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UT Southwestern researchers find protein breakdown contributes to pelvic organ prolapse

DALLAS – May 24, 2011 – A gynecologist and a molecular biologist at UT Southwestern Medical Center have collaborated to show for the first time that pelvic organ prolapse – a condition in which the uterus, bladder or vagina protrude from the body – is caused by a combination of a loss of elasticity and a breakdown of proteins in the vaginal wall.

Pelvic organ prolapse affects many women older than 50 years of age. Besides creating pelvic pressure, prolapse can lead to other pelvic-floor disorders such as urinary and fecal incontinence, and can affect sexual function.

“We found that the protein fibulin-5, which until now simply has been known to be important in generating elastic fibers, actually blocks the enzymes that degrade proteins that support the vaginal wall structure,” said Dr. R. Ann Word, professor of obstetrics and gynecology and a co-senior author of the study in May edition of the *Journal of Clinical Investigation*. “The elastic fibers do play a role, but it’s also the enzymes that degrade the matrix that break down both collagen and elastin over time.”

More than 225,000 inpatient surgical procedures for pelvic organ prolapse are performed each year in the U.S. at an estimated cost of more than \$1 billion. But surgery alone is not always effective in the long run; nearly 30 percent of women report continued problems over a five-year follow-up period because the underlying problem of matrix support has not been corrected. There are no current therapies to prevent the progression of prolapse.

Age and vaginal delivery are the two most common risk factors for prolapse; injury to the vaginal wall may occur during childbirth but prolapse often doesn’t occur until decades later. Obesity and menopause are also contributing factors.

“We still don’t understand why patient A has a terrible delivery, with a large baby, but she never gets prolapse. And then we see patients who are 28 with no children, and they’re already starting to have problems. So we know genetic and environmental factors contribute to this,” Dr. Word said.

Using mice, researchers tested how fibulin-5, a protein that is essential for elastic fiber assembly, regulated the activity of matrix metalloprotease-9 (MMP-9), a group of enzymes that break

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down the matrix of collagen and elastic fibers, leading to a loss of the structural support of the vaginal wall.

Researchers used a fibulin-5 deficient rodent model and a new domain-specific mutant of fibulin-5 to demonstrate that fibulin-5-mediated elastogenesis (development of elastic fibers) is essential to support the pelvic organs. They also showed that prolapse of the vaginal wall requires an increase in MMP-9, but that fibulin-5 inhibits activation of this protease in a tissue-specific manner.

“Matrix assembly of the vaginal wall is a very complicated process,” said Dr. Hiromi Yanagisawa, assistant professor of molecular biology and the study’s other co-senior author. “We need to decode what is necessary in this process, but degrading enzymes are the main therapeutic focus.”

Dr. Word said, “The bottom line is the whole matrix is maintained by a balance between synthesis and degradation. Our goal is to optimize pelvic organ support and target these proteases that degrade the matrix.”

Other UT Southwestern researchers involved in the study were Dr. Madhusudhan Budatha, postdoctoral researcher of molecular biology and lead author; Dr. Quin Zheng, former postdoctoral researcher of molecular biology; Dr. Shayzreen Roshanravan and Dr. Cecilia Weislander, former postdoctoral researchers of obstetrics and gynecology; and Shelby Chapman, senior research associate of molecular biology. Scientists from UT Health Center in Tyler and McGill University in Montreal also participated.

The study was supported by grants from the National Institutes of Health, the American Heart Association and the Welch Foundation.

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