

Regulatory Mechanism of the RNAi Pathway

Ying Liu, Ph.D.

The University of Texas Southwestern Medical Center at Dallas, 2011

Graduate School of Biomedical Sciences

Supervising Professor: Qinghua Liu, Ph.D.

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RNA interference (RNAi) is post-transcriptional gene silencing initiated by Dicer, a RNase III that processes double-stranded RNA (dsRNA) precursors into small interfering RNA (siRNA). In *Drosophila*, Dicer2 and R2D2 coordinately recruit duplex siRNA to the effector RNA-induced silencing complex (RISC), wherein single-stranded siRNA guides the endoribonuclease Argonaute (Ago) to catalyze sequence-specific cleavage of complementary mRNA. It remains unclear as to what constitutes holo-RISC, how is RISC assembled and how is RISC regulated. Here we took a candidate approach to reconstitute for the first time the long double-stranded RNA- and duplex siRNA-initiated RISC activities with the use of recombinant *Drosophila* Dicer-2, R2D2, and Ago2 proteins. We further employed this core reconstitution system to purify a RNAi regulator that we named C3PO (component 3 promoter of RISC), a complex of Translin and Trax. C3PO is a novel Mg²⁺-dependent endoribonuclease that promotes RISC activation by removing the siRNA passenger strand cleavage products. Similar as *Drosophila* C3PO, human C3PO also degrades passenger strand fragments and facilitates RISC activation.

RISC is a multiple-turnover enzyme, wherein single-stranded (ss)-siRNA guides Ago2 to catalyze sequence-specific cleavage of the target mRNA at the effector step. We employed human minimal RISC reconstitution system to purify antoantigen La as a novel activator of the RISC effector step. Biochemical studies indicated that La promotes the multiple-turnover of RISC catalysis by facilitating the release of RISC cleaved products. Moreover, we demonstrated that La is required for efficient RNAi, antiviral defense, and transposon silencing in mammalian and *Drosophila* cells.

Taken together, our findings of C3PO and La reveal a general concept that regulatory factors are required to remove Ago2-cleaved products to assemble or restore active RISC. The robust reconstitution system establishes a powerful platform for in-depth studies of the assembly, function, and regulation of RISC. Similar to the discovery of C3PO and La, it can be used to identify novel regulators and study post-translational regulations of RNAi, therefore, connecting RNAi to other cellular signaling pathways. As such, these biomedical studies could have a major and lasting impact on the biological understanding and therapeutic application of RNAi.