.* 137:109-113.
30. Murray, H.W., Rubin, B.Y., and Rothermel, C.D. 1983. Killing of intracellular *Leishmania donovani* by lymphokine-stimulated human mononuclear phagocytes: Evidence that interferon- γ is the activating lymphokine. *J. Clin. Invest.* 72:1506-1510.
31. Nathan, C.F., Murray, H.W., Wiebe, M.E., and Rubin, B.Y. 1983. Identification of interferon- γ as the lymphokine that activates human macrophage oxidative metabolism and antimicrobial activity. *J. Exp. Med.* 158:670-689.
32. Nathan, C.F., Prendergast, T.J., Wiebe, M.E., Stanley, E.R., Platzer, E., Remold, H.G., Welte, K., Rubin, B.Y., and Murray, H.W. 1984. Activation of human macrophages: Comparison of other cytokines with interferon- γ . *J. Exp. Med.* 160:600-605.
33. Perussia, B., Dayton, E.T., Lazarus, R., Fanning, V., and Trinchieri, G. 1983. Immune interferon induces the receptor for monomeric IgG1 on human monocytic and myeloid cells. *J. Exp. Med.* 158:1092.
34. Rothermel, C.D., Rubin, B.Y., and Murray, H.W. 1983. γ -interferon is the factor in lymphokine that activates human macrophages to inhibit intracellular *Chlamydia psittaci* replication. *J. Immunol.* 131:2542-2544.
35. Strunk, R.C., Cole, F.S., Perlmutter, D.H., and Colten, H.R. 1985. γ -interferon increases expression of class III complement genes C2 and factor B in human monocytes and in murine fibroblasts. *J. Biol. Chem.* 260:15280-15285.
36. Schultz, R.M., and Kleinschmidt, W.J. 1983. Functional identity between immune γ interferon and macrophage activating factor. *Nature* 305:239-240.
37. Arenzana-Seisdedos, F., and Virelizier, J-L. 1983. Interferons as macrophage-activating factors II. Enhanced secretion of interleukin 1 by lipopolysaccharide-stimulated human monocytes. *Eur. J. Immunol.* 13:437-440.

38. Haq, A.U., and Maca, R.D. 1986. Role of IFN- γ and α in IL 1 synthesis and secretion of *in vitro* differentiated human macrophages: A comparative study. *Immunobiol.* 171:451-460.
39. Gerrard, T.L., Siegel, J.P., Dyer, D.R., and Zoon, K.C. 1987. Differential effects of interferon- α and interferon- γ on interleukin 1 secretion by monocytes. *J. Immunol.* 138:2535-2540.
40. Collart, M.A., Belin, D., Vassalli, J-D., deKossodo, S., and Vassalli, P. 1986. γ Interferon enhances macrophage transcription of the tumor necrosis factor/cachectin, interleukin 1, and urokinase genes, which are controlled by short-lived repressors. *J. Exp. Med.* 164:2113-2118.
41. Thurman, G.B., Braude, I.A., Gray, P.W., Oldham, R.K., and Stevenson, H.C. 1985. MIF-like activity of natural and recombinant human interferon- γ and their neutralization by monoclonal antibody. *J. Immunol.* 134:305-309.
42. Basham, T.Y., Nickoloff, B.J., Merigan, T.C., and Morhenn, V.B. 1984. Recombinant gamma interferon induces HLA-DR expression on cultured human keratinocytes. *J. Invest. Dermatol.* 83:88-90.
43. Berman, B., Duncan, M.R., Smith, B., Ziboh, V.A., and Palladino, M. 1985. Interferon enhancement of HLA-DR antigen expression on epidermal Langerhans cells. *J. Invest. Dermatol.* 84:54-58.
44. Houghton, A.N., Thomson, T.M., Gross, D., Oettgen, H.F., and Old, L.J. 1984. Surface antigens of melanoma and melanocytes: Specificity of induction of Ia antigens by human γ -interferon. *J. Exp. Med.* 160:255-269.
45. Inaba, K., Kitaura, M., Kato, T., Watanabe, Y., and Muramatsu, S. 1986. Contrasting effect of α/β - and γ -interferons on expression of macrophage Ia antigens. *J. Exp. Med.* 163:1030-1035.
46. Pober, J.S., Gimbrone, M.A., Jr., Cotran, R.S., Reiss, C.S., Burakoff, S.J., Fiers, W., and Ault, K.A. 1983. Ia expression by vascular endothelium is inducible by activated T cells and human γ interferon. *J. Exp. Med.* 157:1339-1353.
47. Steeg, P.S., Moore, R.N., Johnson, M., and Oppenheim, J.J. 1982. Regulation of murine macrophage Ia antigen expression by a lymphokine with immune interferon activity. *J. Exp. Med.* 156:1780-1793.
48. Korber, B., Mermod, N., Hood, L., and Stroynowski, I. 1988. Regulation of gene expression by interferons: Control of H-2 promoter responses. *Nature* 239:1302-1306.
49. Geppert, T.D., and Lipsky, P.E. 1985. Antigen presentation by gamma interferon treated endothelial cells and fibroblasts: Differential ability to function as antigen presenting cells despite comparable Ia expression. *J. Immunol.* 135:3750-3762.
50. Geppert, T.D., and Lipsky, P.E. 1987. Dissection of defective antigen presentation by gamma interferon treated fibroblasts. *J. Immunol.* 138:385-392.

51. Duijvestijn, A.M., Schreiber, A.B., and Butcher, E.C. 1986. Interferon- γ regulates an antigen specific for endothelial cells involved in lymphocyte traffic. Proc. Natl. Acad. Sci. USA 83:9114-9118.
52. Duncan, M.R., and Berman, M. 1985. Gamma interferon is the lymphokine and beta interferon the monokine responsible for inhibition of fibroblast collagen production and late but not early fibroblast proliferation. J. Exp. Med. 162:516-527.
53. Dustin, M.L., Rothlein, R., Bhan, A.K., Dinarello, C.A., and Springer, T.A. 1986. Induction by IL 1 and interferon-gamma: Tissue distribution, biochemistry, and function of a natural adherence molecule (ICAM-1). J. Immunol. 137:245-254.
54. Goldring, M.B., Sandell, J.J., Stephenson, M.L., and Krane, S.M. 1986. Immune interferon suppresses levels of procollagen mRNA and type II collagen synthesis in cultured human articular and costal chondrocytes. J. Biol. Chem. 261:9049-9055.
55. Jimenez, S.A., Freundlich, B., and Rosenbloom, J. 1984. Selective inhibition of human diploid fibroblast collagen synthesis by interferons. J. Clin. Invest. 74:1112-1116.
56. Turco, J., and Winkler, H.H. 1983. Cloned mouse interferon- γ inhibits the growth of *Rickettsia prowazekii* in cultured mouse fibroblasts. J. Exp. Med. 158:2159-2164.
57. Murphy, M., Loudon, R., Kobayashi, M., and Trinchieri, G. 1986. Gamma interferon and lymphotoxin, released by activated T cells, synergize to inhibit granulocyte/monocyte colony formation. J. Exp. Med. 164:263-279.
58. Zoumbos, N.C., Djeu, J.Y., and Young, N.S. 1984. Interferon is the suppressor of hematopoiesis generated by stimulated lymphocytes *in vitro*. J. Immunol. 133:769-774.
59. Broxmeyer, H.E., Lu, L., Platzer, E., Feit, C., Juliano, L., and Rubin, B.Y. 1983. Comparative analysis of the influences of human gamma, alpha, and beta interferons on human multipotential (CFU-GEMM), erythroid (BFU-E) and granulocyte-macrophage (CFU-GM) progenitor cells. J. Immunol. 131:1300-1305.
60. Rabin, E.M., Mond, J.J., Ohara, J., and Paul, W.E. 1986. Interferon- γ inhibits the action of B cell stimulatory factor (BSF-1) on resting B cells. J. Immunol. 137:1573-1576.
61. Svedersky, L.P., Nedwin, G.E., Goeddel, D.V., and Palladino, M.A., Jr. 1985. Interferon-gamma enhances induction of lymphotoxin in recombinant interleukin 2-stimulated peripheral blood mononuclear cells. J. Immunol. 134:1604-1608.
62. Herrmann, F., Cannistra, S.A., Levine, H., and Griffin, J.D. 1985. Expression of interleukin 2 receptors and binding of interleukin 2 by gamma interferon-induced human leukemic and normal monocytic cells. J. Exp. Med. 162:1111-1116.
63. Tsujimoto, M., Yip, Y.K., and Vilcek, J. 1986. Interferon- γ enhances expression of cellular receptors for tumor necrosis factor. J. Immunol. 136:2441-2444.

64. Patton, J.S., Shepard, H.M., Wilking, H., Lewis, G., Aggarwal, B.B., Eesalu, T.E., Gavin, L.A., and Grunfeld, C. 1986. Interferons and tumor necrosis factors have similar catabolic effects on 3T3 L1 cells. Proc. Natl. Acad. Sci. USA 83:8313-8317.

IN VIVO ACTIVITIES

65. Browning, J. 1987. Interferons and rheumatoid arthritis: insight into interferon biology? Immunol. Today 8:372-374.
66. Degre, M., Mellbye, O.J., and Clarke-Jenssen, O. 1983. Immune interferon in serum and synovial fluid in rheumatoid arthritis and related disorders. Ann. Rheum. Dis. 42:672-676.
67. Husby, G., and Williams, R.C., Jr. 1985. Immunohistochemical studies of interleukin-2 and gamma-interferon in rheumatoid arthritis. Arthritis Rheum. 28:174-181.
68. Kurzrock, R., Rohde, M.F., Quesada, J.R., Gianturco, S.H., Bradley, W.A., Sherwin, S.A., and Guterman, J.U. 1986. Recombinant gamma interferon induces hypertriglyceridemia and inhibits post-heparin lipase activity. J. Exp. Med. 164:1093-1101.
69. Momburg, F., Koch, N., Moller, P., Moldenhauer, G., and Hammerling, G.J. 1986. *In vivo* induction of H-2K/D antigens by recombinant interferon-gamma. Eur. J. Immunol. 16:551-557.
70. Skoskiewicz, M.J., Colvin, R.B., Scheeberger, E.E., and Russell, P.S. 1985. Widespread and selective induction of major histocompatibility complex-determined antigens *in vivo* by gamma interferon. J. Exp. Med. 162:1645-1664.
71. Murray, H.W., Stern, J.J., Welte, K., Rubin, B.Y., Carriero, S., and Nathan, C.F. 1987. Experimental visceral leishmaniasis: Production of interleukin 2 and interferon- γ , tissue immune reaction, and response to treatment with interleukin 2 and interferon- γ . J. Immunol. 138:2290-2297.
72. Kiderlen, A.F., Kaufmann, S.H.E., and Lohmann-Matthes, M-L. 1984. Protection of mice against the intracellular bacterium *Listeria monocytogenes* by recombinant immune interferon. Eur. J. Immunol. 14:964-967.
73. Murray, H.W., Spitalny, G.L., and Nathan, C.F. 1985. Activation of mouse peritoneal macrophages *in vitro* and *in vivo* by interferon- γ . J. Immunol. 134:1619-1622.
74. Buchmeier, N.A., and Schreiber, R.D. 1985. Requirement of endogenous interferon- γ production for resolution of *Listeria monocytogenes* infection. Proc. Natl. Acad. Sci. USA 82:7404-7408.
75. Shalaby, M.R., Hamilton, E.G., Benninger, A.H., and Marafino, B.J., Jr. 1985. *In vivo* antiviral activity of recombinant murine gamma interferon. J. Interferon Res. 5:339-345.

76. Landolfo, S., Cofano, F., Giovarelli, M., Prat, M., Cavallo, G., and Forni, G. 1985. Inhibition of interferon-gamma may suppress allograft reactivity by T lymphocytes *in vitro* and *in vivo*. *Science* 229:176-179.
77. Jacob, C.O., Van Der Meide, P.H., and McDevitt, H.O. 1987. *In vivo* treatment of (NZB X NZW)F₁ lupus-like nephritis with monoclonal antibody to γ interferon. *J. Exp. Med.* 166:798-803.
78. Nathan, C.F., Kaplan, G., Levis, W.R., Nusrat, A., Witmer, M.D., Sherwin, S.A., Job, C.K., Path, F.R.C., Horowitz, C.R., Steinman, R.M., and Cohn, Z.A. 1986. Local and systemic effects of intradermal recombinant interferon- γ in patients with lepromatous leprosy. *N. Engl. J. Med.* 315:6-15.
79. Murray, H.W., Hillman, J.K., Rubin, B.Y., Kelly, C.D., Jacobs, J.L., Tyler, L.W., Donelly, D.M., Carriero, S.M., Godbold, J.H., and Roberts, R.B. 1985. Patients at risk for AIDS-related opportunistic infections: Clinical manifestations and impaired gamma-interferon production. *N. Engl. J. Med.* 313:1504-1510.
80. Panitch, H.S., Hirsch, R.L., Schindler, J., and Johnson, K.P. 1987. Treatment of multiple sclerosis with gamma interferon. Exacerbations associated with activation of the immune system. *Neurology* 37:1097-1102.
81. Ferreira, A., Schofield, L., Enea, V., Schellekens, H., Vander Meide, P., Collins, W.E., Nussenzweig, R.S., and Nussenzweig, V. 1986. Inhibition of development of exoerythrocytic forms of malaria parasites by γ -interferon. *Science* 232:881-884.

Colony Stimulating Factors

GENERAL

1. Metcalf, D. 1985. The granulocyte-macrophage colony-stimulating factors. *Science* 229:16-22.
2. Sieff, C.A. 1987. Hematopoietic growth factors. *J. Clin. Invest.* 79:1549-1557.
3. Clark, S.C., and Kamen, R. 1987. The human hematopoietic colony-stimulating factors. 1987. The human hematopoietic colony-stimulating factors. *Science* 236:1229-1237.
4. Walker, F., Nicola, N.A., Metcalf, D., and Burgess, A.W. 1985. Hierarchical down-modulation of hemopoietic growth factor receptors. *Cell* 43:269-276.

GENE STRUCTURE

5. Wong, G.G, Witek, J.S., Temple, P.A., Wilkens, K.M., Leary, A.C., Luxenberg, D.P., Jones, S.S., Brown, E.L., Kay, R.M., Orr, E.C., Shoemaker, C., Golde, D.W., Kaufman, R.J., Hewick, R.M., Wang, E.A., and Clark, S.C. 1985. Human GM-CSF: Molecular cloning of the complementary DNA and purification of the natural and recombinant proteins. *Science* 228:810-815.
6. Thorens, B., Mermod, J-J., and Vassalli, P. 1987. Phagocytosis and inflammatory stimuli induce GM-CSF mRNA in macrophages through posttranscriptional regulation. *Cell* 48:671-679.
7. Shannon, M.F., Gamble, J.R., and Vadas, M.A. 1988. Nuclear proteins interacting with the promoter region of the human granulocyte/macrophage colony-stimulating factor gene. *Proc. Natl. Acad. Sci. USA* 85:674-678.
8. Huebner, K., Isobe, M., Croce, C.M., Golde, D.W., Kaufman, S.E., and Gasson, J.C. 1985. The human gene encoding GM-CSF is at 5q21-q32, the chromosome region deleted in the 5q- anomaly. *Science* 230:1282-1285.
9. Nagata, S., Tsuchiya, M., Asano, S., Kaziro, Y., Yamazaki, T., Yamamoto, O., Hirata, Y., Kubota, N., Oheda, M., Nomura, H., and Ono, M. 1986. Molecular cloning and expression of cDNA for human granulocyte colony-stimulating factor. *Nature* 319:415-418.
10. Kawasaki, E.S., Ladner, M.B., Wang, A.M., Van Arsdell, J., Warren, M.K., Coyne, M.Y., Schweickart, V.L., Lee, M-T., Wilson, K.J., Boosman, A., Stanley, E.R., Ralph, P., and Mark, D.F. 1985. Molecular cloning of a complementary DNA encoding human macrophage-specific colony-stimulating factor (CSF-1). *Science* 230:291-296.
11. Yang, Y-C., Ciarletta, A.B., Temple, P.A., Chung, M.P., Kovacic, S., Witek-Giannotti, J.S., Leary, A.C., Kriz, R., Donahue, R.E., Wong, G.G., and Clark, S.C. 1986. Human IL-3 (Multi-CSF): Identification by expression cloning of a novel hematopoietic growth factor related to murine IL-3. *Cell* 47:3-10.

RECEPTOR

12. Nicola, N.A. 1987. Why do hemopoietic growth factor receptors interact with each other? *Immunol. Today* 8:134-140.
13. Gasson, J.C., Kaufman, S.E., Weisbart, R.H., Tomonaga, M., and Golde, D.W. 1986. High-affinity binding of granulocyte-macrophage colony-stimulating factor to normal and leukemic human myeloid cells. *Proc. Natl. Acad. Sci. USA* 83:669-673.
14. Nicola, N.A., and Peterson, L. 1986. Identification of distinct receptors for two hemopoietic growth factors (granulocyte colony-stimulating factor and multipotential colony-stimulating factor) by chemical cross-linking. *J. Biol. Chem.* 261:12384-12389.
15. Sherr, C.J., Rettenmier, C.W., Sacca, R., Roussel, M.F., Look, A.T., and Stanley, E.R. 1985. The c-fms proto-oncogene product is related to the receptor for the mononuclear phagocyte growth factor, CSF-1. *Cell* 41:655-676.
16. Rettenmier, C.W., Sacca, R., Furman, W.L., Roussel, M.F., Holt, J.T., Nienhuis, A.W., Stanley, E.R., and Sherr, C.J. 1986. Expression of the human c-fms proto-oncogene product (colony-stimulating factor-1 receptor) on peripheral blood mononuclear cells and choriocarcinoma cell lines. *J. Clin. Invest.* 77:1740-1746.

PHYSIOLOGY

17. Sieff, C.A., Emerson, S.G., Donahue, R.E., Nathan, D.G., Wang, E.A., Wong, G.G., and Clark, S.C. 1985. Human recombinant granulocyte-macrophage colony-stimulating factor: A multilineage hematopoietin. *Science* 230:1171-1173.
18. Ihle, J.N., and Weinstein, Y. 1986. Immunological regulation of hematopoietic/lymphoid stem cell differentiation by interleukin 3. *Adv. Immunol.* 39:1-50.
19. Welte, K., Platzer, E., Lu, L., Gabrilove, J.L., Levi, E., Mertelsmann, R., and Moore, M.A.S. 1985. Purification and biochemical characterization of human pluripotent hematopoietic colony-stimulating factor. *Proc. Natl. Acad. Sci. USA* 82:1526-1530.
20. Gabrilove, J.L., Welte, K., Harris, P., Platzer, E., Lu, L., Levi, E., Mertelsmann, R., and Moore, M.A.S. 1986. Pluripoietin α : A second human hematopoietic colony-stimulating factor produced by the human bladder carcinoma cell line 5637. *Proc. Natl. Acad. Sci. USA* 83:2478-2482.
21. Reed, S.G., Nathan, C.F., Pihl, D.L., Rodricks, P., Shanebeck, K., Conlon, P.J., and Grabstein, K.H. 1987. Recombinant granulocyte/macrophage colony-stimulating factor activates macrophages to inhibit trypanosoma cruzi and release hydrogen peroxide. Comparison with interferon γ . *J. Exp. Med.* 166:1734-1746.
22. Gasson, J.C., Weisbart, R.H., Kaufman, S.E., Clark, S.C., Hewick, R.M., Wong, G.G., and Golde, D.W. 1984. Purified human granulocyte-macrophage colony-stimulating factor: direct action on neutrophils. *Science* 226:1339-1342.

23. Arnaout, M.A., Wang, E.A., Clark, S.C., and Sieff, C.A. 1986. Human recombinant granulocyte-macrophage colony-stimulating factor increases cell-to-cell adhesion and surface expression of adhesion-promoting surface glycoproteins on mature granulocytes. *J. Clin. Invest.* 78:597-601.
24. Gabrilove, J.L., Welte, K., Harris, P., Platzer, E., Lu, L., Levi, E., Mertelsmann, R., and Moore, M.A.S. 1986. Pluripoietin α : A second human hematopoietic colony-stimulating factor produced by the human bladder carcinoma cell line 5637. *Proc. Natl. Acad. Sci. USA* 83:2478-2482.
25. Lopez, A.F., To, L.B., Yang, Y-C., Gamble, J.T., Shannon, M.F., Burns, G.F., Dyson, P.G., Juttner, C.A., Clark, S., and Vadas, M.A. 1987. Stimulation of proliferation, differentiation, and function of human cells by primate interleukin 3. *Proc. Natl. Acad. Sci. USA* 84:2761-2765.
26. Lopez, A.F., Williamson, D.J., Gamble, J.R., Begley, C.G., Harlan, J.M., Klebanoff, S.J., Waltersdorf, A., Wong, G., Clark, S.C., and Vadas, M.A. 1986. Recombinant human granulocyte-macrophage colony-stimulating factor stimulates *in vitro* mature human neutrophil and eosinophil function, surface receptor expression and survival. *J. Clin. Invest.* 78:1220-1228.

IN VIVO ACTIVITIES

27. Groopman, J.E., Mitsuyasu, R.T., DeLeo, M.J., Oette, D.H., and Golde, D.W. 1987. Effect of recombinant human granulocyte-macrophage colony-stimulating factor on myelopoiesis in the acquired immunodeficiency syndrome. *N. Engl. J. Med.* 317:593-598.
28. Vadhan-Raj, S., Keating, M., LeMaistre, A., Hittelman, W.N., McCredie, K., Trujillo, J.M., Broxmeyer, H.E., Henney, C., and Guterman, J.U. 1987. Effects of recombinant human granulocyte-macrophage colony-stimulating factor in patients with myelodysplastic syndromes. *N. Engl. J. Med.* 317:1545-1552.
29. Lang, R.A., Metcalf, D., Cuthbertson, R.A., Lyons, I., Stanley, E., Kelso, A., Kannourakis, G., Williamson, D.J., Klintworth, G.K., Gonda, T.J., and Dunn, A.R. 1987. Transgenic mice expressing a hemopoietic growth factor gene (GM-CSF) develop accumulations of macrophages, blindness, and a fatal syndrome of tissue damage. *Cell* 51:675-686.
30. Moore, M.A.S., and Warren, D.J. 1987. Synergy of interleukin 1 and granulocyte colony-stimulating factor: *In vivo* stimulation of stem-cell recovery and hematopoietic regeneration following 5-fluorouracil treatment of mice. *Proc. Natl. Acad. Sci. USA* 84:7134-7138.
31. Welte, K., Bonilla, M.A., Gillio, A.P., Boone, T.C., Potter, G.K., Gabrilove, J.L., Moore, M.A.S., O'Reilly, R.J., and Souza, L.M. 1987. Recombinant human granulocyte colony-stimulating factor. Effects on hematopoiesis in normal and cyclophosphamide-treated primates. *J. Exp. Med.* 165:941-948.
32. Koren, S., Klimpel, G.R., and Fleischmann, W.R., Jr. 1986. Treatment of mice with macrophage colony stimulating factor (CSF-1) prevents the *in vivo*

myelosuppression induced by murine alpha, beta, and gamma interferons. J. Biol. Response Modifiers 5:481-489.

Other Cytokines

1. Bonnem, E.M., and Spiegel, R.J. 1984. Interferon- α : Current status and future promise. *J. Biol. Response Modifiers* 3:580-598.
2. Sporn, M.B., Roberts, A.B., Wakefield, L.M., and Assoian, R.K. 1986. Transforming growth factor- β : Biological function and chemical structure. *Science* 233:532-534.
3. Derynck, R., Jarrett, J.A., Chen, E.Y., Eaton, D.H., Bell, J.R., Assoian, R.K., Roberts, A.B., Sporn, M.B., and Goeddel, D.V. 1985. Human transforming growth factor- β complementary DNA sequence and expression in normal and transformed cells. *Nature* 316:701-705.
4. Assoian, R.K., Fleurdeley, B.E., Stevenson, H.C., Miller, P.J., Madtes, D.K., Raines, E.W., Ross, R., and Sporn, M.B. 1987. Expression and secretion of type β transforming growth factor by activated human macrophages. *Proc. Natl. Acad. Sci. USA* 84:6020-6024.
5. Shimokado, K., Raines, E.W., Madtes, D.K., Barrett, T.B., Benditt, E.P., and Ross, R. 1985. A significant part of macrophage-derived growth factor consists of at least two forms of PDGF. *Cell* 43:277-286.
6. Martinet, Y., Bitterman, P.B., Mornex, J-F., Grotendorst, G.R., Martin, G.R., and Crystal, R.G. 1986. Activated human monocytes express the c-sis proto-oncogene and release a mediator showing PDGF-like activity. *Nature* 319:158-160.